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(54) Title: PEPTIDE NUCLEIC ACIDS

(57) Abstract

A novel class of compounds, known as peptide nucleic acids, bind complementary ssDNA and RNA strands more strongly than a corresponding DNA. The peptide nucleic acids generally comprise ligands such as naturally occurring DNA bases attached to a peptide backbone through a suitable linker.

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PEPTIDE NUCLEIC ACIDS

RELATED APPLICATIONS

This application is a continuation-in-part of the
5 following Danish Patent Applications: No. 986/91, filed May
24, 1991, No. 987/91, filed May 24, 1991, and No. 510/92,
filed April 15, 1992. The entire disclosure of each
application is incorporated herein by reference.

10 FIELD OF THE INVENTION

This invention is directed to compounds that are not
polynucleotides yet which bind to complementary DNA and RNA
strands more strongly than the corresponding DNA. In particular,
the invention concerns compounds wherein naturally-occurring
15 nucleobases or other nucleobase-binding moieties are
covalently bound to a polyamide backbone.

BACKGROUND OF THE INVENTION

Oligodeoxyribonucleotides as long as 100 base pairs
20 (bp) are routinely synthesized by solid phase methods using
commercially available, fully automatic synthesis machines.
The chemical synthesis of oligoribonucleotides, however, is
far less routine. Oligoribonucleotides also are much less
stable than oligodeoxyribonucleotides, a fact which has
25 contributed to the more prevalent use of oligodeoxyribonucleo-
tides in medical and biological research directed to, for
example, gene therapy or the regulation of transcription or
translation.

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The function of a gene starts by transcription of its information to a messenger RNA (mRNA) which, by interaction with the ribosomal complex, directs the synthesis of a protein coded for by its sequence. The synthetic process is known as 5 translation. Translation requires the presence of various co-factors and building blocks, the amino acids, and their transfer RNAs (tRNA), all of which are present in normal cells.

Transcription initiation requires specific recognition 10 of a promoter DNA sequence by the RNA-synthesizing enzyme, RNA polymerase. In many cases in prokaryotic cells, and probably in all cases in eukaryotic cells, this recognition is preceded by sequence-specific binding of a protein transcription factor to the promoter. Other proteins which bind to the promoter, 15 but whose binding prohibits action of RNA polymerase, are known as repressors. Thus, gene activation typically is regulated positively by transcription factors and negatively by repressors.

Most conventional drugs function by interaction with 20 and modulation of one or more targeted endogenous proteins, e.g., enzymes. Such drugs, however, typically are not specific for targeted proteins but interact with other proteins as well. Thus, a relatively large dose of drug must be used to effectively modulate a targeted protein. Typical 25 daily doses of drugs are from 10^{-5} - 10^{-1} millimoles per kilogram of body weight or 10^{-3} - 10 millimoles for a 100 kilogram person. If this modulation instead could be effected by interaction with and inactivation of mRNA, a dramatic reduction in the necessary amount of drug necessary could 30 likely be achieved, along with a corresponding reduction in side effects. Further reductions could be effected if such interaction could be rendered site-specific. Given that a functioning gene continually produces mRNA, it would thus be even more advantageous if gene transcription could be arrested 35 in its entirety.

Oligodeoxynucleotides offer such opportunities. For example, synthetic oligodeoxynucleotides could be used as

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antisense probes to block and eventually lead to the breakdown of mRNA. Thus, synthetic DNA could suppress translation *in vivo*. It also may be possible to modulate the genome of an animal by, for example, triple helix formation using 5 oligonucleotides or other DNA recognizing agents. However, there are a number of drawbacks associated with triple helix formation. For example, it can only be used for homopurine sequences and it requires unphysiologically high ionic strength and low pH.

10 Furthermore, unmodified oligonucleotides are unpractical both in the antisense approach and in the triple helix approach because they have short *in vivo* half-lives, they are difficult to prepare in more than milligram quantities and, thus, are prohibitively costly, and they are 15 poor cell membrane penetrators.

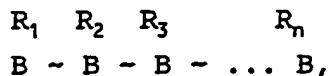
These problems have resulted in an extensive search for improvements and alternatives. For example, the problems arising in connection with double-stranded DNA (dsDNA) recognition through triple helix formation have been 20 diminished by a clever "switch back" chemical linking whereby a sequence of polypurine on one strand is recognized, and by "switching back", a homopurine sequence on the other strand can be recognized. See, e.g., McCurdy, Moulds, and Froehler, *Nucleosides*, in press. Also, good helix formation has been 25 obtained by using artificial bases, thereby improving binding conditions with regard to ionic strength and pH.

In order to improve half life as well as membrane penetration, a large number of variations in polynucleotide backbones has been undertaken, although so far not with 30 desired results. These variations include the use of methylphosphonates, monothiophosphates, dithiophosphates, phosphoramidates, phosphate esters, bridged phosphoroamidates, bridged phosphorothioates, bridged methylenephosphonates, dephospho internucleotide analogs with siloxane bridges, 35 carbonate bridges, carboxymethyl ester bridges, acetamide bridges, carbamate bridges, thioether, sulfoxy, sulfono

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bridges, various "plastic" DNAs, α -anomeric bridges, and borane derivatives.

International patent application WO 86/05518 broadly claims a polymeric composition effective to bind to a single-stranded polynucleotide containing a target sequence of bases. The composition is said to comprise non-homopolymeric, substantially stereoregular polymer molecules of the form:

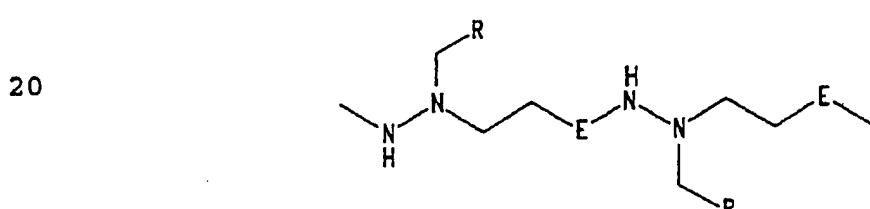
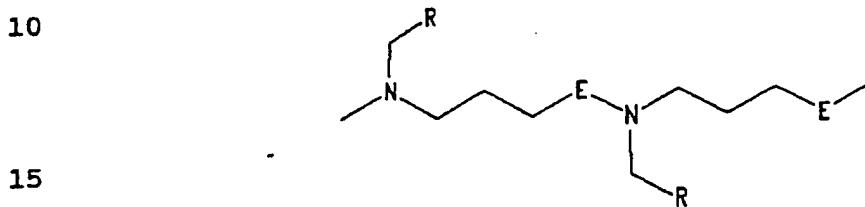
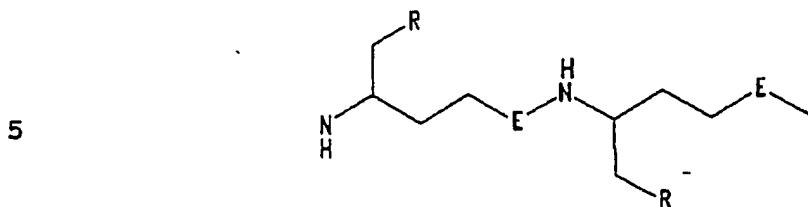


10 where:

- (a) R_1-R_n are recognition moieties selected from purine, purine-like, pyrimidine, and pyrimidine like heterocycles effective to bind by Watson/Crick pairing to corresponding, in-sequence bases in the target sequence;
- (b) n is such that the total number of Watson/Crick hydrogen bonds formed between a polymer molecule and target sequence is at least about 15;
- (c) $B - B$ are backbone moieties joined predominantly by chemically stable, substantially uncharged, predominantly achiral linkages;
- (d) the backbone moiety length ranges from 5 to 7 atoms if the backbone moieties have a cyclic structure, and ranges from 4 to 6 atoms if the backbone moieties have an acyclic structure; and
- (e) the backbone moieties support the recognition moieties at position which allow Watson/Crick base pairing between the recognition moieties and the corresponding, in-sequence bases of the target sequence.

According to WO 86/05518, the recognition moieties are various natural nucleobases and nucleobase-analogs and the backbone moieties are either cyclic backbone moieties comprising furan or morpholine rings or acyclic backbone moieties of the following forms:

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where E is -CO- or -SO₂-.

The specification of the application provides general descriptions for the synthesis 30 of subunits, for backbone coupling reactions, and for polymer assembly strategies. However, the specification provides no example wherein a claimed compound or structure is actually prepared. Although WO 86/05518 indicates that the claimed polymer compositions can bind target sequences and, as a 35 result, have possible diagnostic and therapeutic applications, the application contains no data relating to the binding affinity of a claimed polymer.

International patent application WO 86/05519 claims diagnostic reagents and systems that comprise polymers 40 described in WO 86/05518, but attached to a solid support. WO 86/05519 also provides no examples concerning actually preparation of a claimed diagnostic reagent, much less data showing the diagnostic efficiency of such a reagent.

International patent application WO 89/12060 claims 45 various building blocks for synthesizing oligonucleotide analogs, as well as oligonucleotide analogs formed by joining

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such building blocks in a defined sequence. The building blocks may be either "rigid" (containing a ring) or "flexible" (lacking a ring). In both cases the building blocks contain a hydroxy group and a mercapto group, through which the 5 building blocks are said to join to form oligonucleotide analogs. The linking moiety in the oligonucleotide analogs is selected from the group consisting of sulfide (-S-), sulfoxide (-SO-), and sulfone (-SO₂-). WO 89/12060 provides a general description concerning synthesis of the building 10 blocks and coupling reactions for the synthesis of oligonucleotide analogs, along with experimental examples describing the preparation of building blocks. However, the application provides no examples directed to the preparation of a claimed oligonucleotide analog and no data confirming the 15 specific binding of an oligonucleotide analog to a target oligonucleotide.

Furthermore, oligonucleotides or their derivatives have been linked to intercalators in order to improve binding, to polylysine or other basic groups in order to improve binding 20 both to double-stranded and single-stranded DNA, and to peptides in order to improve membrane penetration. However, such linking has not resulted in satisfactory binding for either double-stranded or single-stranded DNA. Other problems which resulted from, for example, methylphosphonates and 25 monothiophosphates were the occurrence of chirality, insufficient synthetic yield or difficulties in performing solid phase assisted syntheses.

In most cases only a few of these modifications could be used. Even then, only short sequences -- often only dimers 30 -- or monomers could be generated. Furthermore, the oligomers actually produced have rarely been shown to bind to DNA or RNA or have not been examined biologically.

The great majority of these backbone modifications led to decreased stability for hybrids formed between the modified 35 oligonucleotide and its complementary native oligonucleotide, as assayed by measuring T_m values. Consequently, it is generally understood in the art that backbone modifications

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destabilize such hybrids, i.e., result in lower T_m values, and should be kept to a minimum.

OBJECTS OF THE INVENTION

5 It is one object of the present invention to provide compounds that bind ssDNA and RNA strands to form stable hybrids therewith.

It is a further object of the invention to provide compounds that bind ssDNA and RNA strands more strongly than the 10 corresponding DNA.

It is another object to provide compounds wherein naturally-occurring nucleobases or other nucleobase-binding moieties are covalently bound to a peptide backbone.

It is yet another object to provide compounds other 15 than RNA that can bind one strand of a double-stranded polynucleotide, thereby displacing the other strand.

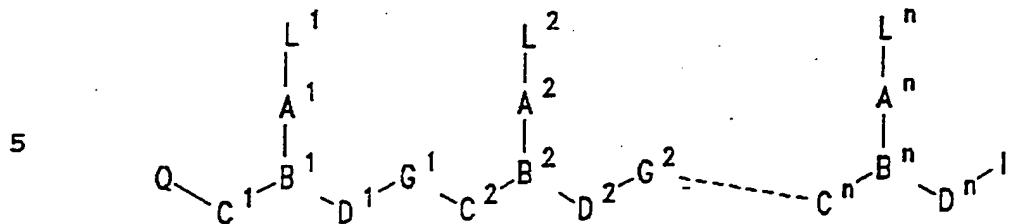
It is still another object to provide therapeutic and prophylactic methods that employ such compounds.

20 SUMMARY OF THE INVENTION

The present invention provides a novel class of compounds, known as peptide nucleic acids (PNAs), that bind complementary ssDNA and RNA strands more strongly than a corresponding DNA. The compounds of the invention generally 25 comprise ligands linked to a peptide backbone via an aza nitrogen. Representative ligands include either the four main naturally occurring DNA bases (i.e., thymine, cytosine, adenine or guanine) or other naturally occurring nucleobases (e.g., inosine, uracil, 5-methylcytosine or thiouracil) or 30 artificial bases (e.g., bromothymine, azaadenines or azaguanines, etc.) attached to a peptide backbone through a suitable linker.

In certain preferred embodiments, the peptide nucleic acids of the invention have the general formula (I):

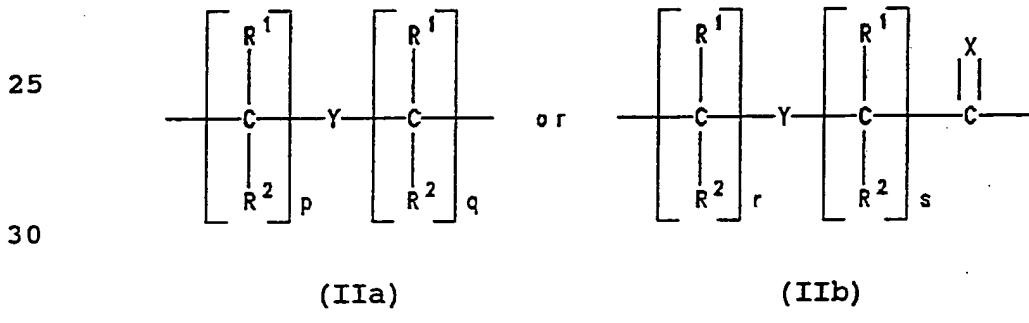
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wherein:

n is at least 2,
each of L^1-L^n is independently selected from the group
consisting of hydrogen, hydroxy, (C_1-C_4) alkanoyl, naturally
15 occurring nucleobases, non-naturally occurring nucleobases,
aromatic moieties, DNA intercalators, nucleobase-binding
groups, heterocyclic moieties, and reporter ligands, at least
one of L^1-L^n being a naturally occurring nucleobase, a non-
naturally occurring nucleobase, a DNA intercalator, or a
20 nucleobase-binding group;

each of A^1-A^n is a single bond, a methylene group or a
group of formula (IIa) or (IIb):



where:

35 X is O, S, Se, NR^3 , CH_2 or $C(CH_3)_2$;
Y is a single bond, O, S or NR^4 ;
each of p and q is zero or an integer from 1 to
5, the sum p+q being not more than 10;
each of r and s is zero or an integer from 1 to
40 5, the sum r+s being not more than 10;
each R^1 and R^2 is independently selected from the
group consisting of hydrogen, (C_1-C_4) alkyl which may be
hydroxy- or alkoxy- or alkylthio-substituted, hydroxy,
alkoxy, alkylthio, amino and halogen; and

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each R³ and R⁴ is independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl, hydroxy- or alkoxy- or alkylthio-substituted (C₁-C₄)alkyl, hydroxy, alkoxy, alkylthio and amino;

5 each of B¹-Bⁿ is N or R³N⁺, where R³ is as defined above; each of C¹-Cⁿ is CR⁶R⁷, CHR⁶CHR⁷ or CR⁶R⁷CH₂, where R⁶ is hydrogen and R⁷ is selected from the group consisting of the side chains of naturally occurring alpha amino acids, or R⁶ and R⁷ are independently selected from the group consisting of 10 hydrogen, (C₂-C₆)alkyl, aryl, aralkyl, heteroaryl, hydroxy, (C₁-C₆)alkoxy, (C₁-C₆)alkylthio, NR³R⁴ and SR⁵, where R³ and R⁴ are as defined above, and R⁵ is hydrogen, (C₁-C₆)alkyl, hydroxy-, alkoxy-, or alkylthio- substituted (C₁-C₆)alkyl, or R⁶ and R⁷ taken together complete an alicyclic or heterocyclic 15 system;

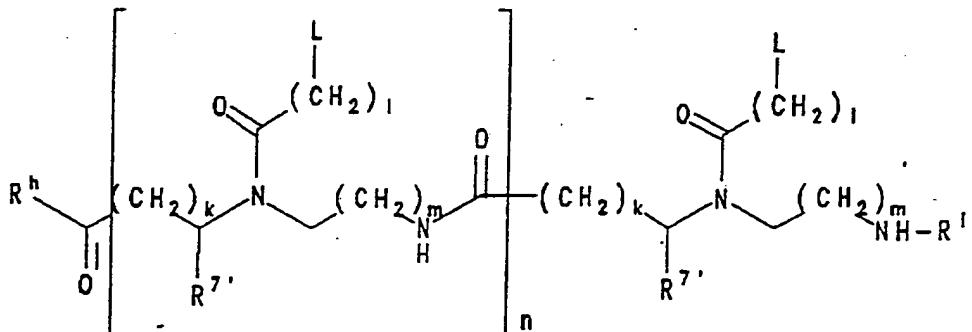
each of D¹-Dⁿ is CR⁶R⁷, CH₂CR⁶R⁷ or CHR⁶CHR⁷, where R⁶ and R⁷ are as defined above;

each of G¹-Gⁿ⁻¹ is -NR³CO-, -NR³CS-, -NR³SO- or -NR³SO₂-, Y in either orientation, where R³ is as defined above; 20 Q is -CO₂H, -CONR'R'', -SO₃H or -SO₂NR'R'' or an activated derivative of -CO₂H or -SO₃H; and I is -NHR'''R'''' or -NR'''C(O)R''''', where R', R'', R''' and R''''' are independently selected from the group consisting of hydrogen, alkyl, amino protecting groups, 25 reporter ligands, intercalators, chelators, peptides, proteins, carbohydrates, lipids, steroids, oligonucleotides and soluble and non-soluble polymers.

The peptide nucleic acids of the invention differ from those disclosed in WO 86/05518 in that their recognition 30 moieties are attached to an aza nitrogen atom in the backbone, rather than to an amide nitrogen atom, a hydrazine moiety or a carbon atom in the backbone.

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Preferred peptide nucleic acids have general formula (III):



(III)

wherein:

5 each L is independently selected from the group consisting of hydrogen, phenyl, heterocyclic moieties, naturally occurring nucleobases, and non-naturally occurring nucleobases;

10 each R' is independently selected from the group consisting of hydrogen and the side chains of naturally occurring alpha amino acids;

n is an integer from 1 to 60;

each of k, l and m is independently zero or an integer from 1 to 5;

15 R^h is OH, NH₂ or -NHLysNH₂; and

R' is H or COCH₃.

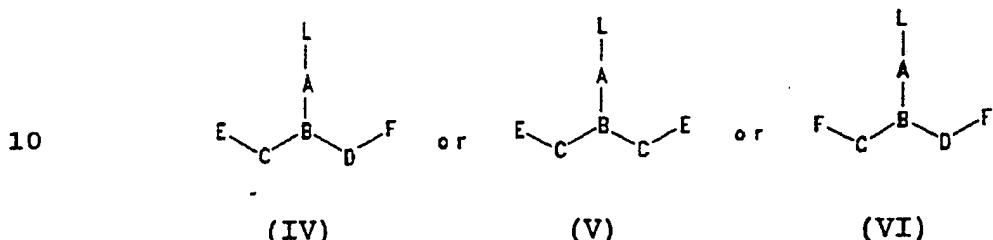
Particularly preferred are compounds having formula (III) wherein each L is independently selected from the group consisting of the nucleobases thymine (T), adenine (A), 20 cytosine (C), guanine (G) and uracil (U), k and m are zero or 1, and n is an integer from 1 to 30, in particular from 4 to 20. An example of such a compound is provided in Figure 1, which shows the structural similarity between such compounds and single-stranded DNA.

25 The peptide nucleic acids of the invention are synthesized by adaptation of standard peptide synthesis procedures, either in solution or on a solid phase. The synthons used are specially designed monomer amino acids or their activated derivatives, protected by standard protecting

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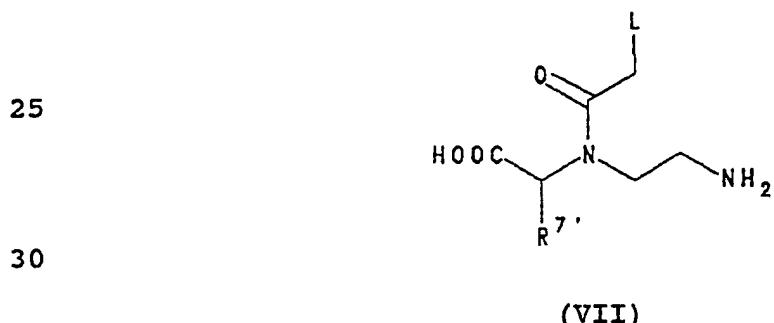
groups. The oligonucleotide analogs also can be synthesized by using the corresponding diacids and diamines.

Thus, the novel monomer synthons according to the invention are selected from the group consisting of amino acids, diacids and diamines having general formulae:



15 wherein L, A, B, C and D are as defined above, except that any amino groups therein may be protected by amino protecting groups; E is COOH, CSOH, SOOH, SO₂OH or an activated derivative thereof; and F is NHR³ or NPgR³, where R³ is as defined above and Pg is an amino protecting group.

20 Preferred monomer synthons according to the invention
are amino acids having formula (VII):



or amino-protected and/or acid terminal activated derivatives thereof, wherein L is selected from the group consisting of
35 hydrogen, phenyl, heterocyclic moieties, naturally occurring nucleobases, non-naturally occurring nucleobases, and protected derivatives thereof; and R⁷ is independently selected from the group consisting of hydrogen and the side chains of naturally occurring alpha amino acids. Especially
40 preferred are such synthons having formula (VII) wherein R⁷ is hydrogen and L is selected from the group consisting of the nucleobases thymine (T), adenine (A), cytosine (C), guanine (G) and uracil (U) and protected derivatives thereof.

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Unexpectedly, these compounds also are able to recognize duplex DNA by displacing one strand, thereby presumably generating a double helix with the other one. Such recognition can take place to dsDNA sequences 5-60 base pairs 5 long. Sequences between 10 and 20 bases are of interest since this is the range within which unique DNA sequences of prokaryotes and eukaryotes are found. Reagents which recognize 17-18 bases are of particular interest since this is the length of unique sequences in the human genome. The 10 compounds of the invention also should be able to form triple helices with dsDNA.

Whereas the improved binding of the compounds of the invention should render them efficient as antisense agents, it is expected that an extended range of related reagents may 15 cause strand displacement, now that this surprising and unexpected new behavior of dsDNA has been discovered.

Thus, in one aspect, the present invention provides methods for inhibiting the expression of particular genes in the cells of an organism, comprising administering to said 20 organism a reagent as defined above which binds specifically to sequences of said genes.

Further, the invention provides methods for inhibiting transcription and/or replication of particular genes or for inducing degradation of particular regions of double stranded 25 DNA in cells of an organism by administering to said organism a reagent as defined above.

Still further, the invention provides methods for killing cells or virus by contacting said cells or virus with a reagent as defined above which binds specifically to 30 sequences of the genome of said cells or virus.

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BRIEF DESCRIPTION OF THE DRAWINGS

The numerous objects and advantages of the present invention may be better understood by those skilled in the art by reference to the accompanying figures, in which:

- 5 Figure 1 shows a naturally occurring deoxyribooligonucleotide (A) and a peptide nucleic acid (PNA) of the invention (B).

Figure 2 provides examples of naturally occurring and non-naturally occurring nucleobases for DNA recognition and 10 reporter groups.

Figure 3 provides a schematic illustration of (a) photocleavage by Acr¹-(Taeg)₁₀-Lys-NH₂ (Acr-T10-LysNH₂); (b) photofootprint by the diazo-linked acridine of Acr¹-(Taeg)₁₀-Lys-NH₂ and preferred KMnO₄-cleavage; and (c) S₁-nuclease 15 enhanced cleavage and (d) micrococcus nuclease cleavage of Acr¹-(Taeg)₁₀-Lys-NH₂ binding site.

Figure 4 provides examples of PNA monomer synthons of the invention.

Figure 5 shows the Acr¹ ligand and a PNA, Acr¹-(Taeg)₁₀-20 Lys-NH₂.

Figure 6 provides a general scheme for the preparation of monomer synthons.

Figure 7 provides a general scheme for the preparation of the Acr¹ ligand.

- 25 Figure 8 provides a general scheme for solid-phase PNA synthesis illustrating the preparation of linear unprotected PNA amides.

Figure 9 shows analytical HPLC chromatograms of: (A) crude H-[Taeg]₁₅-NH₂ after HF-cleavage (before lyophilization); 30 (B) crude Acr¹-[Taeg]₁₅-NH₂ after HF-cleavage (before lyophilization); and (C) purified Acr¹-[Taeg]₁₅-NH₂. Buffer A, 5% CH₃CN/95% H₂O/0.0445% TFA; buffer B, 60% CH₃CN/40% H₂O/0.0390% TFA; linear gradient, 0-100% of B in 30 min; flow rate, 1.2 ml/min; column, Vydac C₁₈ (5 μm, 0.46 x 25 cm).

- 35 Figure 10 shows analytical HPLC chromatograms of: (A) purified H-[Taeg]₁₀-Lys-NH₂ and (B) purified H-[Taeg]₅-Caeg-[Taeg]₄-Lys-NH₂ employing the same conditions as in Figure 9.

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Figures 11a and 11b show binding of AcrT10-Lys to dA₁₀. 5'-³²P-labeled oligonucleotide (1) (5'-GATCCA₁₀G) was incubated in the absence or presence of Acr-T10-LysNH₂ and in the absence or presence of oligonucleotide (2) (5'-GATCCT₁₀G) and 5 the samples were analyzed by polyacrylamide gel electrophoresis (PAGE) and autoradiography under "native conditions" (Figure 11a) or under "denaturing conditions" (Figure 11b).

Figures 12a-c show chemical, photochemical and 10 enzymatic probing of dsDNA-Acr-T10-LysNH₂ complex. Complexes between Acr-T10-LysNH₂ and a ³²P-endlabeled DNA fragment containing a dA₁₀/dT₁₀ target sequence were probed by affinity photocleavage (Figure 12a, lanes 1-3; Figure 12b, lanes 1-3), photofootprinting (Figure 12a, lanes 5-6), potassium 15 permanganate probing (Figure 12b, lanes 4-6) or probing by staphylococcus nuclease (Figure 12b, lanes 8-10) or by nuclease S₁ (Figure 12c). Either the A-strand (Figure 12a) or the T-strand (Figures 12b,c) was probed.

Figure 13 provides a procedure for the synthesis of 20 protected PNA synthons.

Figure 14 provides a procedure for the synthesis of a protected adenine monomer synthon.

Figure 15 provides a procedure for the synthesis of a protected guanine monomer synthon.

25 Figure 16 provides examples of PNA backbone alterations.

Figure 17 provides a procedure for synthesis of thymine monomer synthons with side chains corresponding to the normal amino acids.

30 Figures 18a and 18b provide procedures for synthesis of an aminopropyl analogue and a propionyl analogue, respectively, of a thymine monomer synthon.

Figure 19 provides a procedure for synthesis of an aminoethyl-β-alanine analogue of thymine monomer synthon.

35 Figure 20 shows a PAGE autoradiograph demonstrating that PNAs-T₁₀, -T₉C and -T₈C₂ bind to double stranded DNA with high sequence specificity.

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Figure 21 shows a graph based on densitometric scanning of PAGE autoradiographs demonstrating the kinetics of the binding of PNA-T₁₀ to a double stranded target.

Figure 22 shows a graph based on densitometric scanning 5 of PAGE autoradiographs demonstrating the thermal stabilities of PNAs of varying lengths bound to an A₁₀/T₁₀ double stranded DNA target.

Figure 23 shows an electrophoretic gel staining demonstrating that restriction enzyme activity towards DNA is 10 inhibited when PNA is bound proximal to the restriction enzyme recognition site.

Figure 24 shows a PAGE autoradiograph demonstrating that ¹²⁵I-labeled PNA-T₁₀ binds to a complementary dA₁₀ oligonucleotide.

15 Figure 25 shows a peptide nucleic acid according to the invention.

Figure 26 shows the direction of synthesis for a peptide nucleic acid according to the invention.

Figure 27 provides a test for the tosyl group as a 20 nitrogen protecting group in the synthesis of peptide nucleic acids.

DETAILED DESCRIPTION OF THE INVENTION

In the oligonucleotide analogs and monomer synthons 25 according to the invention, ligand L is primarily a naturally occurring nucleobase attached at the position found in nature, i.e., position 9 for adenine or guanine, and position 1 for thymine or cytosine. Alternatively, L may be a non-naturally occurring nucleobase (nucleobase analog), another base-binding 30 moiety, an aromatic moiety, (C₁-C₄)alkanoyl, hydroxy or even hydrogen. Some typical nucleobase ligands and illustrative synthetic ligands are shown in Figure 2. Furthermore, L can be a DNA intercalator, a reporter ligand such as, for example, a fluorophor, radio label, spin label, hapten, or a protein- 35 recognizing ligand such as biotin.

In monomer synthons, L may be blocked with protecting groups. This is illustrated in Figure 4, where Pg¹ is an

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acid, a base or a hydrogenolytically or photochemically cleavable protecting group such as, for example, t-butoxycarbonyl (Boc), fluorenylmethyloxycarbonyl (Fmoc) or 2-nitrobenzyl (2Nb).

5 Linker A can be a wide variety of groups such as $-\text{CR}^1\text{R}^2\text{CO}-$, $-\text{CR}^1\text{R}^2\text{CS}-$, $-\text{CR}^1\text{R}^2\text{CSe}-$, $-\text{CR}^1\text{R}^2\text{CNHR}^3-$, $-\text{CR}^1\text{R}^2\text{C}=\text{CH}_2-$ and $-\text{CR}^1\text{R}^2\text{C}=\text{C}(\text{CH}_3)_2-$, where R^1 , R^2 and R^3 are as defined above. Preferably, A is methylenecarbonyl ($-\text{CH}_2\text{CO}-$). Also, A can be a longer chain moiety such as propanoyl, butanoyl or 10 pentanoyl, or corresponding derivative, wherein O is replaced by another value of X or the chain is substituted with R^1R^2 or is heterogenous, containing Y. Further, A can be a (C_2-C_6) alkylene chain, a (C_2-C_6) alkylene chain substituted with R^1R^2 or can be heterogenous, containing Y. In certain cases, A can 15 just be a single bond.

In the preferred form of the invention, B is a nitrogen atom, thereby presenting the possibility of an achiral backbone. B can also be R^3N^+ , where R^3 is as defined above.

In the preferred form of the invention, C is $-\text{CR}^6\text{R}^7-$, 20 but can also be a two carbon unit, i.e. $-\text{CHR}^6\text{CHR}^7-$ or $-\text{CR}^6\text{R}^7\text{CH}_2-$, where R^6 and R^7 are as defined above. R^6 and R^7 also can be a heteroaryl group such as, for example, pyrrolyl, furyl, thienyl, imidazolyl, pyridyl, pyrimidinyl, indolyl, or can be taken together to complete an alicyclic system such as, 25 for example, 1,2-cyclobutenediyl, 1,2-cyclopentanediyI or 1,2-cyclohexanediyI.

In the preferred form of the invention, E in the monomer synthon is COOH or an activated derivative thereof, and G in the oligomer is $-\text{CONR}^3-$. As defined above, E may 30 also be CSOH, SOOH, SO_2OH or an activated derivative thereof, whereby G in the oligomer becomes $-\text{CSNR}^3-$, $-\text{SONR}^3-$ and $-\text{SO}_2\text{NR}^3-$, respectively. The activation may, for example, be achieved using an acid anhydride or an active ester derivative, wherein hydrogen in the groups represented by E is replaced by a 35 leaving group suited for generating the growing backbone.

The amino acids which form the backbone may be identical or different. We have found that those based on 2-

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aminoethylglycine are especially well suited to the purpose of the invention.

In some cases it may be of interest to attach ligands at either terminus (Q, I) to modulate the binding characteristics of the PNAs. Representative ligands include DNA intercalators which will improve dsDNA binding or, basic groups, such as lysine or polylysine, which will strengthen the binding of PNA due to electrostatic interaction. To decrease negatively charged groups such as carboxy and sulfo groups could be used. The design of the synthons further allows such other moieties to be located on non-terminal positions.

In a further aspect of the invention, the PNA oligomers are conjugated to low molecular effector ligands such as ligands having nuclease activity or alkylating activity or reporter ligands (fluorescent, spin labels, radioactive, protein recognition ligands, for example, biotin or haptens). In a further aspect of the invention, the PNAs are conjugated to peptides or proteins, where the peptides have signaling activity and the proteins are, for example, enzymes, transcription factors or antibodies. Also, the PNAs can be attached to water-soluble or water-insoluble polymers. In another aspect of the invention, the PNAs are conjugated to oligonucleotides or carbohydrates. When warranted, a PNA oligomer can be synthesized onto some moiety (e.g., a peptide chain, reporter, intercalator or other type of ligand-containing group) attached to a solid support.

Such conjugates can be used for gene modulation (e.g., gene targeted drugs), for diagnostics, for biotechnology, and for scientific purposes.

As a further aspect of the invention, PNAs can be used to target RNA and ssDNA to produce both antisense-type gene regulating moieties and hybridization probes for the identification and purification of nucleic acids. Furthermore, the PNAs can be modified in such a way that they can form triple helices with dsDNA. Reagents that bind sequence-specifically to dsDNA have applications as gene

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targeted drugs. These are foreseen as extremely useful drugs for treating diseases like cancer, AIDS and other virus infections, and may also prove effective for treatment of some genetic diseases. Furthermore, these reagents may be used for 5 research and in diagnostics for detection and isolation of specific nucleic acids.

The triple helix principle is believed to be the only known principle in the art for sequence-specific recognition of dsDNA. However, triple helix formation is largely limited 10 to recognition of homopurine-homopyrimidine sequences. Strand displacement is superior to triple helix recognition in that it allows for recognition of any sequence by use of the four natural bases. Also, in strand displacement recognition readily occurs at physiological conditions, that is, neutral 15 pH, ambient (20-40 C) temperature and medium (100-150 mM) ionic strength.

Gene targeted drugs are designed with a nucleobase sequence (containing 10-20 units) complementary to the regulatory region (the promoter) of the target gene. 20 Therefore, upon administration of the drug, it binds to the promoter and block access thereto by RNA polymerase. Consequently, no mRNA, and thus no gene product (protein), is produced. If the target is within a vital gene for a virus, no viable virus particles will be produced. Alternatively, 25 the target could be downstream from the promoter, causing the RNA polymerase to terminate at this position, thus forming a truncated mRNA/protein which is nonfunctional.

Sequence-specific recognition of ssDNA by base complementary hybridization can likewise be exploited to target 30 specific genes and viruses. In this case, the target sequence is contained in the mRNA such that binding of the drug to the target hinders the action of ribosomes and, consequently, translation of the mRNA into protein. The peptide nucleic acids of the invention are superior to prior reagents in that 35 they have significantly higher affinity for complementary ssDNA. Also, they possess no charge and water soluble, which should facilitate cellular uptake, and they contain amides of

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non-biological amino acids, which should make them biostable and resistant to enzymatic degradation by, for example, proteases.

Certain biochemical/biological properties of PNA oligomers are illustrated by the following experiments.

1. Sequence discrimination at the dsDNA level (Example 63, Figure 20).

Using the S₁-nuclease probing technique, the discrimination of binding of the T₁₀, T₅CT₄(T₉C) & T₂CT₂CT₄(T₈C₂) PNA to the recognition sequences A₁₀, A₅GA₄ (A₉G) & A₂GA₂GA₄ (A₈G₂) cloned into the BamHI, SalI or PstI site of the plasmid pUC19 was analyzed. The results (Figure 20) show that the three PNAs bind to their respective recognition sequences with the following relative efficiencies: PNA - T10: A₁₀ > A₉G >> A₈G₂, PNA - T₉C: A₉G > A₁₀ ~ A₈G₂, PNA - T₈C₂: A₈G₂ ≥ A₉G >> A₁₀. Thus at 37°C one mismatch out of ten gives reduced efficiency (5-10 times estimated) whereas two mismatches are not accepted.

2. Kinetics of PNA-T₁₀ - dsDNA strand displacement complex formation (Example 66, Figure 21).

Complex formation was probed by S₁-nuclease at various times following mixing of PNA and ³²P-endlabeled dsDNA fragment (Figure 21).

3. Stability of PNA-dsDNA complex (Example 67, Figure 22)

Complexes between PNA-T_n and ³²P-dsDNA (A₁₀/T₁₀) target were formed (60 min, 37°C). The complexes were then incubated at the desired temperature in the presence of excess oligo-dA₁₀ for 10 min, cooled to RT and probed with KMnO₄. The results (Figure 22) show that the thermal stability of the PNA-dsDNA complexes mirror that of the PNA oligonucleotide complexes in terms of "Tm".

4. Inhibition of restriction enzyme cleavage by PNA (Example 65, Figure 23)

The plasmid construct, pT10, contains a dA₁₀/dT₁₀ tract cloned into the BamHI site in pUC19. Thus, cleavage of pT10 with BamHI and Pvull results in two small DNA fragments of 211 and 111 bp, respectively. In the presence of PNA-T₁₀, a 336 bp fragment is obtained corresponding to cleavage only by

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PvuII (Figure 23). Thus cleavage by BamHI is inhibited by PNA bound proximal to the restriction enzyme site. The results also show that the PNA-dsDNA complex can be formed in 100% yield. Similar results were obtained using the pT8C2 plasmid 5 and PNA-T8C2.

5. Binding of 125 I-labeled PNA to oligonucleotides (Example 63, Figure 24)

A Tyr-PNA-T₁₀-Lys-NH₂ was labeled with 125 I using Na 125 I and chloramine-T and purified by HPLC. The 125 I-PNA-T₁₀ was 10 shown to bind to oligo-dA₁₀ by PAGE and autoradiography (Figure 24). The binding could be competed by excess denatured calf thymus DNA.

The sequence-specific recognition of dsDNA is illustrated by the binding of a PNA, consisting of 10 thymine substituted 15 2-aminoethylglycyl units, which C-terminates in a lysine amide and N-terminates in a complex 9-aminoacridine ligand (9-Acr¹-(Taeg)₁₀-Lys-NH₂, Figure 11a, 11b) to a dA₁₀/dT₁₀ target sequence. The target is contained in a 248 bp 32 P-end-labelled DNA-fragment.

20 Strand displacement was ascertained by the following type of experiments:

1) The 9-Acr¹ ligand (Figure 5), which is equipped with a 4-nitrobenzamido group to ensures cleavage of DNA upon irradiation, is expected only to cleave DNA in close proximity 25 to its binding site. Upon irradiation of the PNA with the above 248 bp DNA fragment, selective cleavage at the dA₁₀/dT₁₀ sequence is observed (Figure 3a).

2) In a so-called photofootprinting assay, where a synthetic diazo-linked acridine under irradiation cleaves DNA 30 (except where the DNA is protected by said binding substance) upon interaction with DNA in the presence of a DNA-binding substance.

Such an experiment was performed with the above 248 bp dsDNA fragment, which showed clear protection against photocleavage of the PNA binding site (Figure 3b).

3) In a similar type of experiment, the DNA-cleaving enzyme micrococcus nuclease, which is also hindered in its

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action by most DNA-binding reagents, showed increased cleavage at the T₁₀-target (Figure 3c).

4) In yet another type of experiment, the well-known high susceptibility of single strand thymine ligands (as opposed to double strand thymine ligands) towards potassium permanganate oxidation was employed. Oxidation of the 248 bp in the presence of the reagent showed only oxidation of the T₁₀-strand of the target (Figure 3b).

5) In a similar type of demonstration, the single strand specificity of S₁ nuclease clearly showed that only the T₁₀-strand of the target was attacked (Figure 3d).

The very efficient binding of (Taeg)₁₀, (Taeg)₁₀-Lys-NH₂ and Acr¹-(Taeg)₁₀-Lys-NH₂ (Figures 11a. 11b) to the corresponding dA₁₀ was furthermore illustrated in two ways:

15 1. Ligand-oligonucleotide complexes will migrate slower than the naked oligonucleotide upon electrophoresis in polyacrylamide gels. Consequently, such experiments were performed with Acr¹-(Taeg)₁₀-Lys-NH₂ and ³²P-end-labelled dA₁₀. This showed retarded migration under conditions where a normal 20 dA₁₀/dT₁₀ duplex is stable, as well as under conditions where such a duplex is unstable (denaturing gel). A control experiment was performed with a mixture of Acr¹-(Taeg)₁₀-Lys-NH₂ and ³²P-end-labelled dT₁₀ which showed no retardation under the above conditions.

25 2. Upon formation of DNA duplexes (dsDNA) from single strand DNA, the extinction coefficient decreases (hypochromicity). Thus, the denaturing of DNA can be followed by measuring changes in the absorbance, for example, as a function of T_m, the temperature where 50% of a duplex has 30 disappeared to give single strands.

Duplexes were formed from the single-stranded oligodeoxyribonucleotides and the PNAs listed below. Typically 0.3 OD₂₆₀ of the T-rich strand was hybridized with 1 equivalent of the other strand by heating to 90 C for 5 min, 35 cooling to room temperature and kept for 30 min and finally stored in a refrigerator at 5 C for at least 30 min. The buffers used were all 10 mM in phosphate and 1 mM in EDTA.

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The low salt buffer contained no sodium chloride, whereas the medium salt buffer contained 140 mM NaCl and the high salt buffer 500 mM NaCl. The pH of all the buffers was 7.2. The melting temperature of the hybrids were determined on a 5 Gilford Response apparatus. The following extinction coefficients were used A: 15.4 ml/ μ mol·cm; T: 8.8; G: 11.7 and C: 7.3 for both normal oligonucleotides and PNA. The melting curves were recorded in steps of 0.5 C/min. The T_m were determined from the maximum of the 1st derivative of the plot 10 of A_{260} vs temperature.

List of oligodeoxyribonucleotides:

1. 5'-AAA-AAA-AA
2. 5'-AAA-AAA-AAA-A
- 15 3. 5'-TTT-TTT-TTT-T
4. 5'-AAA-AAG-AAA-A
5. 5'-AAG-AAG-AAA-A
6. 5'-AAA-AGA-AAA-A
7. 5'-AAA-AGA-AGA-A
- 20 8. 5'-TTT-TCT-TTT-T
9. 5'-TTT-TCT-TCT-T
10. 5'-TTT-TTC-TTT-T
11. 5'-TTT-TTC-TTC-T
12. 5'-TTC-TTC-TTT-T
- 25 13. 5'-TTT-TTT-TTT-TTT-T
14. 5'-AAA-AAA-AAA-AAA-AAA

List of PNAs

- a. TTT-TTT-TTT-T-Lys-NH₂
- 30 b. TTT-TTT-TT-Lys-NH₂
- c. TTT-TTC-TTT-T-Lys-NH₂
- d. TTC-TTC-TTT-T-Lys-NH₂
- e. Acr-TTT-TTT-TTT-T-Lys-NH₂
- f. Ac-TTT-TTT-TTT-T-Lys-NH₂

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Oligo/PNA	Low Salt	Medium Salt	High Salt
1+b	56.0	51.5	50.0
2+a	73.0	72.5	73.0
2+c		41.5 and 52.0*	
5 2+e	84.5	86.0	~90
2+f		74	
4+a	60.0	59.0	61.5
4+c	74.5	72.0	72.5
4+f		62.0	
10 5+a		47.0	
5+c		57.5	
5+f		46.5	
7+a		46.0	
7+c		58.0	
15 7+f		43.5	
7+12		23.0	
13+14		39.0	

* = Two distinct melting temperatures are seen,
indicating local melting before complete
denaturation.

The hybrid formed between RNA-A (poly rA) and PNA-T₁₀-Lys-NH₂ melts at such high temperature that it cannot be measured (>90 C). But specific hybridization is demonstrated by the large drop in A₂₆₀ by mixing with RNA-A but not G,C and U. The experiment is done by mixing 1 ml of a solution of the PNA and 1 ml of a solution the RNA, each with A₂₆₀ = 0.6, and then measure the absorbance at 260 nm. Thereafter the sample is heated to 90 C for 5 min, cooled to room temperature and kept at this temperature for 30 minutes and finally stored at 5C for 30 min.

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RNA	PNA	A_{260} Before Mixing	A_{260} After Mixing	A_{260} After Mixing and Heating
RNA-A	PNA-T ₁₀ -lys-NH ₂	0.600	0.389	0.360
RNA-U	PNA-T ₁₀ -lys-NH ₂	0.600	0.538	0.528
RNA-G	PNA-T ₁₀ -lys-NH ₂	0.600	0.514	0.517
5 RNA-C	PNA-T ₁₀ -lys-NH ₂	0.600	0.540	0.532

From the above measurements the following conclusions can be made. There is base stacking, since a melting curve is observed. The PNA-DNA hybrid is more stable than a normal DNA-DNA hybrid, and the PNA-RNA is even more stable. Mismatches cause significant drops in the T_m -value, whether the mispaired base is in the DNA or in the PNA-strand. The T_m -value is only slightly dependent on ionic strength, as opposed to normal oligonucleotides.

15 The synthesis of the PNAs according to the invention is discussed in detail in the following, where Figure 1 illustrates one of the preferred PNA examples and compares its structure to that of a complementary DNA.

20 Synthesis of PNA Oligomers and Polymers

The principle of anchoring molecules onto a solid matrix, which helps in accounting for intermediate products during chemical transformations, is known as Solid-Phase Synthesis or Merrifield Synthesis (see, e.g., Merrifield, J. 25 Am. Chem. Soc., 1963, 85, 2149 and Science, 1986, 232, 341). Established methods for the stepwise or fragmentwise solid-phase assembly of amino acids into peptides normally employ a beaded matrix of slightly cross-linked styrene-divinylbenzene copolymer, the cross-linked copolymer having been formed 30 by the pearl polymerization of styrene monomer to which has been added a mixture of divinylbenzenes. A level of 1-2% cross-linking is usually employed. Such a matrix also can be used in solid-phase PNA synthesis in accordance with the present invention (Figure 8).

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Concerning the initial functionalization of the solid phase, more than fifty methods have been described in connection with traditional solid-phase peptide synthesis (see, e.g., Barany and Merrifield in "The Peptides" Vol. 2, 5 Academic Press, New York, 1979, pp. 1-284, and Stewart and Young, "Solid Phase Peptide Synthesis", 2nd Ed., Pierce Chemical Company, Illinois, 1984). Reactions for the introduction of chloromethyl functionality (Merrifield resin; via a chloromethyl methyl ether/SnCl₄ reaction), aminomethyl 10 functionality (via an N-hydroxymethylphthalimide reaction; see, Mitchell, et al., Tetrahedron Lett., 1976, 3795), and benzhydryl amino functionality (Pietta, et al., J. Chem. Soc., 1970, 650) are the most widely applied. Regardless of its nature, the purpose of the functionality is normally to form 15 an anchoring linkage between the copolymer solid support and the C-terminus of the first amino acid to be coupled to the solid support. As will be recognized, anchoring linkages also can be formed between the solid support and the amino acid N-terminus. It is generally convenient to express the 20 "concentration" of a functional group in terms of millimoles per gram (mmol/g). Other reactive functionalities which have been initially introduced include 4-methylbenzhydryl amino and 4-methoxybenzhydryl amino. All of these established methods are in principle useful within the context of the present invention. Preferred methods for PNA synthesis employ aminomethyl as the initial functionality, in that aminomethyl is particularly advantageous with respect to the incorporation 25 of "spacer" or "handle" groups, owing to the reactivity of the amino group of the aminomethyl functionality with respect to the essentially quantitative formation of amide bonds to a carboxylic acid group at one end of the spacer-forming reagent. A vast number of relevant spacer- or handle-forming bifunctional reagents have been described (see, Barany, et al., Int. J. Peptide Protein Res., 1987, 30, 705), especially 30 35 reagents which are reactive towards amino groups such as found in the aminomethyl function. Representative bifunctional reagents include 4-(haloalkyl)aryl-lower alkanoic acids such

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as 4-(bromomethyl)phenylacetic acid, Boc-aminoacyl-4-(oxymethyl)aryl-lower alkanoic acids such as Boc-aminoacyl-4-(oxymethyl)phenylacetic acid, N-Boc-p-acylbenzhydrylamines such as N-Boc-p-glutaroylbenzhydrylamine, N-Boc-4'-lower alkyl-p-acylbenzhydrylamines such as N-Boc-4'-methyl-p-glutaroylbenzhydrylamine, N-Boc-4'-lower alkoxy-p-acylbenzhydrylamines such as N-Boc-4'-methoxy-p-glutaroyl-benzhydrylamine, and 4-hydroxymethylphenoxyacetic acid. One type of spacer group particularly relevant within the context of 10 the present invention is the phenylacetamidomethyl (Pam) handle (Mitchell and Merrifield, *J. Org. Chem.*, 1976, 41, 2015) which, deriving from the electron withdrawing effect of the 4-phenylacetamidomethyl group, is about 100 times more stable than the classical benzyl ester linkage towards the 15 Boc-amino deprotection reagent trifluoroacetic acid (TFA).

Certain functionalities (e.g., benzhydrylarnino, 4-methylbenzhydrylarnino and 4-methoxybenzhydrylarnino) which may be incorporated for the purpose of cleavage of a synthesized PNA chain from the solid support such that the C-terminal of 20 the PNA chain is in amide form, require no introduction of a spacer group. Any such functionality may advantageously be employed in the context of the present invention.

An alternative strategy concerning the introduction of spacer or handle groups is the so-called "preformed handle" 25 strategy (see, Tam, et al., *Synthesis*, 1979, 955-957), which offers complete control over coupling of the first amino acid, and excludes the possibility of complications arising from the presence of undesired functional groups not related to the peptide or PNA synthesis. In this strategy, spacer or handle 30 groups, of the same type as described above, are reacted with the first amino acid desired to be bound to the solid support, the amino acid being N-protected and optionally protected at the other side-chains which are not relevant with respect to the growth of the desired PNA chain. Thus, in those cases in 35 which a spacer or handle group is desirable, the first amino acid to be coupled to the solid support can either be coupled to the free reactive end of a spacer group which has been

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bound to the initially introduced functionality (for example, an aminomethyl group) or can be reacted with the spacer-forming reagent. The space-forming reagent is then reacted with the initially introduced functionality. Other useful 5 anchoring schemes include the "multidetachable" resins (Tam, et al., *Tetrahedron Lett.*, 1979, 4935 and *J. Am. Chem. Soc.*, 1980, 102, 611; Tam, *J. Org. Chem.*, 1985, 50, 5291), which provide more than one mode of release and thereby allow more flexibility in synthetic design.

10 Suitable choices for N-protection are the tert-butyloxycarbonyl (Boc) group (Carpino, *J. Am. Chem. Soc.*, 1957, 79, 4427; McKay, et al., *J. Am. Chem. Soc.*, 1957, 79, 4686; Anderson, et al., *J. Am. Chem. Soc.*, 1957, 79, 6180) normally in combination with benzyl-based groups for the 15 protection of side chains, and the 9-fluorenylmethyloxycarbonyl (Fmoc) group (Carpino, et al., *J. Am. Chem. Soc.*, 1970, 92, 5748 and *J. Org. Chem.*, 1972, 37, 3404), normally in combination with tert-butyl (tBu) for the protection of any side chains, although a number of other possibilities exist 20 which are well known in conventional solid-phase peptide synthesis. Thus, a wide range of other useful amino protecting groups exist, some of which are Adoc (Hass, et al., *J. Am. Chem. Soc.*, 1966, 88, 1988), Bpoc (Sieber, *Helv. Chem. Acta.*, 1968, 51, 614), Mcb (Brady, et al., *J. Org. Chem.*, 25 1977, 42, 143), Bic (Kemp, et al., *Tetrahedron*, 1975, 4624), the o-nitrophenylsulfenyl (Nps) (Zervas, et al., *J. Am. Chem. Soc.*, 1963, 85, 3660), and the dithiasuccinoyl (Dts) (Barany, et al., *J. Am. Chem. Soc.*, 1977, 99, 7363). These amino 30 protecting groups, particularly those based on the widely-used urethane functionality, successfully prohibit racemization (mediated by tautomerization of the readily formed oxazolinone (azlactone) intermediates (Goodman, et al., *J. Am. Chem. Soc.*, 1964, 86, 2918)) during the coupling of most α -amino acids. In addition to such amino protecting groups, a whole range of 35 otherwise "worthless" nonurethane-type of amino protecting groups are applicable when assembling PNA molecules,

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especially those built from achiral units. Thus, not only the above-mentioned amino protecting groups (or those derived from any of these groups) are useful within the context of the present invention, but virtually any amino protecting group which largely fulfills the following requirements: (1) stability to mild acids (not significantly attacked by carboxyl groups); (2) stability to mild bases or nucleophiles (not significantly attacked by the amino group in question); (3) resistance to acylation (not significantly attacked by activated amino acids). Additionally: (4) the protecting group must be close to quantitatively removable, without serious side reactions, and (5) the optical integrity, if any, of the incoming amino acid should preferably be highly preserved upon coupling. Finally, the choice of side-chain protecting groups, in general, depends on the choice of the amino protecting group, since the protection of side-chain functionalities must withstand the conditions of the repeated amino deprotection cycles. This is true whether the overall strategy for chemically assembling PNA molecules relies on, for example, differential acid stability of amino and side-chain protecting groups (such as is the case for the above-mentioned "Boc-benzyl" approach) or employs an orthogonal, that is, chemoselective, protection scheme (such as is the case for the above-mentioned "Fmoc-tBu" approach),

Following coupling of the first amino acid, the next stage of solid-phase synthesis is the systematic elaboration of the desired PNA chain. This elaboration involves repeated deprotection/coupling cycles. The temporary protecting group, such as a Boc or Fmoc group, on the last-coupled amino acid is quantitatively removed by a suitable treatment, for example, by acidolysis, such as with trifluoroacetic acid, in the case of Boc, or by base treatment, such as with piperidine, in the case of Fmoc, so as to liberate the N-terminal amine function.

The next desired N-protected amino acid is then coupled to the N-terminal of the last-coupled amino acid. This coupling of the C-terminal of an amino acid with the N-

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terminal of the last-coupled amino acid can be achieved in several ways. For example, the carboxyl group of the incoming amino acid can be reacted directly with the N-terminal of the last-coupled amino acid with the assistance of a condensation reagent such as, for example, dicyclohexylcarbodiimide (DCC) (Sheehan & Hess, et al., *J. Am. Chem. Soc.*, 1955, 77, 1067) and diisopropylcarbodiimide (DIC) (Sraantakis et al., *Biochem. biophys. res. Commun.*, 1976, 73, 336) or derivatives thereof. Alternatively, it can be bound by providing the incoming amino acid in a form with the carboxyl group activated by any of several methods, including the initial formation of an active ester derivative such as a 2,4,5-trichlorophenyl ester (Pless, et al., *Helv. Chim. Acta*, 1963, 46, 1609), a phthalimido ester (Nefkens, et al., *J. Am. Chem. Soc.*, 1961, 83, 1263), a pentachlorophenyl ester (Kupryszewski, *Roczn. Chem.*, 1961, 35, 595), a pentafluorophenyl ester (Kovacs, et al., *J. Am. Chem. Soc.*, 1963, 85, 183), an o-nitrophenyl ester (Bodanzsky, *Nature*, 1955, 175, 685), an imidazole ester (Li, et al., *J. Am. Chem. Soc.*, 1970, 92, 7608), and a 3-hydroxy-4-oxo-3,4-dihydroquinazoline (Dhbtt-OH) ester (Konig, et al., *Chem. Ber.*, 1973, 103, 2024 and 2034), or the initial formation of an anhydride such as a symmetrical anhydride (Wieland, et al., *Angew. Chem., Int. Ed. Engl.*, 1971, 10, 336). Benzotriazolyl N-oxytrisdimethylaminophosphonium hexafluorophosphate (BOP), "Castro's reagent" (see, e.g., Rivaille, et al., *Tetrahedron*, 1980, 36, 3413) is recommended when assembling PNA molecules containing secondary amino groups. Finally, activated PNA monomers analogous to the recently-reported amino acid fluorides (Carpino, *J. Am. Chem. Soc.*, 1990, 112, 9651) hold considerable promise to be used in PNA synthesis as well.

Following assembly of the desired PNA chain, including protecting groups, the next step will normally be deprotection of the amino acid moieties of the PNA chain and cleavage of the synthesized PNA from the solid support. These processes can take place substantially simultaneously, thereby providing the free PNA molecule in the desired form. Alternatively, in cases in which condensation of two separately synthesized PNA

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chains is to be carried out, it is possible by choosing a suitable spacer group at the start of the synthesis to cleave the desired PNA chains from their respective solid supports (both peptide chains still incorporating their side-chain 5 protecting groups) and finally removing the side-chain protecting groups after, for example, coupling the two side-chain protected peptide chains to form a longer PNA chain.

In the above-mentioned "Boc-benzyl" protection scheme, the final deprotection of side-chains and release of the PNA 10 molecule from the solid support is most often carried out by the use of strong acids such as anhydrous HF (Sakakibara, et al., *Bull. Chem. Soc. Jpn.*, 1965, 38, 4921), boron tris (trifluoroacetate) (Pless, et al., *Helv. Chim. Acta*, 1973, 46, 1609), and sulfonic acids such as trifluoromethanesulfonic 15 acid and methanesulfonic acid (Yajima, et al., *J. Chem. Soc., Chem. Comm.*, 1974, 107). This conventional strong acid (e.g., anhydrous HF) deprotection method, produces very reactive carbocations that may lead to alkylation and acylation of sensitive residues in the PNA chain. Such side-reactions are 20 only partly avoided by the presence of scavengers such as anisole, phenol, dimethyl sulfide, and mercaptoethanol and, therefore, the sulfide-assisted acidolytic S_N2 deprotection method (Tam, et al., *J. Am. Chem. Soc.*, 1983, 105, 6442 and *J. Am. Chem. Soc.*, 1986, 108, 5242), the so-called "low", 25 which removes the precursors of harmful carbocations to form inert sulfonium salts, is frequently employed in peptide and PNA synthesis, either solely or in combination with "high" methods. Less frequently, in special cases, other methods used for deprotection and/or final cleavage of the PNA-solid 30 support bond are, for example, such methods as base-catalyzed alcoholysis (Barton, et al., *J. Am. Chem. Soc.*, 1973, 95, 4501), and ammonolysis as well as hydrazinolysis (Bodanszky, et al., *Chem. Ind.*, 1964 1423), hydrogenolysis (Jones, *Tetrahedron Lett.* 1977 2853 and Schlatter, et al., *Tetrahedron Lett.* 1977 2861)), and photolysis (Rich and Gurwara, *J. Am. Chem. Soc.*, 1975 97, 1575)).

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Finally, in contrast with the chemical synthesis of "normal" peptides, stepwise chain building of achiral PNAs such as those based on aminoethylglycyl backbone units can start either from the N-terminus or the C-terminus, because 5 the coupling reactions are free of racemization. Those skilled in the art will recognize that whereas syntheses commencing at the C-terminus typically employ protected amine groups and free or activated acid groups, syntheses commencing at the N-terminus typically employ protected acid groups and 10 free or activated amine groups.

Based on the recognition that most operations are identical in the synthetic cycles of solid-phase peptide synthesis (as is also the case for solid-phase PNA synthesis), a new matrix, PEPS, was recently introduced (Berg, et al., J. 15 Am. Chem. Soc., 1989, 111, 8024 and International Patent Application WO 90/02749) to facilitate the preparation of large numbers of peptides. This matrix is comprised of a polyethylene (PE) film with pendant long-chain polystyrene (PS) grafts (molecular weight on the order of 10^6). The 20 loading capacity of the film is as high as that of a beaded matrix, but PEPS has the additional flexibility to suit multiple syntheses simultaneously. Thus, in a new configuration for solid-phase peptide synthesis, the PEPS film is fashioned in the form of discrete, labeled sheets, each 25 serving as an individual compartment. During all the identical steps of the synthetic cycles, the sheets are kept together in a single reaction vessel to permit concurrent preparation of a multitude of peptides at a rate close to that of a single peptide by conventional methods. It was reasoned 30 that the PEPS film support, comprising linker or spacer groups adapted to the particular chemistry in question, should be particularly valuable in the synthesis of multiple PNA molecules, these being conceptually simple to synthesize since only four different reaction compartments are normally 35 required, one for each of the four "pseudo-nucleotide" units. Thus, the PEPS film support has been successfully tested in a number of PNA syntheses carried out in a parallel and

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substantially simultaneous fashion. The yield and quality of the products obtained from PEPS were comparable to those obtained by using the traditional polystyrene beaded support. Also, experiments with other geometries of the PEPS polymer 5 such as, for example, non-woven felt, knitted net, sticks or microwellplates have not indicated any limitations of the synthetic efficacy.

Two other methods proposed for the simultaneous synthesis of large numbers of peptides also apply to the 10 preparation of multiple, different PNA molecules. The first of these methods (Geysen, et al., *Proc. Natl. Acad. Sci. USA*, 1984, 81, 3998) utilizes acrylic acid-grafted polyethylene-rods and 96-microtiter wells to immobilize the growing peptide chains and to perform the compartmentalized synthesis. While 15 highly effective, the method is only applicable on a microgram scale. The second method (Houghten, *Proc. Natl. Acad. Sci. USA*, 1985, 82, 5131) utilizes a "tea bag" containing traditionally-used polymer beads. Other relevant proposals for multiple peptide or PNA synthesis in the context of the 20 present invention include the simultaneous use of two different supports with different densities (Tregebar, in "*Chemistry and Biology of Peptides*", J. Meienhofer, ed., Ann Arbor Sci. Publ., Ann Arbor, 1972 pp. 175-178), combining of reaction vessels via a manifold (Gorman, *Anal. Biochem.*, 1984, 25 136, 397), multicolumn solid-phase synthesis (e.g. Krchnak, et al., *Int. J. Peptide Protein Res.*, 1989, 33, 209), and Holm and Meldal, in "*Proceedings of the 20th European Peptide Symposium*", G. Jung and E. Bayer, eds., Walter de Gruyter & Co., Berlin, 1989 pp. 208-210), and the use of cellulose paper 30 (Eichler, et al., *Collect. Czech. Chem. Commun.*, 1989, 54, 1746).

While the conventional cross-linked styrene/divinylbenzene copolymer matrix and the PEPS support are presently preferred in the context of solid-phase PNA 35 synthesis, a non-limiting list of examples of solid supports which may be of relevance are: (1) Particles based upon copolymers of dimethylacrylamide cross-linked with N,N'-

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bisacryloylethylenediamine, including a known amount of N-tert-butoxy carbonyl-beta-alanyl-N'-acryloylhexamethylenediamine. Several spacer molecules are typically added via the beta alanyl group, followed thereafter by the amino acid residue subunits. Also, the beta alanyl-containing monomer can be replaced with an acryloyl sarcosine monomer during polymerization to form resin beads. The polymerization is followed by reaction of the beads with ethylenediamine to form resin particles that contain primary amines as the covalently linked functionality. The polyacrylamide-based supports are relatively more hydrophilic than are the polystyrene-based supports and are usually used with polar aprotic solvents including dimethylformamide, dimethylacetamide, N-methylpyrrolidone and the like (see Atherton, et al., *J. Am. Chem. Soc.*, 1975, 97, 6584, *Bioorg. Chem.* 1979, 8, 351), and J.C.S. Perkin I 538 (1981)); (2) a second group of solid supports is based on silica-containing particles such as porous glass beads and silica gel. One example is the reaction product of trichloro-[3-(4-chloromethyl)phenyl]propylsilane and porous glass beads (see Parr and Grohmann, *Angew. Chem. Internal. Ed.* 1972, 11, 314) sold under the trademark "PORASIL E" by Waters Associates, Framingham, MA, USA. Similarly, a mono ester of 1,4-dihydroxymethylbenzene and silica (sold under the trademark "BIOPAK" by Waters Associates) has been reported to be useful (see Bayer and Jung, *Tetrahedron Lett.*, 1970, 4503); (3) a third general type of useful solid supports can be termed composites in that they contain two major ingredients: a resin and another material that is also substantially inert to the organic synthesis reaction conditions employed. One exemplary composite (see Scott, et al., *J. Chrom. Sci.*, 1971, 9, 577) utilized glass particles coated with a hydrophobic, cross-linked styrene polymer containing reactive chloromethyl groups, and was supplied by Northgate Laboratories, Inc., of Hamden, CT, USA. Another exemplary composite contains a core of fluorinated ethylene polymer onto which has been grafted polystyrene (see Kent and Merrifield, *Israel J. Chem.* 1978,

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17, 243) and van Rietschoten in "Peptides 1974", Y. Wolman, Ed., Wiley and Sons, New York, 1975, pp. 113-116); and (4) contiguous solid supports other than PEPS, such as cotton sheets (Lebl and Eichler, *Peptide Res.* 1989, 2, 232) and 5 hydroxypropylacrylate-coated polypropylene membranes (Daniels, et al., *Tetrahedron Lett.* 1989, 4345), are suited for PNA synthesis as well.

Whether manually or automatically operated, solid-phase PNA synthesis in the context of the present invention is 10 normally performed batchwise. However, most of the syntheses may equally well be carried out in the continuous-flow mode, where the support is packed into columns (Bayer, et al., *Tetrahedron Lett.*, 1970, 4503 and Scott, et al., *J. Chromatogr. Sci.*, 1971, 9, 577). With respect to continuous- 15 flow solid-phase synthesis, the rigid poly(dimethylacrylamide)-Kieselguhr support (Atherton, et al., *J. Chem. Soc. Chem. Commun.*, 1981, 1151) appears to be particularly successful, but another valuable configuration concerns the one worked out for the standard copoly(styrene-1%-divinylbenzene) support 20 (Krchnak, et al., *Tetrahedron Lett.*, 1987, 4469).

While the solid-phase technique is presently preferred in the context of PNA synthesis, other methodologies or combinations thereof, for example, in combination with the solid-phase technique, apply as well: (1) the classical 25 solution-phase methods for peptide synthesis (e.g., Bodanszky, "Principles of Peptide Synthesis", Springer-Verlag, Berlin-New York 1984), either by stepwise assembly or by segment/fragment condensation, are of particular relevance when considering especially large scale productions (gram, kilogram, and even 30 tons) of PNA compounds; (2) the so-called "liquid-phase" strategy, which utilizes soluble polymeric supports such as linear polystyrene (Shemyakin, et al., *Tetrahedron Lett.*, 1965, 2323) and polyethylene glycol (PEG) (Mutter and Bayer, *Angew. Chem., Int. Ed. Engl.*, 1974, 13, 88), is useful; (3) 35 random polymerization (see, e.g., Odian, "Principles of Polymerization", McGraw-Hill, New York (1970)) yielding

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mixtures of many molecular weights ("polydisperse") peptide or PNA molecules are particularly relevant for purposes such as screening for antiviral effects; (4) a technique based on the use of polymer-supported amino acid active esters
5 (Fridkin, et al., *J. Am. Chem. Soc.*, 1965, 87, 4646), sometimes referred to as "inverse Merrifield synthesis" or "polymeric reagent synthesis", offers the advantage of isolation and purification of intermediate products, and may thus provide a particularly suitable method for the synthesis
10 of medium-sized, optionally protected, PNA molecules, that can subsequently be used for fragment condensation into larger PNA molecules; (5) it is envisaged that PNA molecules may be assembled enzymatically by enzymes such as proteases or derivatives thereof with novel specificities (obtained, for
15 example, by artificial means such as protein engineering). Also, one can envision the development of "PNA ligases" for the condensation of a number of PNA fragments into very large PNA molecules; (6) since antibodies can be generated to virtually any molecule of interest, the recently developed
20 catalytic antibodies (abzymes), discovered simultaneously by the groups of Lerner (Tramantano, et al., *Science*, 1986, 234, 1566) and of Schultz (Pollack, et al., *Science*, 1986, 234, 1570), should also be considered as potential candidates for assembling PNA molecules. Thus, there has been considerable
25 success in producing abzymes catalyzing acyl-transfer reactions (see for example Shokat, et al., *Nature*, 1989, 338, 269) and references therein). Finally, completely artificial enzymes, very recently pioneered by Stewart's group (Hahn, et al., *Science*, 1990, 248, 1544), may be developed to suit PNA
30 synthesis. The design of generally applicable enzymes, ligases, and catalytic antibodies, capable of mediating specific coupling reactions, should be more readily achieved for PNA synthesis than for "normal" peptide synthesis since PNA molecules will often be comprised of only four different
35 amino acids (one for each of the four native nucleobases) as compared to the twenty natural by occurring (proteinogenic) amino acids constituting peptides. In conclusion, no single

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strategy may be wholly suitable for the synthesis of a specific PNA molecule, and therefore, sometimes a combination of methods may work best.

The present invention also is directed to therapeutic or prophylactic uses for peptide nucleic acids. Likely therapeutic and prophylactic targets include herpes simplex virus (HSV), human papillomavirus (HPV), human immunodeficiency virus (HIV), candida albicans, influenza virus, cytomegalovirus (CMV), intracellular adhesion molecules 10 (ICAM), 5-lipoxygenase (5-LO), phospholipase A₂ (PLA₂), protein kinase C (PKC), and RAS oncogene. Potential applications of such targeting include treatments for ocular, labial, genital, and systemic herpes simplex I and II infections; genital warts; cervical cancer; common warts; 15 Kaposi's sarcoma; AIDS; skin and systemic fungal infections; flu; pneumonia; retinitis and pneumonitis in immunosuppressed patients; mononucleosis; ocular, skin and systemic inflammation; cardiovascular disease; cancer; asthma; psoriasis; cardiovascular collapse; cardiac infarction; 20 gastrointestinal disease; kidney disease; rheumatoid arthritis; osteoarthritis; acute pancreatitis; septic shock; and Crohn's disease.

For therapeutic or prophylactic treatment, the peptide nucleic acids of the invention can be formulated in a pharmaceutical composition, which may include carriers, thickeners, diluents, buffers, preservatives, surface active agents and the like. Pharmaceutical compositions may also include one or more active ingredients such as antimicrobial agents, antiinflammatory agents, anesthetics, and the like in 30 addition to peptide nucleic acid.

The pharmaceutical composition may be administered in a number of ways depending on whether local or systemic treatment is desired, and on the area to be treated. Administration may be done topically (including 35 ophthalmically, vaginally, rectally, intranasally), orally, by inhalation, or parenterally, for example by intravenous

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drip or subcutaneous, intraperitoneal or intramuscular injection.

Formulations for topical administration may include ointments, lotions, creams, gels, drops, suppositories, 5 sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable. Coated condoms may also be useful.

Compositions for oral administration include powders 10 or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets, or tablets. Thickeners, flavorings, diluents, emulsifiers, dispersing aids or binders may be desirable.

Formulations for parenteral administration may include 15 sterile aqueous solutions which may also contain buffers, diluents and other suitable additives.

Dosing is dependent on severity and responsiveness of the condition to be treated, but will normally be one or more doses per day, with course of treatment lasting from several 20 days to several months or until a cure is effected or a diminution of disease state is achieved. Persons of ordinary skill can easily determine optimum dosages, dosing methodologies and repetition rates.

Treatments of this type can be practiced on a variety 25 of organisms ranging from unicellular prokaryotic and eukaryotic organisms to multicellular eukaryotic organisms. Any organism that utilizes DNA-RNA transcription or RNA-protein translation as a fundamental part of its hereditary, metabolic or cellular control is susceptible to therapeutic and/or 30 prophylactic treatment in accordance with the invention. Seemingly diverse organisms such as bacteria, yeast, protozoa, algae, all plants and all higher animal forms, including warm-blooded animals, can be treated. Further, since each cell of multicellular eukaryotes can be treated since they include 35 both DNA-RNA transcription and RNA-protein translation as integral parts of their cellular activity. Furthermore, many of the organelles (e.g., mitochondria and chloroplasts) of

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eukaryotic cells also include transcription and translation mechanisms. Thus, single cells, cellular populations or organelles can also be included within the definition of organisms that can be treated with therapeutic or diagnostic 5 phosphorothioate oligonucleotides. As used herein, therapeutics is meant to include the eradication of a disease state, by killing an organism or by control of erratic or harmful cellular growth or expression.

The present invention also pertains to the advantageous 10 use of PNA molecules in solid-phase biochemistry (see, e.g., "Solid-Phase Biochemistry - Analytical and Synthetic Aspects", W. H. Scouten, ed., John Wiley & Sons, New York, 1983), notably solid-phase biosystems, especially bioassays or solid-phase techniques which concerns diagnostic detection/quantification 15 or affinity purification of complementary nucleic acids (see, e.g., "Affinity Chromatography - A Practical Approach", P. D. G. Dean, W. S. Johnson and F. A. Middle, eds., IRL Press Ltd., Oxford 1986; "Nucleic Acid Hybridization - A Practical Approach", B. D. Barnes and S. J. Higgins, IRL Press Ltd., 20 Oxford 1987). Present day methods for performing such bioassays or purification techniques almost exclusively utilize "normal" or slightly modified oligonucleotides either physically adsorbed or bound through a substantially permanent covalent anchoring linkage to beaded solid supports such as 25 cellulose, glass beads, including those with controlled porosity (Mizutani, et al., *J. Chromatogr.*, 1986, 356, 202), "Sephadex", "Sephadose", agarose, polyacrylamide, porous particulate alumina, hydroxyalkyl methacrylate gels, diol-bonded silica, porous ceramics, or contiguous-materials such 30 as filter discs of nylon and nitrocellulose. One example employed the chemical synthesis of oligo-dT on cellulose beads for the affinity isolation of poly A tail containing mRNA (Gilham in "Methods in Enzymology," L. Grossmann and K. Moldave, eds., vol. 21, part D, page 191, Academic Press, New 35 York and London, 1971). All the above-mentioned methods are applicable within the context of the present invention. However, when possible, covalent linkage is preferred over the

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physical adsorption of the molecules in question, since the latter approach has the disadvantage that some of the immobilized molecules can be washed out (desorbed) during the hybridization or affinity process. There is, thus, little
5 control of the extent to which a species adsorbed on the surface of the support material is lost during the various treatments to which the support is subjected in the course of the bioassay/purification procedure. The severity of this problem will, of course, depend to a large extent on the rate
10 at which equilibrium between adsorbed and "free" species is established. In certain cases it may be virtually impossible to perform a quantitative assay with acceptable accuracy and/or reproducibility. Loss of adsorbed species during treatment of the support with body fluids, aqueous reagents
15 or washing media will, in general, be expected to be most pronounced for species of relatively low molecular weight. In contrast with oligonucleotides, PNA molecules are easier to attach onto solid supports because they contain strong nucleophilic and/or electrophilic centers. In addition, the
20 direct assembly of oligonucleotides onto solid supports suffers from an extremely low loading of the immobilized molecule, mainly due to the low surface capacity of the materials that allow the successful use of the state-of-the-art phosphoramidite chemistry for the construction of oligo-
25 nucleotides. (Beaucage and Caruthers, *Tetrahedron Lett.*, 1981,
22, 1859; Caruthers, *Science*, 1985, 232, 281). It also suffers from the fact that by using the alternative phosphite triester method (Letsinger and Mahadevan, *J. Am. Chem. Soc.*
1976, 98, 3655), which is suited for solid supports with a
30 high surface/loading capacity, only relatively short oligonucleotides can be obtained. As for conventional solid-phase peptide synthesis, however, the latter supports are excellent materials for building up immobilized PNA molecules (the side-chain protecting groups are removed from the synthesized PNA
35 chain without cleaving the anchoring linkage holding the chain to the solid support). Thus, PNA species benefit from the above-described solid-phase techniques with respect to the

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much higher (and still sequence-specific) binding affinity for complementary nucleic acids and from the additional unique sequence-specific recognition of (and strong binding to) nucleic acids present in double-stranded structures. They
5 also can be loaded onto solid supports in large amounts, thus further increasing the sensitivity/capacity of the solid-phase technique. Further, certain types of studies concerning the use of PNA in solid-phase biochemistry can be approached, facilitated, or greatly accelerated by use of the recently-
10 reported "light-directed, spatially addressable, parallel chemical synthesis" technology (Fodor, et al., *Science*, 1991, 251, 767), a technique that combines solid-phase chemistry and photolithography to produce thousands of highly diverse, but identifiable, permanently immobilized compounds (such as
15 peptides) in a substantially simultaneous way.

Additional objects, advantages, and novel features of this invention will become apparent to those skilled in the art upon examination of the following examples thereof, which
20 are not intended to be limiting.

synthesis of monomeric building blocks

The monomers preferably are synthesized by the general scheme outlined in Figure 13. This involves preparation of
25 either the methyl or ethyl ester of (Bocaminoethyl)glycine, by a protection/deprotection procedure as described in Examples 24-26. The synthesis of thymine monomer is described in Examples 27-28, and that of the protected cytosine monomer is described in Example 29.

30 The synthesis of the protected adenine monomer (Figure 14) involved alkylation with ethyl bromoacetate (Example 30) and verification of the position of substitution by X-ray crystallography, as being the wanted 9-position. The N⁶-amino group then was protected with the benzyloxycarbonyl group by
35 the use of the reagent N-ethyl-benzyloxycarbonylimidazole tetrafluoroborate (Example 31). Simple hydrolysis of the product ester (Example 32) gave N⁶-benzyloxycarbonyl-9-

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carboxymethyl adenine, which then was used in the standard procedure (Examples 33-34, Figure 13). The adenine monomer has been built into two different PNA-oligomers (Examples 56, 57, 71 and 73).

5 The synthesis of the protected G-monomer is outlined in Figure 15. The starting material, 2-amino-6-chloropurine, was alkylated with bromoacetic acid (Example 35) and the chlorine atom was then substituted with a benzyloxy group (Example 36). The resulting acid was coupled to the
10 (bocaminoethyl) glycine methyl ester (from Example 26) with agent PyBrop™, and the resulting ester was hydrolysed (Example 37). The O⁶-benzyl group was removed in the final HF-cleavage step in the synthesis of the PNA-oligomer. Cleavage was verified by finding the expected mass of the final PNA-
15 oligomer, upon incorporation into an PNA-oligomer using diisopropyl carbodiimide as the condensation agent (Examples 55 and 71).

Extended Backbones

Alterations of the groups A, C and D (figure 16) is
20 demonstrated by the synthesis of monomeric building blocks and incorporation into PNA-oligomers.

In one example, the C group was a CH(CH₃) group. The synthesis of the corresponding monomer is outlined in Figure 17. It involves preparation of Boc-protected 1-amino-2,3-
25 propanediol (Example 38), which is cleaved by periodate to give bocaminoacetaldehyde, which is used directly in the next reaction. The bocaminoacetaldehyde can be condensed with a variety of amines; in Example 39, alanine ethyl ester was used. In Examples 40-42, the corresponding thymine monomers
30 were prepared. The monomer has been incorporated into an 8-mer (Example 60) by the DCC-coupling protocol (Examples 56 and 57).

In another example, the D group is a (CH₂)₃ group. The synthesis of the corresponding monomer is outlined in figure 35 18.A and described in Examples 43-44.

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In another example, the A group is a $(\text{CH}_2)_2\text{CO}$ group. The synthesis of the corresponding thymine monomer is outlined figure 18.B and Examples 46 through 48.

In yet another example, the C group is a $(\text{CH}_2)_2$ group.
5 The synthesis of the thymine and protected cytosine monomer is outlined in Figure 19 and Examples 49 through 54. Hybridization experiments with a PNA-oligomer containing one unit is described in Examples 61 and 81, which shows a significant lowering of affinity but a retention of
10 specificity.

General Remarks

The following abbreviations are used in the experimental examples: DMF, N,N-dimethylformamide; DCC, N,N-dicyclohexyl carbodiimide; DCU, N,N-dicyclohexyl urea; THF, 15 tetrahydrofuran; aeg, N-acetyl (2'-aminoethyl)glycine; pfp, pentafluorophenyl; Boc, tert-butoxycarbonyl; Z, benzyloxy-carbonyl; NMR, nuclear magnetic resonance; s, singlet; d, doublet; dd, doublet of doublets; t; triplet; q, quartet; m, multiplet; b, broad; δ , chemical shift;

20 NMR spectra were recorded on either a JEOL FX 90Q spectrometer, or a Bruker 250 MHz with tetramethylsilane as internal standard. Mass spectrometry was performed on a MassLab VG 12-250 quadropole instrument fitted with a VG FAB source and probe. Melting points were recorded on Buchi 25 melting point apparatus and are uncorrected. N,N-Dimethylformamide was dried over 4 Å molecular sieves, distilled and stored over 4 Å molecular sieves. Pyridine (HPLC quality) was dried and stored over 4 Å molecular sieves. Other solvents used were either the highest quality obtainable 30 or were distilled before use. Dioxane was passed through basic alumina prior to use. Bocanhydride, 4-nitrophenol, methyl bromoacetate, benzyloxycarbonyl chloride, pentafluorophenol were all obtained through Aldrich Chemical Company. Thymine, cytosine, adenine were all obtained through 35 Sigma.

Thin layer chromatography (Tlc) was performed using the following solvent systems: (1) chloroform:triethyl

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amine:methanol, 7:1:2; (2) methylene chloride:methanol, 9:1; (3) chloroform:methanol:acetic acid 85:10:5. Spots were visualized by UV (254 nm) or/and spraying with a ninhydrin solution (3 g ninhydrin in 1000 ml 1-butanol and 30 ml acetic acid), after heating at 120°C for 5 min and, after spraying, heating again.

EXAMPLE 1

tert-Butyl 4-nitrophenyl carbonate

10 Sodium carbonate (29.14 g; 0.275 mol) and 4-nitrophenol (12.75 g; 91.6 mmol) were mixed with dioxane (250 ml). Boc-anhydride (20.0 g; 91.6 mmol) was transferred to the mixture with dioxane (50 ml). The mixture was refluxed for 1 h, cooled to 0°C, filtered and concentrated to 1/3, and then 15 poured into water (350 ml) at 0°C. After stirring for 1/2 h., the product was collected by filtration, washed with water, and then dried over sicapent, *in vacuo*. Yield 21.3 g (97%). M.p. 73.0-74.5°C (litt. 78.5-79.5°C). Anal. for C₁₁H₁₃NO₅ found(calc.) C: 55.20(55.23) H: 5.61(5.48) N: 5.82(5.85).

20

EXAMPLE 2

(N'-Boc-2'-aminoethyl)glycine (2)

The title compound was prepared by a modification of the procedure by Heimer, et al. *Int. J. Pept.*, 1984, 23, 203-211. N-(2-Aminoethyl)glycine (1, 3.00 g; 25.4 mmol) was dissolved in water (50 ml), dioxane (50 ml) was added, and the pH was adjusted to 11.2 with 2 N sodium hydroxide. tert-Butyl-4-nitrophenyl carbonate (7.29 g; 30.5 mmol) was dissolved in dioxane (40 ml) and added dropwise over a period 30 of 2 h, during which time the pH was maintained at 11.2 with 2 N sodium hydroxide. The pH was adjusted periodically to 11.2 for three more hours and then the solution was left overnight. The solution was cooled to 0°C and the pH was carefully adjusted to 3.5 with 0.5 M hydrochloric acid. The 35 aqueous solution was washed with chloroform (3 x 200 ml), the pH adjusted to 9.5 with 2N sodium hydroxide and the solution was evaporated to dryness, *in vacuo* (14 mmHg). The residue

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was extracted with DMF (25+2x10 ml) and the extracts filtered to remove excess salt. This results in a solution of the title compound in about 60% yield and greater than 95% purity by tlc (system 1 and visualised with ninhydrin, Rf=0.3). The 5 solution was used in the following preparations of Boc-aeg derivates without further purification.

EXAMPLE 3

N-1-Carboxymethylthymine (4)

10 This procedure is different from the literature synthesis, but is easier, gives higher yields, and leaves no unreacted thymine in the product. To a suspension of thymine (3, 40.0 g; 0.317 mol) and potassium carbonate (87.7 g; 0.634 mmol) in DMF (900 ml) was added methyl bromoacetate (30.00 ml; 15 0.317 mmol). The mixture was stirred vigorously overnight under nitrogen. The mixture was filtered and evaporated to dryness, *in vacuo*. The solid residue was treated with water (300 ml) and 4 N hydrochloric acid (12 ml), stirred for 15 min at 0°C, filtered, and washed with water (2 x 75 ml). The 20 precipitate was treated with water (120 ml) and 2N sodium hydroxide (60 ml), and was boiled for 10 minutes. The mixture was cooled to 0°C, filtered, and the pure title compound was precipitated by the addition of 4 N hydrochloric acid (70 ml). Yield after drying, *in vacuo* over sicapent: 37.1 g (64%). ¹H-NMR: (90 MHz; DMSO-d₆): 11.33 ppm (s, 1H, NH); 25 7.49 (d, J=0.92Hz, 1H, ArH); 4.38 (s, 2H, CH₂); 1.76 (d, J=0.92Hz, T-CH₃)

EXAMPLE 4

30 N-1-Carboxymethylthymine pentafluorophenyl ester (5)

N-1-Carboxymethylthymine (4, 10.0g; 54.3 mmol) and pentafluorophenol (10.0 g; 54.3 mmol) were dissolved in DMF (100 ml) and cooled to 5°C in ice water. DCC (13.45 g; 65.2 mmol) then was added. When the temperature passed below 5°C, 35 the ice bath was removed and the mixture was stirred for 3 h at ambient temperature. The precipitated DCU was removed by filtration and washed twice with DMF (2 x 10 ml). The

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combined filtrate was poured into ether (1400 ml) and cooled to 0°C. Petroleum ether (1400 ml) was added and the mixture was left overnight. The title compound was isolated by filtration and was washed thoroughly with petroleum ether.

5 Yield: 14.8 g(78%). The product was pure enough to carry out the next reaction, but an analytical sample was obtained by recrystallization from 2-propanol. M.p. 200.5-206°C Anal. for C₁₃H₇F₅N₂O₄. Found(calc.) C: 44.79(44.59); H: 2.14(2.01) N: 8.13(8.00). FAB-MS: 443 (M+1+glycerol), 351 (M+1). ¹H-NMR (90 MHz; DMSO-d₆): 11.52 ppm (s,1H,NH); 7.64 (s,1H,ArH); 4.99 (s,2H,CH₂); 1.76 (s,3H,CH₃).

EXAMPLE 5

1-(Boc-aeg)thymine (6)

15 To the DMF-solution from above was added triethyl amine (7.08 ml; 50.8 mmol) followed by N-1-carboxymethylthymine pentafluorophenyl ester (5, 4.45 g; 12.7 mmol). The resultant solution was stirred for 1 h. The solution was cooled to 0°C and treated with cation exchange material ("Dowex 50W X-8", 20 40 g) for 20 min. The cation exchange material was removed by filtration, washed with dichloromethane (2 x 15 ml), and dichloromethane (150 ml) was added. The resulting solution was washed with saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness, *in vacuo*, first 25 by a water aspirator and then by an oil pump. The residue was shaken with water (50 ml) and evaporated to dryness. This procedure was repeated once. The residue then was dissolved in methanol (75 ml) and poured into ether (600 ml) and petroleum ether (1.4 L). After stirring overnight, the white 30 solid was isolated by filtration and was washed with petroleum ether. Drying over sicapent, *in vacuo*, gave 3.50 g (71.7%). M.p. 142-147°C. Anal. for C₁₆H₂₄N₄O₇. Found(calc.) C: 49.59(50.00) H: 6.34(6.29) N: 14.58(14.58). ¹H-NMR (250 MHz, DMSO-d₆): Due to the limited rotation around the secondary 35 amide bond several of the signals were doubled in the ratio 2:1,(indicated in the list by mj. for major and mi. for minor). 12.73 ppm (b,1H, -CO₂H); 11.27 ppm (s, mj., imide);

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11.25 ppm (s, mi., imide); 7.30 ppm (s, mj., ArH); 7.26 ppm (s, mi., ArH); 6.92 ppm (unres. t, mj., BocNH); 6.73 ppm (unres. t; mi., BocNH); 4.64 ppm (s, mj., T-CH₂-CO-); 4.47 ppm (s, mi., T-CH₂-CO-); 4.19 ppm (s, mi., CONRCH₂CO₂H); 3.97 ppm (s, mj., CONRCH₂CO₂H); 3.41-2.89 ppm (unres. m, -CH₂CH₂- and water); 1.75 ppm (s, 3H, T-CH₃); 1.38 ppm (s, 9H, t-Bu). ¹³C-NMR: 170.68 ppm (CO); 170.34 (CO); 167.47 (CO); 167.08 (CO); 164.29 (CO); 150.9 (C5''); 141.92 (C6''); 108.04 (C2'); 77.95 and 77.68 (Thy-CH₂CO); 48.96, 47.45 and 46.70 (-CH₂CH₂- and NCH₂CO₂H); 37.98 (Thy-CH₃); 28.07 (t-Bu). FAB-MS: 407 (M+Na⁺); 385 (M+H⁺).

EXAMPLE 6

1-(Boc-aeg)thymine pentafluorophenyl ester (7, Boc-Taeg.OPfp)

15 1-(Boc-aeg)thymine (6) (2.00 g; 5.20 mmol) was dissolved in DMF (5 ml) and methylene chloride (15 ml) was added. Pentafluorophenol (1.05 g; 5.72 mmol) was added and the solution was cooled to 0°C in an ice bath. DDC then was added (1.29 g; 6.24 mmol) and the ice bath was removed after 20 2 min. After 3 h with stirring at ambient temperature, the precipitated DCU was removed by filtration and washed with methylene chloride. The combined filtrate was washed twice with aqueous sodium hydrogen carbonate and once with saturated sodium chloride, dried over magnesium sulfate, and evaporated 25 to dryness, *in vacuo*. The solid residue was dissolved in dioxane (150 ml) and poured into water (200 ml) at 0°C. The title compound was isolated by filtration, washed with water, and dried over sicapent, *in vacuo*. Yield: 2.20 g (77%). An analytical sample was obtained by recrystallisation from 2-30 propanol. M.p. 174-175.5°C. Analysis for C₂₂H₂₃N₄O₇F₅, found(- calc.): C: 48.22(48.01); H: 4.64(4.21); N: 9.67(10.18). ¹H-NMR (250 MHz, CDCl₃): Due to the limited rotation around the secondary amide bond several of the signals were doubled in the ratio 6:1 (indicated in the list by mj. for major and mi. for minor). 7.01 ppm (s, mi., ArH); 6.99 ppm (s, mj., ArH); 5.27 ppm (unres. t, BocNH); 4.67 ppm (s, mj., T-CH₂-CO-); 4.60 ppm (s, mi., T-CH₂-CO-); 4.45 ppm (s, mj., CONRCH₂CO₂Pfp); 4.42

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ppm (s, m., CONRCH₂CO₂Pfp); 3.64 ppm (t, 2H, BocNHCH₂CH₂-); 3.87 ppm ("q", 2H, BocNHCH₂CH₂-); 1.44(s, 9H, t-Bu). FAB-MS: 551 (10; M+1); 495 (10; M+1-tBu); 451 (80; -Boc).

5 EXAMPLE 7

N⁴-Benzylloxycarbonyl cytosine (9)

Over a period of about 1 h, benzylloxycarbonyl chloride (52 ml; 0.36 mol) was added dropwise to a suspension of cytosine (8, 20.0 g; 0.18 mol) in dry pyridine (1000 ml) at 0°C under nitrogen in oven-dried equipment. The solution then was stirred overnight, after which the pyridine suspension was evaporated to dryness, *in vacuo*. Water (200 ml) and 4 N hydrochloric acid were added to reach pH ~1. The resulting white precipitate was filtered off, washed with water and partially dried by air suction. The still-wet precipitate was boiled with absolute ethanol (500 ml) for 10 min, cooled to 0°C, filtered, washed thoroughly with ether, and dried, *in vacuo*. Yield 24.7 g (54%). M.p.>250°C. Anal. for C₁₂H₁₇N₃O₃. Found(calc.); C: 58.59(58.77); H: 4.55(4.52); N: 17.17(17.13). No NMR spectra were recorded since it was not possible to get the product dissolved.

EXAMPLE 8

N⁴-Benzylloxycarbonyl-N¹-carboxymethyl cytosine (10)

In a three necked round bottomed flask equipped with mechanical stirring and nitrogen coverage was placed methyl bromacetate (7.82 ml; 82.6 mmol) and a suspension of N⁴-benzylloxycarbonyl-cytosine (9, 21.0 g; 82.6 mmol) and potassium carbonate (11.4 g; 82.6 mmol) in dry DMF (900 ml). The mixture was stirred vigorously overnight, filtered, and evaporated to dryness, *in vacuo*. Water (300 ml) and 4 N hydrochloric acid (10 ml) were added, the mixture was stirred for 15 minutes at 0°C, filtered, and washed with water (2 x 75 ml). The isolated precipitate was treated with water (120 ml), 2N sodium hydroxide (60 ml), stirred for 30 min, filtered, cooled to 0°C, and 4 N hydrochloric acid (35 ml) was added. The title compound was isolated by filtration, washed

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thoroughly with water, recrystallized from methanol (1000 ml) and washed thoroughly with ether. This afforded 7.70 g (31%) of pure compound. The mother liquor from the recrystallization was reduced to a volume of 200 ml and cooled to 0°C.

5 This afforded an additional 2.30 g of a material that was pure by tlc but had a reddish color. M.p. 266-274°C. Anal. for C₁₄H₁₃N₃O₅. Found(calc.); C: 55.41(55.45); H: 4.23(4.32); N: 14.04(13.86). ¹H-NMR (90 MHz; DMSO-d₆): 8.02 ppm (d,J=7.32Hz,1H,H-6); 7.39 (s,5H,Ph); 7.01 (d,J=7.32Hz,1H,H-5); 10 5.19 (s,2H,PhCH₂-); 4.52 (s,2H).

EXAMPLE 9

N⁴-Benzylloxycarbonyl-N¹-carboxymethyl-cytosine penta-fluorophenyl ester (11)

15 N⁴-Benzylloxycarbonyl-N¹-carboxymethyl-cytosine (10, 4.00 g; 13.2 mmol) and pentafluorophenol (2.67 g; 14.5 mmol) were mixed with DMF (70 ml), cooled to 0°C with ice-water, and DCC (3.27 g; 15.8 mmol) was added. The ice bath was removed after 3 min and the mixture was stirred for 3 h at room 20 temperature. The precipitated DCU was removed by filtration, washed with DMF, and the filtrate was evaporated to dryness, in vacuo (0.2 mmHg). The solid residue was treated with methylene chloride (250 ml), stirred vigorously for 15 min, filtered, washed twice with diluted sodium hydrogen carbonate 25 and once with saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness, in vacuo. The solid residue was recrystallized from 2-propanol (150 ml) and the crystals were washed thoroughly with ether. Yield 3.40 g (55%). M.p. 241-245°C. Anal. for C₂₀H₁₂N₃F₅O₅. Found(calc.); C: 51.56(51.18); H: 2.77(2.58); N: 9.24(8.95). ¹H-NMR (90 MHz; CDCl₃): 7.66 ppm (d,J=7.63Hz,1H,H-6); 7.37 (s,5H,Ph); 7.31 (d,J=7.63Hz,1H,H-5); 5.21 (s,2H,PhCH₂-); 4.97 (s,2H,NCH₂-). FAB-MS: 470 (M+1)

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EXAMPLE 10

N⁴-Benzylloxycarbonyl-1-Boc-aeg-cytosine (12)

To a solution of (N-Boc-2-aminoethyl)glycine (2) in DMF, prepared as described above, was added triethyl amine 5 (7.00 ml; 50.8 mmol) and N⁴-benzylloxycarbonyl-N¹-carboxymethyl-cytosine pentafluorophenyl ester (11, 2.70 g; 5.75 mmol). After stirring the solution for 1 h at room temperature, methylene chloride (150 ml), saturated sodium chloride (250 ml), and 4 N hydrochloric acid to pH ~1 were 10 added. The organic layer was separated and washed twice with saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness, *in vacuo*, first with a water aspirator and then with an oil pump. The oily residue was treated with water (25 ml) and was again evaporated to dryness, *in vacuo*. 15 This procedure then was repeated. The oily residue (2.80 g) was then dissolved in methylene chloride (100 ml), petroleum ether (250 ml) was added, and the mixture was stirred overnight. The title compound was isolated by filtration and washed with petroleum ether. Tlc (system 1) indicated 20 substantial quantities of pentafluorophenol, but no attempt was made to remove it. Yield: 1.72 g (59%). M.p. 156°C(decomp.). ¹H-NMR (250 MHz, CDCl₃): Due to the limited rotation around the secondary amide bond several of the signals were doubled in the ratio 2:1,(indicated in the list 25 by mj. for major and mi. for minor). 7.88 ppm (dd,1H,H-6); 7.39 (m,5H,Ph); 7.00 (dd,1H,H-5); 6.92 (b,1H,BocNH); 6.74 (b,1H,ZNH)-?; 5.19 (s,2H,Ph-CH₃); 4.81 ppm (s, mj., Cyt-CH₂-CO-); 4.62 ppm (s, mi., Cyt-CH₂-CO-); 4.23 (s, mi., CONRCH₂CO₂H); 3.98 ppm (s, mj., CONRCH₂CO₂H); 3.42-3.02 (unres. 30 m, -CH₂CH₂- and water); 1.37 (s,9H,tBu). FAB-MS: 504 (M+1); 448 (M+1-tBu).

EXAMPLE 11

N⁴-Benzylloxycarbonyl-1-B c-aeg-cytosine pentafluorophenyl 35 ester (13)

N⁴-Benzylloxycarbonyl-1-Boc-aeg-cytosine (12, 1.50 g; 2.98 mmol) and pentafluorophenol (548 mg; 2.98 mmol) was

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dissolved in DMF (10 ml) Methylene chloride (10 ml) was added, the reaction mixture was cooled to 0°C in an ice bath, and DCC (676 mg; 3.28 mmol) was added. The ice bath was removed after 3 min and the mixture was stirred for 3 h at ambient 5 temperature. The precipitate was isolated by filtration and washed once with methylene chloride. The precipitate was dissolved in boiling dioxane (150 ml) and the solution was cooled to 15°C, whereby DCU precipitated. The DCU was removed by filtration and the resulting filtrate was poured into water 10 (250 ml) at 0°C. The title compound was isolated by filtration, was washed with water, and dried over sicapent, *in vacuo*. Yield 1.30 g (65%). Analysis for C₂₉H₂₈N₅O₈F₅. Found(calc.); C: 52.63(52.02); H: 4.41(4.22); N: 10.55(10.46). ¹H-NMR (250 MHz; DMSO-d₆): showed essentially the spectrum of 15 the above acid, most probably due to hydrolysis of the ester. FAB-MS: 670 (M+1); 614 (M+1-tBu)

EXAMPLE 12

4-Chlorocarboxy-9-chloroacridine

20 4-Carboxyacridone (6.25 g; 26.1 mmol), thionyl chloride (25 ml), and 4 drops af DMF were heated gently under a flow of nitrogen until all solid material had dissolved. The solution then was refluxed for 40 min. The solution was cooled and excess thionyl chloride was removed *in vacuo*. The 25 last traces of thionyl chloride were removed by coevaporation with dry benzene (dried over Na-Pb) twice. The remaining yellow powder was used directly in the next reaction.

EXAMPLE 13

30 4-(5-Methoxycarbonylpentylamidocarbonyl)-9-chloroacridine

Methyl 6-aminohexanoate hydrochloride (4.70 g; 25.9 mmol) was dissolved in methylene chloride (90 ml), cooled to 0°C, triethyl amine (15 ml) was added, and the resulting solution then was immediately added to the acid chloride from 35 above. The roundbottomed flask containingthe acid chloride was cooled to 0°C in an ice bath. The mixture was stirred vigorously for 30 min at 0°C and 3 h at room temperature. The

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resulting mixture was filtered to remove the remaining solids, which were washed with methylene chloride (20 ml). The red-brown methylene chloride filtrate was subsequently washed twice with saturated sodium hydrogen carbonate, once with 5 saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness, in vacuo. To the resulting oily substance was added dry benzene (35 ml) and ligroin (60-80°C, dried over Na-Pb). The mixture was heated to reflux. Activated carbon and celite were added and mixture was 10 refluxed for 3 min. After filtration, the title compound crystallised upon cooling with magnetic stirring. It was isolated by filtration and washed with petroleum ether. The product was stored over solid potassium hydroxide. Yield 5.0 g (50%).

15

EXAMPLE 14

4-(5-Methoxycarbonylpentyl)amidocarbonyl-9-[6'-(4''-nitrobenzamido)hexylamino]-aminoacridine

4-(5-Methoxycarbonylpentylamidocarbonyl)-9-20 chloroacridine (1.30 g; 3.38 mmol) and phenol (5 g) were heated to 80°C for 30 min under a flow of nitrogen, after which 6-(4'-nitrobenzamido)-1-hexylamine (897 mg; 3.38 mmol) was added. The temperature raised to 120°C for 2 h. The reaction mixture was cooled and methylene chloride (80 ml) was 25 added. The resulting solution was washed three times with 2N sodium hydroxide (60 ml portions) and once with water, dried over magnesium sulfate, and evaporated to dryness, in vacuo. The resulting red oil (1.8 g) was dissolved in methylene chloride (40 ml), cooled to 0°C. Ether (120 ml) was added and 30 the resultant solution was stirred overnight. This results in a mixture of solid material and an oil. The solid was isolated by filtration. The solid and the oil were re-dissolved in methylene chloride (80 ml) and added dropwise to cold ether (150 ml). After 20 minutes of stirring, the title 35 compound was isolated by filtration in the form of orange crystals. The product was washed with ether and dried in

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vacuo over potassium hydroxide. Yield 1.60 g (77%). M.p. 145-147°C.

EXAMPLE 15

5 4-(5-Carboxypentyl)amidocarbonyl-9-[6'-(4''-nitrobenzamido)hexylamino]-aminoacridine

4-(5-Methoxycarbonylpentyl)amidocarbonyl-9-[6'-(4''-nitrobenzamido)hexylamino]aminoacridine (503 mg; 0.82 mmol) was dissolved in DMF (30 ml), and 2 N sodium hydroxide (30 ml) 10 was added. After stirring for 15 min, 2 N hydrochloric acid (35 ml) and water (50 ml) were added at 0°C. After stirring for 30 min, the solution was decanted, leaving an oily substance which was dissolved in boiling methanol (150 ml), filtered and concentrated to 1/3 volume. To the methanol 15 solution were added ether (125 ml) and 5-6 drops of HCl in ethanol. The solution was decanted after 1 h of stirring at 0°C. The oily substance was redissolved in methanol (25 ml) and precipitated with ether (150 ml). The title compound was isolated as yellow crystals after stirring overnight. Yield 20 417 mg (80%). M.p. 173°C (decomp.).

EXAMPLE 16

(a) 25 4-(5-pentafluorophenoxy carbonylpentyl)-amidocarbonyl-9-[6'-(4''-nitrobenzamido)-hexylamino]-aminoacridine(Acr¹O_{pfp})

The acid from above (300 mg; 0.480 mmol) was dissolved in DMF (2 ml) and methylene chloride (8 ml) was added. Pentafluorophenol (97 mg; 0.53 mmol), transferred with 2 x 2 ml of the methylene chloride, was added. The resulting 30 solution was cooled to 0°C after which DCC (124 mg; 0.60 mmol) was subsequently added. The ice bath was removed after 5 minutes and the mixture was left with stirring overnight. The precipitated DCU was removed by centrifugation and the centrifugate was evaporated to dryness, in vacuo, first by a 35 water aspirator and then by an oil pump. The residue was dissolved in methylene chloride (20 ml), filtered, and evaporated to dryness, in vacuo. The residue was again dissolved

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in methylene chloride and petroleum ether (150 ml). A 1 ml portion of 5M HCl in ether was added. The solvent was removed by decanting after 30 min of stirring at 0°C. The residual oily substance was dissolved in methylene chloride (100 ml).
5 Petroleum ether (150 ml) was added and the mixture was left with stirring overnight. The next day the yellow precipitated crystalline material was isolated by filtration and was washed with copious amounts of petroleum ether. Yield, after drying, 300 mg (78%). M.p. 97.5°C (decomp.) All samples showed
10 satisfactory elemental analysis, ¹H- and ¹³C-NMR and mass spectra.

(b) Experimental for the synthesis of PNA compounds,
cf. Figure 8

Materials: Boc-Lys (ClZ), benzhydrylamine-copoly-
15 (styrene-1%-divinylbenzene) resin (BHA resin), and p-methylbenzhydrylamine-copoly(styrene-1%-divinylbenzene)resin (MBHA resin) were purchased from Peninsula Laboratories. Other reagents and solvents were: Biograde trifluoroacetic acid from Halocarbon Products; diisopropylethylamine (99%; was
20 not further distilled) and N-acetylimidazole (98%) from Aldrich; H₂O was distilled twice; anhydrous HF from Union Carbide; synthesis grade N,N-dimethylformamide and analytical grade methylene chloride (was not further distilled) from Merck; HPLC grade acetonitrile from Lab-Scan; purum grade
25 anisole, N,N'-dicyclohexylcarbodiimide, diisopropylcarbodiimide, puriss. grade 2,2,2-trifluoroethanol from Fluka and trifluoromethanesulfonic acid from Flourad.

(b) General Methods and Remarks

Except where otherwise stated, the following applies.

30 The PNA compounds were synthesized by the stepwise solid-phase approach (Merrifield, J. Am. Chem. Soc., 1963, 85, 2149) employing conventional peptide chemistry utilizing the TFA-labile tert-butyloxycarbonyl (Boc) group for "temporary" N-protection (Merrifield, J. Am. Chem. Soc., 1964, 86, 304) and
35 the more acid-stable benzyloxycarbonyl (Z) and 2-chlorobenzyloxycarbonyl (ClZ) groups for "permanent" side chain protection. To obtain C-terminal amides, the PNAs were assembled onto the HF-labile BHA or MBHA resins (the MBHA

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resin has increased susceptibility to the final HF cleavage relative to the unsubstituted BHA resin (Matsueda, et al., *Peptides*, 1981, 2, 45). All reactions (except HF reactions) were carried out in manually operated standard solid-phase 5 reaction vessels fitted with a coarse glass frit (Merrifield, et al., *Biochemistry*, 1982, 21, 5020). The quantitative ninhydrin reaction (Kaiser test), originally developed by Merrifield and co-workers (Sarin, et al., *Anal. Biochem.*, 1981, 117, 147) for peptides containing "normal" amino acids, 10 was successfully applied (see Table I - III) using the "normally" employed effective extinction coefficient $\epsilon = 15000 \text{ M}^{-1}\text{cm}^{-1}$ for all residues to determine the completeness of the individual couplings as well as to measure the number of growing peptide chains. The theoretical substitution 15 S_{n-1} upon coupling of residue number n (assuming both complete deprotection and coupling as well as neither chain termination nor loss of PNA chains during the synthetic cycle) is calculated from the equation:

$$S_n = S_{n-1} \times (1 + (S_{n-1} \times \Delta MW \times 10^{-3} \text{ mmol/mol}))^{-1}$$

20 where ΔMW is the gain in molecular weight ($[\Delta MW] = \text{g/mol}$) and S_{n-1} is the theoretical substitution upon coupling of the preceding residue n-1 ($[S] = \text{mmol/g}$). The estimated value (%) on the extent of an individual coupling is calculated relative to the measured substitution (unless S was not determined) and 25 include correction for the number of remaining free amino groups following the previous cycle. HF reactions were carried out in a Diaflon HF apparatus from Toho Kasei (Osaka, Japan). Vydac C₁₈ (5 μm , 0.46 x 25 cm and 5 μm , 1.0 x 25 cm) reverse-phase columns, respectively were used for analytical 30 and semi-preparative HPLC on an SP8000 instrument. Buffer A was 5 vol % acetonitrile in water containing 445 μl trifluoroacetic acid per liter, and buffer B was 60 vol % acetonitrile in water containing 390 μl trifluoroacetic acid per liter. The linear gradient was 0-100% of buffer B in 30 35 min, flow rates 1.2 ml/min (analytical) and 5 ml/min (semi-preparative). The eluents were monitored at 215 nm (analytical) and 230 nm (semi-preparative). Molecular weights

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of the PNAs were determined by ^{252}Cf plasma desorption time-of-flight mass spectrometry from the mean of the most abundant isotopes.

5 EXAMPLE 17

Solid-Phase Synthesis of Acr¹-[Taeg]₁₅-NH₂ and Shorter Derivatives

(a) Stepwise Assembly of Boc-[Taeg]₁₅-BHA Resin

The synthesis was initiated on 100 mg of preswollen and 10 neutralized BHA resin (determined by the quantitative ninhydrin reaction to contain 0.57 mmol NH₂/g) employing single couplings ("Synthetic Protocol 1") using 3.2 equivalents of BocTaeg-OPfp in about 33% DMF/CH₂Cl₂. The individual coupling reactions were carried out by shaking for 15 at least 12 h in a manually operated 6 ml standard solid-phase reaction vessel and unreacted amino groups were blocked by acetylation at selected stages of the synthesis. The progress of chain elongation was monitored at several stages by the quantitative ninhydrin reaction (see Table I). Portions of 20 protected Boc-[Taeg]₅-BHA, Boc-[Taeg]₁₀-BHA, and Boc-[Taeg]₁₅-BHA resins were taken out after assembling 5, 10, and 15 residues, respectively.

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	Synthetic Step	Residue Coupled	Substitution After Deprotection (mmol/g)		Remaining Free Amino Groups After ($\mu\text{mol/g}$)		Estimated Extent of Coupling (%)
			Measd	Theoretol	Single Coupling	Acetylation	
	"0"		0.57				
5	1	BocTaeg	ND	0.50	1.30		<99.7
	2	BocTaeg	ND	0.44	1.43		<99.9
	3	BocTaeg	0.29	0.39	3.33		99.3
	4	BocTaeg	0.27	0.35	13.30		96.3
	5	BocTaeg	0.26	0.32	8.33		>99.9
10	6	BocTaeg	ND	0.30	7.78		>99.9
	7	BocaTeg	ND	0.28	13.81	7.22	<97.8
	8	BocTaeg	ND	0.26	14.00		<99.9
	9	BocTaeg	ND	0.24	30.33		93.2
	10	BocTaeg	0.16	0.23	11.67	2.67	>99.9
15	11	BocTaeg	ND	0.21	4.58		>99.9
	12	BocTaeg	ND	0.20	5.87		<99.4
	13	BocTaeg	ND	0.19	1.67		>99.9
	14	BocTaeg	ND	0.18	14.02		<93.1
	15	BocTaeg	0.07	0.17	4.20	3.33	>99.9

20

(b) Synthesis of $\text{Acr}^1\text{-[Taeg]}_{15}\text{-BHA Resin}$

Following deprotection of the residual $\text{Boc-[\Taeg]}_{15}\text{-BHA}$ resin (estimated dry weight is about 30 mg; -0.002 mmol growing chains), the $\text{H-[\Taeg]}_{15}\text{-BHA}$ resin was reacted with 25 about 50 equivalents (80 mg; 0.11 mmol) of $\text{Acr}^1\text{-OPfp}$ in 1 ml of about 66% $\text{DMF}/\text{CH}_2\text{Cl}_2$ (i.e., a 0.11 M solution of the pentafluorophenylester) in a 3 ml solid-phase reaction vessel. As judged by a qualitative ninhydrin reaction, coupling of the acridine moiety was close to quantitative.

30

(c) Cleavage, Purification, and Identification of $\text{H-[\Taeg]}_5\text{-NH}_2$

A portion of protected $\text{Boc-[\Taeg]}_5\text{-BHA}$ resin was treated with 50% trifluoroacetic acid in methylene chloride to remove the N-terminal Boc group (which is a precursor of

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the potentially harmful tert-butyl cation) prior to the HF cleavage. Following neutralization and washing (performed in a way similar to those of steps 2-4 in "Synthetic Protocol 1"), and drying for 2 h in vacuum, the resulting 67.1 mg (dry weight) of H-[Taeg]₅-BHA resin was cleaved with 5 ml of HF:anisole (9:1, v/v) stirring at 0°C for 60 min. After removal of HF, the residue was stirred with dry diethyl ether (4 x 15 ml, 15 min each) to remove anisole, filtered under gravity through a fritted glass funnel, and dried. The PNA 10 was then extracted into a 60 ml (4 x 15 ml, stirring 15 min each) 10% aqueous acetic acid solution. Aliquots of this solution were analyzed by analytical reverse-phase HPLC to establish the purity of the crude PNA. The main peak at 13.0 min accounted for about 93% of the total absorbance. The 15 remaining solution was frozen and lyophilized to afford about 22.9 mg of crude material. Finally, 19.0 mg of the crude product was purified from five batches, each containing 3.8 mg in 1 ml of H₂O. The main peak was collected by use of a semi-preparative reverse-phase column. Acetonitrile was 20 removed on a speed vac and the residual solution was frozen (dry ice) and subsequently lyophilized to give 13.1 mg of >99% pure H-[Taeg]₅-NH₂. The PNA molecule readily dissolved in water and had the correct molecular weight based on mass spectral determination. For (M+H)⁺ the calculated m/z value 25 was 1349.3 and the measured m/z value was 1347.8.

(d) Cleavage, Purification, and Identification of H-[Taeg]₁₀-NH₂

A portion of protected Boc-[Taeg]₁₀-BHA resin was treated as described in section (c) to yield 11.0 mg of crude 30 material upon HF cleavage of 18.9 mg dry H-[Taeg]₁₀-BHA resin. The main peak at 15.5 min accounted for about 53% of the total absorbance. About 1 mg of the crude product was purified repeatedly (for reasons described below) to give approximately 0.1 mg of at least 80% but presumably >99% pure H-[Taeg]₁₀-NH₂. 35 A rather broad tail eluting after the target peak and accounting for about 20% of the total absorbance could not be removed (only slightly reduced) upon the repeated

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purification. Judged by the mass spectrum, which only confirms the presence of the correct molecular weight H-[Taeg]₁₀-NH₂, the tail phenomenon is ascribed to more or less well-defined aggregational/conformational states of the target molecule. Therefore, the crude product is likely to contain more than the above-mentioned 53% of the target molecule. H-[Taeg]₁₀-NH₂ is readily dissolved in water. For (M+H)⁺ the calculated m/z value was 2679.6 and the measured m/z value was 2681.5.

10 (e) Cleavage, Purification, and Identification of H-[Taeg]₁₅-NH₂.

A portion of protected Boc-[Taeg]₁₅-BHA resin was treated as described in section (c) to yield 3.2 mg of crude material upon HF cleavage of 13.9 mg dry H-[Taeg]₁₅-BHA resin. The main peak at 22.6 min was located in a broad bulge accounting for about 60% of the total absorbance (Fig. 12a). Again (see the preceding section), this bulge is ascribed to aggregational/conformational states of the target molecule H-[Taeg]₁₅-NH₂ since mass spectral analysis of the collected "bulge" did not significantly reveal the presence of other molecules. All of the crude product was purified collecting the "bulge" to give approximately 2.8 mg material. For (M+Na)⁺ the calculated m/z value was 4033.9 and the measured m/z value was 4032.9.

25 (f) Cleavage, Purification, and Identification of Acr¹-[Taeg]₁₅-NH₂.

A portion of protected Acr¹-[Taeg]₁₅-BHA resin was treated as described in section (b) to yield 14.3 mg of crude material upon HF cleavage of 29.7 mg dry Acr¹-[Taeg]₁₅-BHA resin. Taken together, the main peak at 23.7 min and a "dimer" (see below) at 29.2 min accounted for about 40% of the total absorbance (Fig. 12b). The crude product was purified repeatedly to give approximately 1 mg of presumably >99% pure Acr¹-[Taeg]₁₅-NH₂ "contaminated" with self-aggregated molecules eluting at 27.4 min, 29.2 min, and finally as a huge broad bulge eluting with 100% buffer B (Fig. 12c). This interpretation is in agreement with the observation that those

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peaks grow upon standing (for hours) in aqueous acetic acid solution, and finally precipitate out quantitatively. For $(M+H)^+$ the calculated m/z value was 4593.6 and the measured m/z value was 4588.7.

5 (g) Synthetic Protocol 1

(1) Boc-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 3 ml, 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 3 ml, 6 x 1 min; (3) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 3 ml, 3 x 2 min; (4) washing with CH₂Cl₂, 3 ml, 6 x 1 min, and drain 10 for 1 min; (5) 2-5 mg sample of PNA-resin may be taken out and dried thoroughly for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 3.2 equiv. (0.18 mmol; 100 mg) BocTaeg-OPfp dissolved in 1 ml CH₂Cl₂ followed by addition of 0.5 ml DMF (final concentration of 15 pentafluorophenylester ~0.12 M); the coupling reaction was allowed to proceed for a total of 12-24 h shaking at room temperature; (7) washing with DMF, 3 ml, 1 x 2 min; (8) washing with CH₂Cl₂, 3 ml, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 3 ml, 2 x 2 min; (10) washing with 20 CH₂Cl₂, 3 ml, 6 x 1 min; (11) 2-5 mg sample of protected PNA-resin is taken out for a rapid qualitative ninhydrin test and further 2-5 mg is dried thoroughly for a quantitative ninhydrin analysis to determine the extent of coupling (after cycles 7, 10, and 15 unreacted amino groups were blocked by 25 acetylation with N-acetylimidazol in methylene chloride).

EXAMPLE 18

Solid-Phase Synthesis of Acr¹-[Taeg]₁₅-Lys-NH₂ and Shorter Derivatives

30 (a) Stepwise Assembly of Boc-[Taeg]₁₅-Lys(ClZ)-BHA Resin

The synthesis was initiated by a quantitative loading (standard DCC *in situ* coupling in neat CH₂Cl₂) of Boc-Lys(ClZ) onto 100 mg of preswollen and neutralized BHA resin (0.57 mmol 35 NH₂/g). Further extension of the protected PNA chain employed single couplings ("Synthetic Protocol 2") for cycles 1 to 5 and cycles 10 to 15 using 3.2 equivalents of BocTaeg-

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OPfp in about 33% DMF/CH₂Cl₂. Cycles 5 to 10 employed an extra straight DCC (i.e., *in situ*) coupling of the free acid Boc-Taeg-OH in about 33% DMF/CH₂Cl₂. All coupling reactions were carried out by shaking for at least 12 h in a manually
5 operated 6 ml standard solid-phase reaction vessel. Unreacted amino groups were blocked by acetylation at the same stages of the synthesis, as was done in Example 17. Portions of protected Boc-[Taeg]₅-Lys(ClZ)-BHA and Boc-[Taeg]₁₀-Lys(ClZ)-BHA resins were taken out after assembling 5 and 10 PNA
10 residues, respectively. As judged by the analytical HPLC chromatogram of the crude cleavage product from the Boc-[Taeg]₁₀-Lys(ClZ)-BHA resin (see section (e)), an additional "free acid" coupling of PNA residues 5 to 10 gave no significant improvement of the synthetic yield as compared to
15 the throughout single-coupled residues in Example 17.

25 (b) Synthesis of Acr¹-[Taeg]₁₀-Lys(ClZ)-BHA Resin

Following deprotection of a portion of Boc-[Taeg]₁₀-Lys(ClZ)-BHA resin (estimated dry weight is about 90 mg; ~ 0.01 mmol growing chains), the H-[Taeg]₁₅-BHA resin was
20 reacted with about 20 equivalents (141 mg; 0.19 mmol) of Acr¹-OPfp in 1 ml of about 66% DMF/CH₂Cl₂ in a 3 ml solid-phase reaction vessel. As judged by a qualitative ninhydrin reaction, coupling of the acridine moiety was close to quantitative.

25 (c) Synthesis of Acr¹-[Taeg]₁₅-Lys(ClZ)-BHA Resin

Following deprotection of the residual Boc-[Taeg]₁₅-Lys(ClZ)-BHA resin (estimated dry weight about 70 mg; ~ 0.005 mmol growing chains), the H-[Taeg]₁₅-Lys(ClZ)-BHA resin was
30 reacted with about 25 equivalents (91 mg; 0.12 mmol) of Acr¹-OPfp in 1 ml of about 66% DMF/CH₂Cl₂ in a 3 ml solid-phase reaction vessel. As judged by a qualitative ninhydrin reaction, coupling of the acridine moiety was close to quantitative.

35 (d) Cleavage, Purification, and Identification of H-[Taeg]₅-Lys-NH₂

A portion of protected Boc-[Taeg]₅-Lys(ClZ)-BHA resin was treated as described in Example 17c to yield 8.9 mg of

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crude material upon HF cleavage of 19.0 mg dry H-[Taeg]₅-Lys(ClZ)-BHA resin. The main peak at 12.2 min (eluted at 14.2 min if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) accounted for about 90% of the total absorbance. About 2.2 mg of the crude product was purified to give approximately 1.5 mg of 99% pure H-[Taeg]₅-Lys-NH₂.

(e) Cleavage, Purification, and Identification of H-[Taeg]₁₀-Lys-NH₂

10 A portion of protected Boc-[Taeg]₁₀-Lys(ClZ)-BHA resin was treated as described in Example 17c to yield 1.7 mg of crude material upon HF cleavage of 7.0 mg dry H-[Taeg]₁₀-Lys(ClZ)-BHA resin. The main peak at 15.1 min (eluted at 17.0 min if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) accounted for about 50% of the total absorbance. About 1.2 mg of the crude product was purified to give approximately 0.2 mg of >95% pure H-[Taeg]₁₀-Lys-NH₂. Figure 13a. For (M+H)⁺ the calculated m/z value was 2807.8 and the measured m/z value was 2808.2.

20 (f) Cleavage, Purification, and Identification of Acr¹-[Taeg]₁₀-Lys-NH₂

99.1 mg protected Acr¹-[Taeg]₁₀-Lys(ClZ)-BHA resin (dry weight) was cleaved as described in Example 17c to yield 42.2 mg of crude material. The main peak at 25.3 min (eluted at 23.5 min if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) accounted for about 45% of the total absorbance. An 8.87 mg portion of the crude product was purified to give approximately 5.3 mg of >97% pure H-[Taeg]₁₀-Lys-NH₂. For (M+H)⁺ the calculated m/z value was 2850.8 and the measured m/z value was 2849.8.

(g) Cleavage and Purification of Acr¹-[Taeg]₁₅-Lys-NH₂

A 78.7 mg portion of protected Acr¹-[Taeg]₁₅-Lys(ClZ)-BHA resin (dry weight) was cleaved as described in Example I section (c) to yield 34.8 mg of crude material. The main peak at 23.5 min (about the same elution time if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) and a "dimer" at 28.2 min accounted for about 35%

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of the total absorbance. About 4.5 mg of the crude product was purified to give approximately 1.6 mg of presumably >95% pure H-[Taeg]₁₀-Lys-NH₂. This compound could not be free of the "dimer" peak, which grew upon standing in aqueous acetic acid solution.

(h) Synthetic Protocol 2

(1) Boc-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 3 ml, 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 3 ml, 6 x 1 min; (3) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 3 ml, 10 3 x 2 min; (4) washing with CH₂Cl₂, 3 ml, 6 x 1 min, and drain for 1 min; (5) 2-5 mg sample of PNA-resin can be taken out and dried thoroughly for a qualitative ninhydrin analysis; (6) for cycles 1 to 5 and cycles 10 to 15 the coupling reaction was carried out by addition of 3.2 equiv. (0.18 mmol; 100 mg) 15 BocTaeg-OPfp dissolved in 1 ml CH₂Cl₂ followed by addition of 0.5 ml DMF (final concentration of pentafluorophenylester ~ 0.12 M); the coupling reaction was allowed to proceed for a total of 12-24 h with shaking; cycles 5 to 10 employed an additional 0.12 M DCC coupling of 0.12 M BocTaeg-OH in 1.5 ml 20 DMF/CH₂Cl₂ (1:2, v/v); (7) washing with DMF, 3 ml, 1 x 2 min; (8) washing with CH₂Cl₂, 3 ml, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 3 ml, 2 x 2 min; (10) washing with CH₂Cl₂, 3 ml, 6 x 1 min; (11) 2-5 mg sample of protected 25 PNA-resin is taken out for a qualitative ninhydrin test (after cycles 7, 10, and 15 unreacted amino groups were blocked by acetylation with N-acetylimidazol in methylene chloride).

EXAMPLE 19

Improved Solid-Phase Synthesis of H-[Taeg]₁₀-Lys-NH₂

30 The protected PNA was assembled onto an MBHA resin, using approximately half the loading of the BHA resin used in the previous examples. Furthermore, all cycles except one was followed by acetylation of uncoupled amino groups. The following describes the synthesis in full detail:

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(a) Preparation of Boc-Lys(ClZ)-NH-CH(p-CH₃-C₆H₄)-C₆H₄ Resin (MBHA Resin) with an Initial Substitution of 0.3 mmol/g

The desired substitution of Boc-Lys(ClZ)-MBHA resin was 5 0.25 - 0.30 mmol/g. In order to get this value, 1.5 mmol of Boc-Lys(ClZ) was coupled to 5.0 g of neutralized and preswollen MBHA resin (determined by the quantitative ninhydrin reaction to contain 0.64 mmol NH₂/g) using a single "in situ" coupling (1.5 mmol of DCC) in 60 ml of CH₂Cl₂. The 10 reaction was carried out by shaking for 3 h in a manually operated, 225 ml, standard, solid-phase reaction vessel. Unreacted amino groups were then blocked by acetylation with a mixture of acetic anhydride/pyridine/CH₂Cl₂ (1:1:2, v/v/v) for 18 h. A quantitative ninhydrin reaction on the 15 neutralized resin showed that only 0.00093 mmol/g free amine remained (see Table I), i.e. 0.15% of the original amino groups. The degree of substitution was estimated by deprotection and ninhydrin analysis, and was found to be 0.32 mmol/g for the neutralized H-Lys(ClZ)-MBHA resin. This 20 compares well with the maximum value of 0.28 mmol/g for a quantitative coupling of 0.30 mmol Boc-Lys(ClZ)/g resin (see Table II).

(b) Stepwise Assembly of Boc-[Taeg]₃-Lys(ClZ)-MBHA Resin

25 The entire batch of H-Lys(ClZ)-MBHA resin prepared in section (a) was used directly (in the same reaction vessel) to assemble Boc-[Taeg]₃-Lys(ClZ)-MBHA resin by single couplings ("Synthetic Protocol 3") utilizing 2.5 equivalents of BocTaeg-OPfp in neat CH₂Cl₂. The quantitative ninhydrin 30 reaction was applied throughout the synthesis (see Table II).

(c) Stepwise Assembly of Boc-[Taeg]₈-Lys(ClZ)-MBHA Resin

About 4.5 g of wet Boc-[Taeg]₃-Lys(ClZ)-MBHA resin (~0.36 mmol growing chains; taken out of totally ~ 19 g wet 35 resin prepared in section (b)) was placed in a 55 ml SPPS reaction vessel. Boc-[Taeg]₈-Lys(ClZ)-MBHA resin was assembled by single couplings ("Synthetic Protocol 4")

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utilizing 2.5 equivalents of BocTaeg-OPfp in about 30% DMF/CH₂Cl₂. The progress of the synthesis was monitored at all stages by the quantitative ninhydrin reaction (see Table II).

5 (d) Stepwise Assembly of Boc-[Taeg]₁₀-Lys(ClZ)-MBHA Resin

About 1 g of wet Boc-[Taeg]₈-Lys(ClZ)-MBHA resin (~0.09 mmol growing chains; taken out of totally ~4 g wet resin prepared in section (c)) was placed in a 20 ml SPPS reaction 10 vessel. Boc-[Taeg]₁₀-Lys(ClZ)-MBHA resin was assembled by the single-coupling protocol employed in the preceding section utilizing 2.5 equivalents of BocTaeg-OPfp in about 30% DMF/CH₂Cl₂. The reaction volume was 3 ml (vigorous shaking). The synthesis was monitored by the quantitative ninhydrin 15 reaction (see Table II).

	Synthetic Step	Residue Coupled	Substitution After Deprotection (mmol/g)		Remaining Free Amino Groups After (μ mol/g)		Estimated Extent of Coupling (%)
			Measd	Theoret	Single Coupling	Acetylation	
20	"0"	BocLys(ClZ)	0.32	0.28		0.93	
	1	BocTaeg	0.23	0.26	0.97	0.54	>99.9
	2	BocTaeg	0.21	0.24	0.92	0.46	99.8
	3	BocTaeg	0.19	0.23	1.00	0.57	99.7
	4	BocTaeg	0.18	0.21	1.85		99.3
25	5	BocTaeg	0.17	0.20	2.01	0.19	99.9
	6	BocTaeg	0.15	0.19	1.69	0.10	99.0
	7	BocTaeg	0.11	0.18	1.11	0.66	99.1
	8	BocTaeg	0.12	0.17	1.82	0.44	99.0
	9	BocTaeg	0.10	0.17	5.63	0.56	94.8
30	10	BocTaeg	0.11	0.16	1.54	0.67	99.1

(e) Synthesis of Ac-[Taeg]₁₀-Lys(ClZ)-MBHA Resin

Following deprotection of a portion of Boc-[Taeg]₁₀-Lys(ClZ)-MBHA resin (estimated dry weight is about 45 mg), the 35 resin was next acetylated quantitatively with a 2 ml mixture

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of acetic anhydride/pyridine/CH₂Cl₂ (1:1:2, v/v/v) for 2 h in a 3 ml solid-phase reaction vessel.

(f) Cleavage, Purification, and Identification of H-[Taeg]₁₀-Lys-NH₂

5 A portion of protected Boc-[Taeg]₁₀-Lys(ClZ)-BHA resin was treated as described in Example 17c to yield about 24 mg of crude material upon HF cleavage of 76 mg dry H-[Taeg]₅-Lys(ClZ)-BHA resin. The main peak at 15.2 min (which includes impurities such as deletion peptides and various byproducts) 10 accounted for about 78% of the total absorbance. The main peak also accounted for about 88% of the "main peak plus deletion peaks" absorbance, which is in good agreement with the overall estimated coupling yield of 90.1% obtained by summarizing the individual coupling yields in Table II. A 7.2 15 mg portion of the crude product was purified from two batches by use of a semi-preparative reverse-phase column, (collecting the main peak in a beaker cooled with dry ice/2-propanol). Each contained 3.6 mg in 1 ml of H₂O. The frozen solution was lyophilized directly (without prior removal of acetonitrile 20 on a speed vac) to give 4.2 mg of 82% pure H-[Taeg]₁₀-Lys-NH₂.

(g) Cleavage, Purification, and Identification of Ac-[Taeg]₁₀-Lys-NH₂

A 400.0 mg portion of protected Ac-[Taeg]₁₀-Lys(ClZ)-BHA resin (dry weight) was cleaved as described in Example 25 17c, except for the TFA treatment to yield 11.9 mg of crude material. The main peak at 15.8 min accounted for about 75% of the total absorbance. A 4.8 mg portion of the crude product was purified to give approximately 3.5 mg of >95% pure Ac-[Taeg]₁₀-Lys-NH₂. For (M+H)⁺ the calculated m/z value = 30 2849.8 and the measured m/z value = 2848.8.

(h) Synthetic Protocol 3.

(1) Boc-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 100 ml, 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 100 ml, 6 x 1 min; (3) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 35 100 ml, 3 x 2 min; (4) washing with CH₂Cl₂, 100ml, 6 x 1 min, and drain for 1 min; (5) 2-5 mg sample of PNA-resin is taken out and dried thoroughly for a quantitative ninhydrin analysis

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to determine the substitution; (6) addition of 2.5 equiv. (3.75 mmol; 2.064 g) BocTaeg-OPfp dissolved in 35 ml CH₂Cl₂ (final concentration of pentafluorophenylester ~0.1 M); the coupling reaction was allowed to proceed for a total of 20-24
5 h with shaking; (7) washing with DMF, 100 ml, 1 x 2 min (to remove precipitate of BocTaeg-OH); (8) washing with CH₂Cl₂, 100 ml, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 100 ml, 2 x 2 min; (10) washing with CH₂Cl₂, 100 ml, 6 x 1 min; (11) 2-5 mg sample of protected PNA-resin is taken
10 out for a rapid qualitative ninhydrin test and a further 2-5 mg is dried thoroughly for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted amino groups by acetylation with a 100 ml mixture of acetic anhydride / pyridine / CH₂Cl₂ (1:1:2, v/v/v) for 2
15 h; (13) washing with CH₂Cl₂, 100 ml, 6 x 1 min; (14) 2 x 2-5 mg samples of protected PNA-resin are taken out, neutralized with DIEA/CH₂Cl₂ (1: 19, v/v) and washed with CH₂Cl₂ for qualitative and quantitative ninhydrin analyses.

(i) Synthetic Protocol 4.

20 (1) Boc-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 25 ml, 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 25 ml, 6 x 1 min; (3) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 25 ml, 3 x 2 min; (4) washing with CH₂Cl₂, 25 ml, 6 x 1 min, and drain for 1 min; (5) 2-5 mg sample of PNA-resin is taken out
25 and dried thoroughly for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 2.5 equiv. (0.92 mmol; 0.506 g) BocTaeg-OPfp dissolved in 6 ml CH₂Cl₂ followed by addition of 3 ml DMF (final concentration of pentafluorophenylester ~0.1 M); the coupling reaction was
30 allowed to proceed for a total of 20-24 hrs with shaking; (7) washing with DMF, 25 ml, 1 x 2 min; (8) washing with CH₂Cl₂, 25 ml, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 25 ml, 2 x 2 min; (10) washing with CH₂Cl₂, 25 ml, 6 x 1 min; (11) 2-5 mg sample of protected PNA-resin is taken out
35 for a rapid qualitative ninhydrin test and a further 2-5 mg is dried thoroughly for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted

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amino groups by acetylation with a 25 ml mixture of acetic anhydride/pyridine/CH₂Cl₂ (1:1:2, v/v/v) for 2 h (except after the first cycle); (13) washing with CH₂Cl₂, 25 ml, 6 x 1 min; (14) 2 x 2-5 mg samples of protected PNA-resin are taken out, 5 neutralized with DIEA/CH₂Cl₂ (1: 19, v/v) and washed with CH₂Cl₂ for qualitative and quantitative ninhydrin analyses.

EXAMPLE 20**Solid-Phase Synthesis of H-[Taeg]₅-Caeg-[Taeg]₄-Lys-NH₂**10 (a) Stepwise Assembly of Boc-[Taeg]₅-C(z)aeg-[Taeg]₄-Lys(Clz)-MBHA Resin

About 2.5 g of wet Boc-[Taeg]₃-Lys(Clz)-MBHA resin (~ 1/6 of the total remaining about 16 g wet resin; ~0.75 g dry resin ~0.15 mmol growing chains) was placed in a 6 ml SPPS 15 reaction vessel. Boc-[Taeg]₅-Caeg-[Taeg]₄-Lys(Clz)-MBHA resin was assembled by double coupling of all Taeg-residues utilizing the usual 2.5 equivalents of BocTaeg-OPfp in 2.5 ml about 30% DMF/CH₂Cl₂, except that the first residue was single-coupled. Incorporation of the C(Z)aeg-residue was 20 accomplished by coupling with 2.0 equivalents of BocC(Z)aeg-OPfp in TFE/CH₂Cl₂ (1:2, v/v). The progress of the synthesis was monitored at all stages by the quantitative ninhydrin reaction (see Table III).

	Synthetic Step	Residue Coupled	Substitution After Deprotection (mmol/g)		Remaining Free Amino Groups After (μ mol/g)			Estimated Extent of Coupling
			Measd.	Theoret.	1st Coupl	2nd Coupl	Acetyl-ation	
	3		0.19	0.23	1.00		0.57	
	4	BocTaeg	0.17	0.21	4.88		97.3	97.3
30	5	BocC(Z)aeg	0.11	0.20	70.20	27.98	1.33	78.4 (46)
	6	BocTaeg	0.10	0.19	24.79	4.58	2.40	95.4 (75)
	7	BocTaeg	0.09	0.18	8.55	1.61	0.20	>99.9 (93)
	8	BocTaeg	0.08	0.17	6.53	0.80	0.45	99.0 (91)
	9	BocTaeg	0.07	0.16	9.26	3.66	0.61	94.8 (86)
35	10	BocTaeg	0.07	0.15	5.32	1.48	0.60	98.8 (93)

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(b) Cleavage, Purification, and Identification of H-[Taeg]₅-Caeg-[Taeg]₄-Lys-NH₂

A portion of protected Boc-[Taeg]₅-Caeg-[Taeg]₄-Lys(ClZ)-BHA resin was treated as described in Example I section (c) to yield about 14.4 mg of crude material upon HF cleavage of 66.9 mg dry H-[Taeg]₅-Caeg-[Taeg]₄-Lys(ClZ)-BHA resin. The main peak at 14.5 min accounted for >50% of the total absorbance. A 100.0 mg portion of the crude product was purified (8 batches; each dissolved in 1 ml H₂O) to give approximately 9.1 mg of 96% pure H-[Taeg]₅-Caeg-[Taeg]₄-Lys-NH₂ (Figure 13b). For (M+H)⁺ the calculated m/z value = 2793,8 and the measured m/z value = 2790,6.

EXAMPLE 21

15 Binding of Acr¹-(Taeg)₁₀-Lys-NH₂ to dA₁₀ (Figure 11a)

Acr¹-(Taeg)₁₀-Lys (100 ng) was incubated for 15 min at room temperature with 50 cps 5'-[³²P]-end-labelled oligonucleotide [d(GATCCA₁₀G)] in 20 µl TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 7.4). The sample was cooled in ice (15 min) and analyzed by gel electrophoresis in polyacrylamide (PAGE). To 10 µl of the sample was added 2 µl 50% glycerol, 5 TBE (TBE = 90 mM Tris-borate, 1 mM EDTA, pH 8.3), and the sample was analysed by PAGE (15% acrylamide, 0.5% bisacrylamide) in TBE buffer at 4°C. A 10 µl portion of the 25 sample was lyophilized and redissolved in 10 µl 80% formamide, 1 TBE, heated to 90°C (5 min), and analyzed by urea/PAGE (15% acrylamide, 0.5% bisacrylamide, 7 M urea) in TBE. [³²P]-containing DNA bands were visualized by autoradiography using intensifying screens and Agfa Curix RPI X-ray films exposed 30 at -80°C for 2 h.

Oligonucleotides were synthesized on a Biosearch 7500 DNA synthesizer, labelled with γ [³²P]-ATP (Amersham, 5000 Ci/mmol) and polynucleotide kinase, and purified by PAGE using standard techniques (Maniatis et al. (1986): A laboratory manual, Cold Spring Harbor Laboratories).

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EXAMPLE 22

Formation of strand displacement complex

A dA₁₀-dT₁₀ target sequence contained within a plasmid DNA sequence was constructed by cloning of two oligonucleotides (d(GATCCA₁₀G) + d(GATCC^TT₁₀G)) into the BamHI restriction enzyme site of pUC19 using the Escherichia coli JM101 strain by standard techniques (Maniatis et al., 1986). The desired plasmid (designated pT10) was isolated from one of the resulting clones and purified by the alkaline extraction procedure and CsCl centrifugation (Maniatis et al., 1986). A 3'-[³²P]-end-labelled DNA fragment of 248 bp containing the dA₁₀/dT₁₀ target sequence was obtained by cleaving the pT10 DNA with restriction enzymes EcoRI and PvuII, labelling of the cleaved DNA with α [³²P]-dATP (4000 Ci/mmol, Amersham) using the Klenow fragment of E. coli DNA polymerase (Boehringer Mannheim), and purifying the 248 bp DNA fragment by PAGE (5% acrylamide, 0.06% bisacrylamide, TBE buffer). This DNA fragment was obtained with [³²P]-end-labelling at the 5'-end by treating the EcoRI-cleaved pT10 plasmid with bacterial alkaline phosphatase (Boehringer Mannheim), purifying the plasmid DNA by gel electrophoresis in low melting agarose, and labelling with γ [³²P] ATP and polynucleotide kinase. Following treatment with PvuII, the 248 bp DNA fragment was purified as above.

The complex between Acr¹-(Taeg)₁₀-Lys-NH₂ and the 248 bp DNA fragment was formed by incubating 50 ng of Acr¹-(Taeg)₁₀-Lys-NH₂ with 500 cps ³²P-labelled 248 bp fragment and 0.5 μ g calf thymus DNA in 100 μ l buffer for 60 min at 37°C.

EXAMPLE 23

Probing of strand displacement complex with:

(a) Staphylococcus nuclease (Figure 12b)

The strand displacement complex was formed in 25 mM Tris-HCl, 1 mM MgCl₂, 0.1 mM CaCl₂, pH 7.4 as described above. The complex was treated with Staphylococcus nuclease (Boehringer Mannheim) at 750 U/ml for 5 min at 20°C and the reaction was stopped by addition of EDTA to 25 mM. The DNA

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was precipitated with 2 vols. of ethanol, 2% potassium acetate redissolved in 80% formamide, TBE, heated to 90°C (5 min), and analyzed by high resolution PAGE (10% acrylamide, 0.3% bisacrylamide, 7 M urea) and autoradiography.

5 (b) Affinity photocleavage (Figure 12a + 12b)

The complex was formed in TE buffer. A sample contained in an Eppendorf tube was irradiated from above at 300 nm (Philips TL 20 W/12 fluorescent light tube, $24 \text{ Jm}^{-2} \text{s}^{-1}$) for 30 min. The DNA was precipitated as above, taken up in 1 M 10 piperidine, and heated to 90°C for 20 min. Following lyophilization, the DNA was analysed by PAGE as above.

(c) Potassium permanganate (Figure 12b)

The complex was formed in 100 μl TE and 5 μl 20 mM KMnO_4 was added. After 15 s at 20°C, the reaction was stopped 15 by addition of 50 μl 1.5 M sodium acetate, pH 7.0, 1 M 2-mercaptoproethanol. The DNA was precipitated, treated with piperidine and analyzed, as above.

(d) Photofootprinting (Figure 12b)

The complex was formed in 100 μl TE and diazo-linked 20 acridine (0.1 $\mu\text{g}/\mu\text{l}$) (DHA, Nielsen et al. (1988) Nucl. Acids Res. 16, 3877-88) was added. The sample was irradiated at 365 nm (Philips TL 20 W/09N, $22 \text{ Jm}^{-2} \text{s}^{-1}$) for 30 min and treated as described for "affinity photocleavage".

(e) S₁-nuclease (Figure 12c)

25 The complex was formed in 50 mM sodium acetate, 200 mM NaCl, 0.5% glycerol, 1 mM ZnCl₂, pH 4.5 and treated with nuclease S₁ (Boehringer Mannheim) at 0.5 U/ml for 5 min at 20°C. The reaction was stopped and treated further as described under "Staphylococcus nuclease".

30

EXAMPLE 24

N-Benzylloxycarbonyl-N'-(bocaminoethyl)glycine.

Aminoethyl glycine (52.86 g; 0.447 mol) was dissolved in water (900 ml) and dioxane (900 ml) was added. The pH was 35 adjusted to 11.2 with 2N NaOH. While the pH was kept at 11.2, tert-butyl-p-nitrophenyl carbonate (128.4 g; 0.537 mol) was dissolved in dioxane (720 ml) and added dropwise over the

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course of 2 hours. The pH was kept at 11.2 for at least three more hours and then left with stirring overnight. The yellow solution was cooled to 0°C and the pH was adjusted to 3.5 with 2 N HCl. The mixture was washed with chloroform (4x100 ml), 5 and the pH of the aqueous phase was readjusted to 9.5 with 2 N NaOH at 0°C. Benzyloxycarbonyl chloride (73.5 ml; 0.515 mol) was added over half an hour, while the pH was kept at 9.5 with 2 N NaOH. The pH was adjusted frequently over the next 4 hours, and the solution was left with stirring overnight. 10 On the following day the solution was washed with ether (3x600 ml) and the pH of the solution was afterwards adjusted to 1.5 with 2 N HCl at 0°C. The title compound was isolated by extraction with ethyl acetate (5x1000 ml). The ethyl acetate solution was dried over magnesium sulfate and evaporated to 15 dryness, *in vacuo*. This afforded 138 g, which was dissolved in ether (300 ml) and precipitated by the addition of petroleum ether (1800 ml). Yield 124.7 g (79%). M.p. 64.5-85 °C. Anal. for C₁₇H₂₄N₂O₆ found(calc.) C: 58.40(57.94); H: 7.02(6.86); N: 7.94(7.95). ¹H-NMR (250 MHz, CDCl₃) 7.33 & 7.32 (5H, Ph); 5.15 & 5.12 (2H, PhCH₂); 4.03 & 4.01 (2H, NCH₂CO₂H); 20 3.46 (b, 2H, BocNHCH₂CH₂); 3.28 (b, 2H, BocNHCH₂CH₂); 1.43 & 1.40 (9H, 'Bu). HPLC (260 nm) 20.71 min. (80.2%) and 21.57 min. (19.8%). The UV-spectra (200 nm - 300 nm) are identical, indicating that the minor peak consists of Bis-Z-AEG.

25

EXAMPLE 25

N'-Boc-aminoethyl glycine ethyl ester.

N-Benzyloxycarbonyl-N'-(bocaminoethyl)glycine (60.0 g; 0.170 mol) and N,N-dimethyl-4-aminopyridine (6.00 g) were 30 dissolved in absolute ethanol (500 ml), and cooled to 0°C before the addition of DCC (42.2 g; 0.204 mol). The ice bath was removed after 5 minutes and stirring was continued for 2 more hours. The precipitated DCU (32.5 g dried) was removed by filtration and washed with ether (3x100 ml). The combined 35 filtrate was washed successively with diluted potassium hydrogen sulfate (2x400 ml), diluted sodium hydrogencarbonate (2x400 ml) and saturated sodium chloride (1x400 ml). The

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organic phase was filtered, then dried over magnesium sulfate, and evaporated to dryness, *in vacuo*, which yielded 66.1 g of an oily substance which contained some DCU.

The oil was dissolved in absolute ethanol (600 ml) and 5 was added 10% palladium on carbon (6.6 g) was added. The solution was hydrogenated at atmospheric pressure, where the reservoir was filled with 2 N sodium hydroxide. After 4 hours, 3.3 L was consumed out of the theoretical 4.2 L. The reaction mixture was filtered through celite and evaporated 10 to dryness, *in vacuo*, affording 39.5 g (94%) of an oily substance. A 13 g portion of the oily substance was purified by silica gel (600 g SiO₂) chromatography. After elution with 300 ml 20% petroleum ether in methylene chloride, the title compound was eluted with 1700 ml of 5% methanol in methylene 15 chloride. The solvent was removed from the fractions with satisfactory purity, *in vacuo* and the yield was 8.49 g. Alternatively 10 g of the crude material was purified by Kugel Rohr distillation. ¹H-NMR (250 MHz, CD₃OD); 4.77 (b, s, NH); 4.18 (q, 2H, MeCH₂-); 3.38 (s, 2H, NCH₂CO₂Et); 3.16 (t, 2H, 20 BocNHCH₂CH₂); 2.68 (t, 2H, BocNHCH₂CH₂); 1.43 (s, 9H, 'Bu) and 1.26 (t, 3H, CH₃) ¹³C-NMR 171.4 (COEt); 156.6 (CO); 78.3 ((CH₃)₂C); 59.9 (CH₂); 49.0 (CH₂); 48.1 (CH₂); 39.0 (CH₂); 26.9 (CH₂) and 12.6 (CH₃).

25 EXAMPLE 26

N'-Boc-aminoethyl glycine methyl ester.

The above procedure was used, with methanol being substituted for ethanol. The final product was purified by column purification.

30

EXAMPLE 27

1-(Boc-aeg)thymine ethyl ester.

N'-Boc-aminoethyl glycine ethyl ester (13.5 g; 54.8 mmol), DhbtOH (9.84 g; 60.3 mmol) and 1-carboxymethyl thymine 35 (11.1 g; 60.3 mmol) were dissolved in DMF (210 ml). Methylene chloride (210 ml) then was added. The solution was cooled to 0°C in an ethanol/ice bath and DCC (13.6 g; 65.8

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mmol) was added. The ice bath was removed after 1 hour and stirring was continued for another 2 hours at ambient temperature. The precipitated DCU was removed by filtration and washed twice with methylene chloride (2 x 75 ml). To the 5 combined filtrate was added more methylene chloride (650 ml). The solution was washed successively with diluted sodium hydrogen carbonate (3 x 500 ml), diluted potassium hydrogen sulfate (2 x 500 ml), and saturated sodium chloride (1 x 500 ml). Some precipitate was removed from the organic phase by 10 filtration. The organic phase was dried over magnesium sulfate and evaporated to dryness, *in vacuo*. The oily residue was dissolved in methylene chloride (150 ml), filtered, and the title compound was precipitated by the addition of petroleum ether (350 ml) at 0°C. The methylene 15 chloride/petroleum ether procedure was repeated once. This afforded 16.0 g (71%) of a material which was more than 99% pure by HPLC.

EXAMPLE 28

20 1-(Boc-aeg)thymine.

The material from above was suspended in THF (194 ml, gives a 0.2 M solution), and 1 M aqueous lithium hydroxide (116 ml) was added. The mixture was stirred for 45 minutes at ambient temperature and then filtered to remove residual 25 DCU. Water (40 ml) was added to the solution which was then washed with methylene chloride (300 ml). Additional water (30 ml) was added, and the alkaline solution was washed once more with methylene chloride (150 ml). The aqueous solution was cooled to 0°C and the pH was adjusted to 2 by the dropwise 30 addition of 1 N HCl (approx. 110 ml). The title compound was extracted with ethyl acetate (9 x 200 ml), the combined extracts were dried over magnesium sulfate and were evaporated to dryness, *in vacuo*. The residue was evaporated once from methanol, which after drying overnight afforded a colorless 35 glassy solid. Yield 9.57 g (64%). HPLC > 98% R_r=14.8 min. Anal. for C₁₆H₂₄N₄O₇·0.25 H₂O Found (calc.) C: 49.29(49.42); H: 6.52(6.35); N: 14.11(14.41). Due to the limited rotation

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around the secondary amide, several of the signals were doubled in the ratio 2:1 (indicated in the list by mj. for major and mi. for minor). $^1\text{H-NMR}$ (250 MHz, DMSO-d₆): 12.75 (b.s., 1H, CO₂H); 11.28 (s, "1H", mj., imide NH); 11.26 (s, 5 "1H", mi., imide NH); 7.30 (s, "1H", mj., T H-6); 7.26 (s, "1H", mi., T H-6); 6.92 (b.t., "1H", mj., BocNH); 6.73 (b.t., "1H", mi., BocNH); 4.64 (s, "2H", mj., CH₂CON); 4.46 (s, "2H", mj., CH₂CON); 4.19 (s, "2H", mi., CH₂CO₂H); 3.97 (s, "2H", mj., CH₂CO₂H); 3.63-3.01 (unresolved m, includes water, 10 CH₂CH₂); 1.75 (s, 3H, CH₃) and 1.38 (s, 9H, ^tBu).

EXAMPLE 29

N'-Benzylloxycarbonyl-1-(Boc-aeg)cytosine.

N'-Boc-aminoethyl glycine ethyl ester (5.00 g; 20.3 mmol), DhbtOH (3.64 g; 22.3 mmol) and N'-benzylloxycarbonyl-1-carboxymethyl cytosine (6.77 g; 22.3 mmol) were suspended in DMF (100 ml). Methylene chloride (100 ml) then was added. The solution was cooled to 0°C and DCC (5.03 g; 24.4 mmol) was added. The ice bath was removed after 2 h and stirring was 20 continued for another hour at ambient temperature. The reaction mixture then was evaporated to dryness, *in vacuo*. The residue was suspended in ether (100 ml) and stirred vigorously for 30 min. The solid material was isolated by filtration and the ether wash procedure was repeated twice. 25 The material was then stirred vigorously for 15 min with dilute sodium hydrogencarbonate (aprox. 4% solution, 100 ml), filtered and washed with water. This procedure was then repeated once, which after drying left 17.0 g of yellowish solid material. The solid was then boiled with dioxane (200 ml) and filtered while hot. After cooling, water (200 ml) was 30 added. The precipitated material was isolated by filtration, washed with water, and dried. According to HPLC (observing at 260 nm) this material has a purity higher than 99%, besides the DCU. The ester was then suspended in THF (100 ml), cooled 35 to 0°C, and 1 N LiOH (61 ml) was added. After stirring for 15 minutes, the mixture was filtered and the filtrate was washed with methylene chloride (2 x 150 ml). The alkaline

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solution then was cooled to 0°C and the pH was adjusted to 2.0 with 1 N HCl. The title compound was isolated by filtration and was washed once with water, leaving 11.3 g of a white powder after drying. The material was suspended in methylene chloride (300 ml) and petroleum ether (300 ml) was added. Filtration and wash afforded 7.1 g (69%) after drying. HPLC showed a purity of 99% R_f = 19.5 min, and a minor impurity at 12.6 min (approx. 1%) most likely the Z-de protected monomer. Anal. for $C_{22}H_{24}N_4O_8$ found(calc.) C: 54.16(54.87); H: 5.76(5.81)
10 and N: 13.65(13.91). 1H -NMR (250 MHz, DMSO-d₆). 10.78 (b.s, 1H, CO₂H); 7.88 (2 overlapping doublets, 1H, Cyt H-5); 7.41-7.32 (m, 5H, Ph); 7.01 (2 overlapping doublets, 1H, Cyt H-6); 6.94 & 6.78 (unres. triplets, 1H, BocNH); 5.19 (s, 2H, PhCH₂); 4.81 & 4.62 (s, 2H, CH₂CON); 4.17 & 3.98 (s, 2H, CH₂CO₂H);
15 3.42-3.03 (m, includes water, CH₂CH₂) and 1.38 & 1.37 (s, 9H, 'Bu). ^{13}C -NMR. 150.88; 128.52; 128.18; 127.96; 93.90; 66.53; 49.58 and 28.22. IR: Frequency in cm⁻¹ (intensity). 3423 (26.4), 3035 (53.2), 2978(41.4), 1736(17.3), 1658(3.8), 1563(23.0), 1501(6.8) and 1456 (26.4).

20

EXAMPLE 30

9-Carboxymethyl adenine ethyl ester.

Adenine (10.0 g, 74 mmol) and potassium carbonate (10.29 g, 74.0 mmol) were suspended in DMF and ethyl bromoacetate (8.24 ml, 74 mmol) was added. The suspension was stirred for 2.5 h under nitrogen at room temperature and then filtered. The solid residue was washed three times with DMF (10 ml). The combined filtrate was evaporated to dryness, *in vacuo*. The yellow-orange solid material was poured into water (200 ml) and 4 N HCl was added to pH≈6. After stirring at 0°C for 10 min, the solid was filtered off, washed with water, and recrystallized from 96% ethanol (150 ml). The title compound was isolated by filtration and washed thoroughly with ether. Yield 3.4 g (20%). M.p. 215.5-220°C. Anal. for $C_9H_{11}N_5O_2$ found(calc.): C: 48.86(48.65); H: 5.01(4.91); N: 31.66(31.42). 1H -NMR (250 MHz; DMSO-d₆): (s, 2H, H-2 & H-8), 7.25 (b. s., 2H, NH₂), 5.06 (s, 2H, NCH₂), 4.17 (q, 2H, J=7.11 Hz, OCH₂) and

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1.21 (t, 3H, J=7.13 Hz, NCH₂). ¹³C-NMR. 152.70, 141.30, 61.41, 43.97 and 14.07. FAB-MS. 222 (MH⁺). IR: Frequency in cm⁻¹ (intensity). 3855 (54.3), 3274(10.4), 3246(14.0), 3117(5.3), 2989(22.3), 2940(33.9), 2876(43.4), 2753(49.0), 2346(56.1), 5 2106(57.1), 1899(55.7), 1762(14.2), 1742(14.2), 1742(1.0), 1671(1.8), 1644(10.9), 1606(0.6), 1582(7.1), 1522(43.8), 1477(7.2), 1445(35.8) and 1422(8.6). The position of alkylation was verified by X-ray crystallography on crystals, which were obtained by recrystallization from 96% ethanol.

10 Alternatively, 9-carboxymethyl adenine ethyl ester can be prepared by the following procedure. To a suspension of adenine (50.0 g, 0.37 mol) in DMF (1100 ml) in 2 L three-necked flask equipped with a nitrogen inlet, a mechanical stirrer and a dropping funnel was added 16.4 g (0.407 mol) 15 hexane washed sodium hydride- mineral oil dispersion. The mixture was stirred vigorously for 2 hours, whereafter ethyl bromacetate 75 ml, 0.67 mol) was added dropwise over the course of 3 hours. The mixture was stirred for one additional hour, whereafter tlc indicated complete conversion of adenine. 20 The mixture was evaporated to dryness at 1 mmHg and water (500 ml) was added to the oily residue which caused crystallisation of the title compound. The solid was recrystallised from 96% ethanol (600 ml). Yield after drying 53.7 (65.6%). HPLC (215 nm) purity > 99.5%.

25

EXAMPLE 31

N^BBenzylloxycarbonyl-9-carboxymethyl adenine ethyl ester.

9-Carboxymethyladenine ethyl ester (3.40 g, 15.4 mmol) was dissolved in dry DMF (50 ml) by gentle heating, cooled to 30 20°C, and added to a solution of N-ethyl- benzylloxycarbonylimidazole tetrafluoroborate (62 mmol) in methylene chloride (50 ml) over a period of 15 min with ice-cooling. Some precipitation was observed. The ice bath was removed and the solution was stirred overnight. The reaction 35 mixture was treated with saturated sodium hydrogen carbonate (100 ml). After stirring for 10 min, the phases were separated and the organic phase was washed successively with

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one volume of water, dilute potassium hydrogen sulfate (twice), and with saturated sodium chloride. The solution was dried over magnesium sulfate and evaporated to dryness, *in vacuo*, which afforded 11 g of an oily material. The material 5 was dissolved in methylene chloride (25 ml), cooled to 0°C, and precipitated with petroleum ether (50 ml). This procedure was repeated once to give 3.45 g (63%) of the title compound. M.p. 132-35°C. Analysis for $C_{17}H_{17}N_5O_4$ found (calc.): C: 56.95(57.46); H: 4.71(4.82); N: 19.35(19.71). 1H -
10 NMR (250 MHz; $CDCl_3$): 8.77 (s, 1H, H-2 or H-8); 7.99 (s, 1H, H-2 or H-8); 7.45-7.26 (m, 5H, Ph); 5.31 (s, 2H, N-CH₂); 4.96 (s, 2H, Ph-CH₂); 4.27 (q, 2H, J=7.15 Hz, CH₂CH₃) and 1.30 (t, 3H, J=7.15 Hz, CH₂CH₃). ^{13}C -NMR: 153.09; 143.11; 128.66; 67.84; 62.51; 44.24 and 14.09. FAB-MS: 356 (MH⁺) and 312
15 (MH⁺-CO₂). IR: frequency in cm^{-1} (intensity). 3423 (52.1); 3182 (52.8); 3115(52.1); 3031(47.9); 2981(38.6); 1747(1.1); 1617(4.8); 15.87(8.4); 1552(25.2); 1511(45.2); 1492(37.9); 1465(14.0) and 1413(37.3).

20 EXAMPLE 32

N^6 -Benzylloxycarbonyl-9-carboxymethyl adenine.

N^6 -Benzylloxycarbonyl-9-carboxymethyladenine ethyl ester (3.20 g; 9.01 mmol) was mixed with methanol (50 ml) cooled to 0°C. Sodium Hydroxide Solution (50 ml; 2N) was added, whereby 25 the material quickly dissolved. After 30 min at 0°C, the alkaline solution was washed with methylene chloride (2x50ml). The aqueous solution was brought to pH 1.0 with 4 N HCl at 0°C, whereby the title compound precipitated. The yield after filtration, washing with water, and drying was 3.08 g (104%).
30 The product contained salt and elemental analysis reflected that. Anal. for $C_{15}H_{13}N_5O_4$ found(calc.): C: 46.32(55.05); H: 4.24(4.00); N: 18.10(21.40) and C/N: 2.57(2.56). 1H -NMR(250 MHz; DMSO-d₆): 8.70 (s, 2H, H-2 and H-8); 7.50-7.35 (m, 5H, Ph); 5.27 (s, 2H, N-CH₂); and 5.15 (s, 2H, Ph-CH₂). ^{13}C -NMR.
35 168.77, 152.54, 151.36, 148.75, 145.13, 128.51, 128.17, 127.98, 66.76 and 44.67. IR (KBr) 3484(18.3); 3109(15.9); 3087(15.0); 2966(17.1); 2927(19.9); 2383(53.8); 1960(62.7); 1739(2.5);

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1688(5.2); 1655(0.9); 1594(11.7); 1560(12.3); 1530(26.3);
1499(30.5); 1475(10.4); 1455(14.0); 1429(24.5) and 1411(23.6).
FAB-MS: 328 (MH^+) and 284 (MH^+-CO_2). HPLC (215 nm, 260 nm) in
system 1: 15.18 min, minor impurities all less than 2%.

5

EXAMPLE 33

N'-Benzylloxycarbonyl-1-(Boc-aeg)adenine ethyl ester.

N'-Boc-aminoethyl glycine ethyl ester (2.00 g; 8.12 mmol), DhbtOH (1.46 g; 8.93 mmol) and N'-benzylloxycarbonyl-9-10 carboxymethyl adenine (2.92 g; 8.93 mmol) were dissolved in DMF (15 ml). Methylene chloride (15 ml) then was added. The solution was cooled to 0°C in an ethanol/ice bath. DCC (2.01 g; 9.74 mmol) was added. The ice bath was removed after 2.5 h and stirring was continued for another 1.5 hour at ambient 15 temperature. The precipitated DCU was removed by filtration and washed once with DMF (15 ml), and twice with methylene chloride (2 x 15 ml). To the combined filtrate was added more methylene chloride (100 ml). The solution was washed successively with dilute sodium hydrogen carbonate (2 x 100 20 ml), dilute potassium hydrogen sulfate (2 x 100 ml), and saturated sodium chloride (1 x 100 ml). The organic phase was evaporated to dryness, *in vacuo*, which afforded 3.28 g (73%) of a yellowish oily substance. HPLC of the raw product showed a purity of only 66% with several impurities, both more and 25 less polar than the main peak. The oil was dissolved in absolute ethanol (50 ml) and activated carbon was added. After stirring for 5 minutes, the solution was filtered. The filtrate was mixed with water (30 ml) and was left with stirring overnight. The next day, the white precipitate was 30 removed by filtration, washed with water, and dried, affording 1.16 g (26%) of a material with a purity higher than 98% by HPLC. Addition of water to the mother liquor afforded another 0.53 g with a purity of approx. 95%. Anal. for $C_{22}H_{21}N_5O_7$ found(calc.) C: 55.01(54.44; H: 6.85(6.15) and N: 35 16.47(17.09). 1H -NMR (250 MHz, CDCl₃) 8.74 (s, 1H, Ade H-2); 8.18 (b. s, 1H, ZNH); 8.10 & 8.04 (s, 1H, H-8); 7.46-7.34 (m, 5H, Ph); 5.63 (unres. t, 1H, BocNH); 5.30 (s, 2H, PhCH₂); 5.16

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& 5.00 (s, 2H, CH_2CON); 4.29 & 4.06 (s, 2H, $\text{CH}_2\text{CO}_2\text{H}$); 4.20 (q, 2H, OCH_2CH_3); 3.67-3.29 (m, 4H, CH_2CH_2); 1.42 (s, 9H, 'Bu) and 1.27 (t, 3H, OCH_2CH_3). The spectrum shows traces of ethanol and DCU.

5

EXAMPLE 34

$\text{N}^6\text{-Benzylloxycarbonyl-1-(Boc-aeg)adenine}$.

$\text{N}^6\text{-Benzylloxycarbonyl-1-(Boc-aeg)adenine ethyl ester}$ (1.48 g; 2.66 mmol) was suspended in THF (13 ml) and the mixture was cooled to 0°C. Lithium hydroxide (8 ml; 1 N) was added. After 15 min of stirring, the reaction mixture was filtered, extra water (25 ml) was added, and the solution was washed with methylene chloride (2 x 25 ml). The pH of the aqueous solution was adjusted to pH 2.0 with 1 N HCl. The precipitate was isolated by filtration, washed with water, and dried, and dried affording 0.82 g (58%). The product reprecipitated twice with methylene chloride/petroleum ether, 0.77 g (55%) after drying. M.p. 119°C (decomp.) Anal. for $\text{C}_{24}\text{H}_{28}\text{N}_4\text{O}_4\text{H}_2\text{O}$ found(calc.) C: 53.32(52.84); H: 5.71(5.73); N: 17.68(17.97). FAB-MS. 528.5 (MH^+). $^1\text{H-NMR}$ (250 MHz, DMSO-d_6) . 12.75 (very b, 1H, CO_2H); 10.65 (b. s, 1H, ZNH); 8.59 (d, 1H, $J = 2.14$ Hz, Ade H-2); 8.31 (s, 1H, Ade H-8); 7.49-7.31 (m, 5H, Ph); 7.03 & 6.75 (unresol. t, 1H, BocNH); 5.33 & 5.16 (s, 2H, CH_2CON); 5.22 (s, 2H, PhCH_3); 4.34-3.99 (s, 2H, $\text{CH}_2\text{CO}_2\text{H}$); 3.54-3.03 (m's, includes water, CH_2CH_2) and 1.39 & 1.37 (s, 9H, 'Bu). $^{13}\text{C-NMR}$. 170.4; 166.6; 152.3; 151.5; 149.5; 145.2; 128.5; 128.0; 127.9; 66.32; 47.63; 47.03; 43.87 and 28.24.

EXAMPLE 35

30 2-Amino-6-chloro-9-carboxymethylpurine.

To a suspension of 2-amino-6-chloropurine (5.02 g; 29.6 mmol) and potassium carbonate (12.91 g; 93.5 mmol) in DMF (50 ml) was added bromoacetic acid (4.70 g; 22.8 mmol). The mixture was stirred vigorously for 20 h. under nitrogen. Water (150 ml) was added and the solution was filtered through Celite to give a clear yellow solution. The solution was acidified to a pH of 3 with 4 N hydrochloric acid. The

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precipitate was filtered and dried, *in vacuo*, over sicapent. Yield (3.02 g; 44.8%). $^1\text{H-NMR}$ (DMSO-d₆): δ = 4.88 ppm (s, 2H); 6.95 (s, 2H); 8.10 (s, 1H).

5 EXAMPLE 36

2-Amino-6-benzyloxy-9-carboxymethylpurine.

Sodium (2.0 g; 87.0 mmol) was dissolved in benzyl alcohol (20 ml) and heated to 130°C for 2 h. After cooling to 0°C, a solution of 2-amino-6-chloro-9-carboxymethylpurine (4.05 g; 18.0 mmol) in DMF (85 ml) was slowly added, and the resulting suspension stirred overnight at 20°C. Sodium hydroxide solution (1N, 100 ml) was added and the clear solution was washed with ethyl acetate (3 x 100 ml). The water phase then was acidified to a pH of 3 with 4 N hydrochloric acid. The precipitate was taken up in ethyl acetate (200 ml), and the water phase was extracted with ethyl acetate (2 x 100 ml). The combined organic phases were washed with saturated sodium chloride solution (2 x 75 ml), dried with anhydrous sodium sulfate, and taken to dryness by evaporation, *in vacuo*. The residue was recrystallized from ethanol (300 ml). Yield after drying, *in vacuo*, over sicapent: 2.76 g (52%). M.p. 159–65°C. Anal. (calc., found) C(56.18; 55.97), H(4.38; 4.32), N(23.4; 23.10). $^1\text{H-NMR}$ (DMSO-d₆): 4.82 ppm. (s, 2H); 5.51 (s, 2H); 6.45 (s, 2H); 7.45 (m, 5H); 7.82 (s, 1H).

EXAMPLE 37

N-([2-Amino-6-benzyloxy-purine-9-yl]-acetyl)-N-(2-Boc-aminoethyl)-glycine [BocGaeg-OH monomer].

2-Amino-6-benzyloxy-9-carboxymethyl-purine (0.50 g; 1.67 mmol), methyl-N(2-[tert-butoxycarbonylamino]ethyl)-glycinate (0.65 g; 2.80 mmol), diisopropylethyl amine (0.54 g; 4.19 mmol), and bromo-tris-pyrrolidino-phosphonium-hexafluoro-phosphate (PyBroP®) (0.798 g; 1.71 mmol) were stirred in DMF (2 ml) for 4 h. The clear solution was poured into an ice-cooled solution of sodium hydrogen carbonate (1 N; 40 ml) and extracted with ethyl acetate (3 X 40 ml). The

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organic layer was washed with potassium hydrogen sulfate solution (1 N; 2 X 40 ml), sodium hydrogen carbonate (1 N; 1 X 40 ml) and saturated sodium chloride solution (60 ml). After drying with anhydrous sodium sulfate and evaporation, 5 in vacuo, the solid residue was recrystallized from ethyl acetate/hexane (20 ml (2:1)) to give the methyl ester in 63% yield (MS-FAB 514 (M+1)). Hydrolysis was accomplished by dissolving the ester in ethanol/water (30 ml (1:2)) containing conc. sodium hydroxide (1 ml). After stirring for 2 h, the 10 solution was filtered and acidified to a pH of 3, by the addition of 4 N hydrochloric acid. The title compound was obtained by filtration. Yield: 370 mg (72% for the hydrolysis). Purity by HPLC was more than 99%. Due to the limited rotation around the secondary amide several of the 15 signals were doubled in the ratio 2:1 (indicated in the list by mj. for major and mi. for minor). ¹H-NMR(250, MHz, DMSO-d₆): d = 1.4 ppm. (s, 9H); 3.2 (m, 2H); 3.6 (m, 2H); 4.1 (s, mj., CONRCH₂COOH); 4.4 (s, mi., CONRCH₂COOH); 5.0 (s, mi., Gua-CH₂CO-); 5.2 (s, mj., Gua-CH₂CO); 5.6 (s, 2H); 6.5 (s, 2H); 6.9 20 (m, mi., BocNH); 7.1 (m, mj., BocNH); 7.5 (m., 3H); 7.8 (s, 1H); 12,8 (s;1H). ¹³C-NMR. 170.95; 170.52; 167.29; 166.85; 160.03; 159.78; 155.84; 154.87; 140.63; 136.76; 128.49; 128.10; 113.04; 78.19; 77.86; 66.95; 49.22; 47.70; 46.94; 45.96; 43.62; 43.31 and 28.25.

25

EXAMPLE 38

3-Boc-amino-1,2-propanediol.

3-Amino-1,2-propanediol (40.00 g, 0.440 mol, 1.0 eq.) was dissolved in water (1000 ml) and cooled to 0°C. Di-tert-butyl dicarbonate (115.0 g, 0.526 mol, 1.2 eq.) was added in one portion. The reaction mixture was heated to room temperature on a water bath during stirring. The pH was maintained at 10.5 with a solution of sodium hydroxide (17.56 g, 0.440 mol, 1.0 eq.) in water (120 ml). When the addition 35 of aqueous sodium hydroxide was completed, the reaction mixture was stirred overnight at room temperature. Subsequently, ethyl acetate (750 ml) was added to the reaction

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mixture, followed by cooling to 0°C. The pH was adjusted to 2.5 with 4 N sulphuric acid with vigorous stirring. The phases were separated and the water phase was washed with additional ethyl acetate (6x350 ml). The volume of the 5 organic phase was reduced to 900 ml by evaporation under reduced pressure. The organic phase then was washed with a saturated aqueous solution of potassium hydrogen sulfate diluted to twice its volume (1x1000 ml) and with saturated aqueous sodium chloride (1x500 ml). The organic phase was 10 dried ($MgSO_4$) and evaporated under reduced pressure to yield 50.12 g (60%) of the title compound. The product could be solidified by evaporation from methylene chloride and subsequent freezing. 1H -NMR ($CDCl_3/TMS$): δ = 1.43 (s, 9H, Me_3C), 3.25 (m, 2H, CH_2), 3.57 (m, 2H, CH_2), 3.73 (m, 1H, CH). 15 ^{13}C -NMR ($CDCl_3/TMS$): δ = 28.2 (Me_3C), 42.6 (CH_2), 63.5, 71.1 (CH_2OH , $CHOH$), 79.5 (Me_3C), 157.0 (C=O).

EXAMPLE 39

2-(Boc-amino)ethyl-L-alanine methyl ester.

20 3-Boc-amino-1,2-propanediol (20.76 g, 0.109 mol, 1 eq.) was suspended in water (150 ml). Potassium *m*-periodate (24.97 g, 0.109 mol, 1 eq.) was added and the reaction mixture was stirred for 2 h at room temperature under nitrogen. The reaction mixture was filtered and the water phase extracted 25 with chloroform (6x250 ml). The organic phase was dried ($MgSO_4$) and evaporated to afford an almost quantitative yield of Boc-aminoacetaldehyde as a colourless oil, which was used without further purification in the following procedure.

30 Palladium-on-carbon (10%, 0.8 g) was added to MeOH (250 ml) under nitrogen with cooling (0°C) and vigorous stirring. Anhydrous sodium acetate (4.49 g, 54.7 mmol, 2 eqv) and L-alanine methyl ester, hydrochloride (3.82 g, 27.4 mmol, 1 eqv) were added. Boc-aminoacetaldehyde (4.79 g, 30.1 mmol, 1.1 eqv) was dissolved in MeOH (150 ml) and added to the reaction 35 mixture. The reaction mixture was hydrogenated at atmospheric pressure and room temperature until hydrogen uptake had ceased. The reaction mixture was filtered through celite,

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which was washed with additional MeOH. The MeOH was removed under reduced pressure. The residue was suspended in water (150 ml) and pH adjusted to 8.0 by dropwise addition of 0.5 N NaOH with vigorous stirring. The water phase was extracted 5 with methylene chloride (4x250 ml). The organic phase was dried ($MgSO_4$), filtered through celite, and evaporated under reduced pressure to yield 6.36 g (94%) of the title compound as a clear, slightly yellow oil. MS (FAB-MS): m/z (%) = 247 (100, M+1, 191 (90), 147 (18)). 1H -NMR (250 MHz, $CDCl_3$): 1.18 (d, J=7.0 Hz, 3H, Me), 1.36 (s, 9H, Me_3C), 1.89 (b, 1H, NH), 2.51 (m, 1H, CH_2), 2.66 (m, 1H, CH_2), 3.10 (m, 2H, CH_2), 3.27 (q, J=7.0 Hz, 1H, CH), 3.64 (s, 3H, OMe), 5.06 (b, 1H, carbamate NH). ^{13}C -NMR: d = 18.8 (Me), 28.2 (Me_3C), 40.1, 47.0 (CH_2), 51.6 (OMe), 56.0 (CH), 155.8 (carbamate C=O), 175.8 15 (ester C=O).

EXAMPLE 40

N-(Boc-aminoethyl)-N-(1-thyminylacetyl)-L-alanine methyl ester.

20 To a solution of Boc-aminoethyl-(L)-alanine methyl ester (1.23 g, 5.0 mmol) in DMF (10 ml) was added Dhbt-OH (0.90 g, 5.52 mmol) and 1-thyminylacetic acid (1.01 g, 5.48 mmol). When the 1-thyminylacetic acid was dissolved, dichloromethane (10 ml) was added and the solution was cooled 25 on an ice bath. After the reaction mixture had reached 0°C, DCC (1.24 g, 6.01 mmol) was added. Within 5 min after the addition, a precipitate of DCU was seen. After a further 5 min, the ice bath was removed. Two hours later, TLC analysis showed the reaction to be finished. The mixture was filtered 30 and the precipitate washed with dichloromethane (100ml). The resulting solution was extracted twice with 5% sodium hydrogen carbonate (150 ml) and twice with saturated potassium hydrogen sulfate (25 ml) in water (100 ml). After a final extraction with saturated sodium chloride (150 ml), the solution was 35 dried with magnesium sulfate and evaporated to give a white foam. The foam was purified by column chromatography on silica gel using dichloromethane with a methanol gradient as

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eluent. This yielded a pure compound (>99% by HPLC) (1.08 g, 52.4%). FAB-MS: 413 (M+1) and 431 (M+1 + water). ¹H-NMR (CDCl₃): 4.52 (s, 2 H, CH₂); 3.73 (s, 3 H, OMe); 3.2-3.6 (m, 4 H, ethyl CH₂'s); 1.90 (s, 3 H, Me in T); 1.49 (d, 3 H, Me in Ala, J=7.3 Hz); 1.44 (s, 9 H, Boc).

EXAMPLE 41

N-(Boc-aminoethyl)-N-(1-thyminylacetyl)-L-alanine.

The methyl ester of the title compound (2.07 g, 5.02 mmol) was dissolved in methanol (100 ml), and cooled on an ice bath. 2 M sodium hydroxide (100 ml) was added. After stirring for 10 min, the pH of the mixture was adjusted to 3 with 4 M hydrogen chloride. The solution was subsequently extracted with ethyl acetate (3 x 100 ml). The combined organic extracts were dried over magnesium sulfate. After evaporation, the resulting foam was dissolved in ethyl acetate (400 ml) and a few ml of methanol to dissolve the solid material. Petroleum ether then was added until precipitation started. After standing overnight at -20°C, the precipitate was removed by filtration. This gave 1.01 g (50.5%) of pure compound (>99% by HPLC). The compound can be recrystallized from 2-propanol. FAB-MS: 399 (M+1). ¹H-NMR (DMSO-d₆): 11.35 (s, 1 H, COO); 7.42 (s, 1 H, H_{6'}); 4.69 (s, 2 H, CH₂); 1.83 (s, 3 H, Me in T); 1.50-1.40 (m, 12 H, Me in Ala + Boc).

25

EXAMPLE 42

(a) **N-(Boc-aminoethyl)--N-(1-thyminylacetyl)-D-alanine methyl ester.**

To a solution of Boc-aminoethyl alanine methyl ester (2.48 g, 10.1 mmol) in DMF (20 ml) was added Dhbt-OH (1.80 g, 11.0 mmol) and thyminylacetic acid (2.14 g, 11.6 mmol). After dissolution of the 1-thyminylacetic acid, methylene chloride (20 ml) was added and the solution cooled on an ice bath. When the reaction mixture had reached 0°C, DCC (2.88 g, 14.0 mmol) was added. Within 5 min after the addition a precipitate of DCU was seen. After 35 min the ice bath was removed. The reaction mixture was filtered 3.5 h later and

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the precipitate washed with methylene chloride (200 ml). The resulting solution was extracted twice with 5% sodium hydrogen carbonate (200 ml) and twice with saturated potassium hydrogen sulfate in water (100 ml). After a final extraction with 5 saturated sodium chloride (250 ml), the solution was dried with magnesium sulfate and evaporated to give an oil. The oil was purified by short column silica gel chromatography using methylene chloride with a methanol gradient as eluent. This yielded a compound which was 96% pure according to HPLC (1.05 10 g, 25.3%) after precipitation with petroleum ether. FAB-MS: 413 (M+1). $^1\text{H-NMR}$ (CDCl_3): 5.64 (t, 1 H, BocNH, $J=5.89$ Hz); 4.56 (d, 2 H, CH'_2); 4.35 (q, 1 H, CH in Ala, $J=7.25$ Hz); 3.74 (s, 3 H, OMe); 3.64-3.27 (m, 4 H, ethyl H's); 1.90 (s, 3 H, Me in T); 1.52-1.44 (t, 12 H, Boc+Me in Ala).

15 (b) N-(Boc-aminoethyl)-N-(1-thyminylacetyl)-D-alanine
The methyl ester of the title compound (1.57 g, 3.81 mmol) was dissolved in methanol (100 ml) and cooled on an ice bath. Sodium hydroxide (100 ml; 2 M) was added. After stirring for 10 min the pH of the mixture was adjusted to 3 20 with 4 M hydrogen chloride. The solution then was extracted with ethyl acetate (3 x 100 ml). The combined organic extracts were dried over magnesium sulfate. After evaporation, the oil was dissolved in ethyl acetate (200 ml). Petroleum ether was added (to a total volume of 600 ml) until 25 precipitation started. After standing overnight at -20°C, the precipitate was removed by filtration. This afforded 1.02 g (67.3%) of the title compound, which was 94% pure according to HPLC. FAB-MS: 399 (M+1). $^1\text{H-NMR}$: 11.34 (s, 1 H, COOH); 7.42 (s, 1 H, H'_6); 4.69 (s, 2 H, CH'_2); 4.40 (q, 1 H, CH in 30 Ala, $J=7.20$ Hz); 1.83 (s, 3 H, Me in T); 1.52-1.40 (m, 12 H, Boc + Me in Ala).

EXAMPLE 43

N-(N'-Boc-3'-aminopropyl)-N-[(1-thyminyl)acetyl]glycine methyl 35 ester.

N-(N'-Boc-3'-aminopropyl)glycine methyl ester (2.84 g, 0.0115 mol) was dissolved in DMF (35 ml), followed by addition

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of DhbtOH (2.07 g, 0.0127 mol) and 1-thyminylacetic acid (2.34 g, 0.0127 mol). Methylene chloride (35 ml) was added and the mixture cooled to 0°C on an ice bath. After addition of DCC (2.85 g, 0.0138 mol), the mixture was stirred at 0°C for 2 h, 5 followed by 1 h at room temperature. The precipitated DCU was removed by filtration, washed with methylene chloride (25 ml), and a further amount of methylene chloride (150 ml) was added to the filtrate. The organic phase was extracted with sodium hydrogen carbonate (1 volume saturated diluted with 1 volume 10 water, 6 x 250 ml), potassium sulfate (1 volume saturated diluted with 4 volumes water, 3 x 250 ml), and saturated aqueous sodium chloride (1 x 250 ml), dried over magnesium sulfate, and evaporated to dryness, *in vacuo*. The solid residue was suspended in methylene chloride (35 ml) and 15 stirred for 1h. The precipitated DCU was removed by filtration and washed with methylene chloride (25 ml). The filtrate was evaporated to dryness, *in vacuo*, and the residue purified by column chromatography on silica gel, eluting with a mixture of methanol and methylene chloride (gradient from 20 3-7% methanol in methylene chloride). This afforded the title compound as a white solid (3.05 g, 64%). M.p.. 76-79°C (decomp.). Anal. for C₁₈H₂₈N₄O₇, found (calc.) C: 52.03 (52.42) H: 6.90 (6.84) N: 13.21 (13.58). The compound showed satisfactory ¹H and ¹³C-NMR spectra.

25

EXAMPLE 44

N-(N'-Boc-3'-aminopropyl)-N-[(1-thyminyl)acetyl]glycine.

N-(N'-Boc-3'-aminopropyl)-N-[(1-thyminyl)acetyl]glycine methyl ester (3.02 g, 0.00732 mol) was dissolved in methanol 30 (25 ml) and stirred for 1.5 h with 2 M sodium hydroxide (25 ml). The methanol was removed by evaporation, *in vacuo*, and pH adjusted to 2 with 4 M hydrochloric acid at 0°C. The product was isolated as white crystals by filtration, washed with water (3 x 10 ml), and dried over sicapent, *in vacuo*. 35 Yield 2.19 g (75%). Anal. for C₁₇H₂₆N₄O₇, H₂O, found (calc.) C: 49.95 (49.03) H: 6.47 (6.29) N: 13.43 (13.45). The compound showed satisfactory ¹H and ¹³C-NMR spectra.

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EXAMPLE 45

3-(1-Thyminyl)-propanoic acid methyl ester.

Thymine (14.0 g, 0.11 mol) was suspended in methanol. Methyl acrylate (39.6 ml, 0.44 mol) was added, along with 5 catalytic amounts of sodium hydroxide. The solution was refluxed in the dark for 45 h, evaporated to dryness, in vacuo, and the residue dissolved in methanol (8 ml) with heating. After cooling on an ice bath, the product was precipitated by addition of ether (20 ml), isolated by 10 filtration, washed with ether (3 x 15 ml), and dried over sicapent, in vacuo. Yield 11.23 g (48%). M.p. 112-119°C. Anal. for $C_9H_{12}N_2O_4$, found (calc.) C: 51.14 (50.94) H: 5.78 (5.70) N: 11.52 (13.20). The compound showed satisfactory 1H and ^{13}C -NMR spectra.

15

EXAMPLE 46

3-(1-Thyminyl)-propanoic acid.

3-(1-Thyminyl)-propanoic acid methyl ester (1.0 g, 0.0047 mol) was suspended in 2 M sodium hydroxide (15 ml), 20 boiled for 10 min. The pH was adjusted to 0.3 with conc. hydrochloric acid. The solution was extracted with ethyl acetate (10 x 25 ml). The organic phase was extracted with saturated aqueous sodium chloride, dried over magnesium sulfate, and evaporated to dryness, in vacuo, to give the 25 title compound as a white solid (0.66 g, 71%). M.p. 118-121°C. Anal. for $C_8H_{10}N_2O_4$, found (calc.) C: 48.38 (48.49) H: 5.09 (5.09) N: 13.93 (14.14). The compound showed satisfactory 1H and ^{13}C -NMR spectra.

30 **EXAMPLE 47**

N-(N'-Boc-aminoethyl)-N-[(1-thyminyl)propanoyl]glycine ethyl ester.

N-(N'-Boc-aminoethyl)glycine ethyl ester (1.0 g, 0.0041 mol) was dissolved in DMF (12 ml). DhbtOH (0.73 g, 0.0045 mol) and 3-(1-thyminyl)-propanoic acid (0.89 g, 0.0045 mol) were added. Methylene chloride (12 ml) then was added and the mixture was cooled to 0°C on an ice bath. After addition of

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DCC (1.01 g, 0.0049 mol), the mixture was stirred at 0°C for 2 h, followed by 1 h at room temperature. The precipitated DCU was removed by filtration, washed with methylene chloride (25 ml), and a further amount of methylene chloride (50 ml) 5 was added to the filtrate. The organic phase was extracted with sodium hydrogen carbonate (1 volume saturated diluted with 1 volume water, 6 x 100 ml), potassium sulfate (1 volume saturated diluted with 4 volumes water, 3 x 100 ml), and saturated aqueous sodium chloride (1 x 100 ml), dried over 10 magnesium sulfate, and evaporated to dryness, *in vacuo*. The solid residue was suspended in methylene chloride (15 ml), and stirred for 1h. The precipitated DCU was removed by filtration and washed with methylene chloride. The filtrate was evaporated to dryness, *in vacuo*, and the residue purified 15 by column chromatography on silica gel, eluting with a mixture of methanol and methylene chloride (gradient from 1 to 6% methanol in methylene chloride). This afforded the title compound as a white solid (1.02 g, 59%). Anal. for C₁₉H₃₀N₄O₇, found (calc.) C: 53.15 (53.51) H: 6.90 (7.09) N: 12.76 20 (13.13). The compound showed satisfactory ¹H and ¹³C-NMR spectra.

EXAMPLE 48

N-(N'-Boc-aminoethyl)-N-[(1-thyminyl)propanoyl]glycine .
25 N-(N'-Boc-aminoethyl)-N-[(1-thyminyl)propanoyl]glycine ethyl ester (0.83 g, 0.00195 mol) was dissolved in methanol (25 ml). Sodium hydroxide (25 ml; 2 M) was added. The solution was stirred for 1 h. The methanol was removed by evaporation, *in vacuo*, and the pH adjusted to 2 with 4 M 30 hydrochloric acid at 0°C. The product was isolated by filtration, washed with ether (3 x 15 ml), and dried over sicapent, *in vacuo*. Yield 0.769 g, 99%). M.p. 213°C (de-comp.).

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EXAMPLE 49

Mono-Boc- ϵ -ethylenediamine (2).

tert-Butyl-4-nitrophenyl carbonate (1) (10.0 g; 0.0418 mol) dissolved in DMF (50 ml) was added dropwise over a period of 30 min to a solution of ethylenediamine (27.9 ml; 0.418 mol) and DMF (50 ml) and stirred overnight. The mixture was evaporated to dryness, *in vacuo*, and the resulting oil dissolved in water (250 ml). After cooling to 0°C, pH was adjusted to 3.5 with 4 M hydrochloric acid. The solution then was filtered and extracted with chloroform (3x250 ml). The pH was adjusted to 12 at 0°C with 2 M sodium hydroxide, and the aqueous solution extracted with methylene chloride (3x300 ml). After treatment with sat. aqueous sodium chloride (250 ml), the methylene chloride solution was dried over magnesium sulfate. After filtration, the solution was evaporated to dryness, *in vacuo*, resulting in 4.22 g (63%) of the product (oil). $^1\text{H-NMR}$ (90 MHz; CDCl_3): δ 1.44 (s, 9H); 2.87 (t, 2H); 3.1 (q, 2H); 5.62 (s, broad).

20 EXAMPLE 50

(N-Boc-aminoethyl)- β -alanine methyl ester, HCl.

Mono-Boc-ethylenediamine (2) (16.28 g; 0.102 mol) was dissolved in acetonitrile (400 ml) and methyl acrylate (91.50 ml; 1.02 mol) was transferred to the mixture with acetonitrile (200 ml). The solution was refluxed overnight under nitrogen in the dark to avoid polymerization of methyl acrylate. After evaporation to dryness, *in vacuo*, a mixture of water and ether (200 + 200 ml) was added, and the solution was filtered and vigorously stirred. The aqueous phase was extracted one more time with ether and then freeze dried to yield a yellow solid. Recrystallization from ethyl acetate yielded 13.09 g (46%) of the title compound. M.p. 138-140°C. Anal. for $\text{C}_{11}\text{H}_{23}\text{N}_2\text{O}_4\text{Cl}$, found (calc.) C: 46.49 (46.72) H: 8.38 (8.20) N: 9.83 (9.91) Cl: 12.45 (12.54). $^1\text{H-NMR}$ (90 MHz; DMSO-d_6): δ 1.39 (s, 9H); 2.9 (m, 8H); 3.64 (s, 3H).

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EXAMPLE 51

N-[(1-Thyminyl)acetyl]-N'-Boc-aminoethyl- β -alanine methyl ester.

(N-Boc-amino-ethyl)- β -alanine methyl ester, HCl (3)
5 (2.0 g; 0.0071 mol) and 1-thyminylacetic acid pentafluorophenyl ester (5) (2.828 g; 0.00812 mol) were dissolved in DMF (50 ml). Triethyl amine (1.12 ml; 0.00812 mol) was added and the mixture stirred overnight. After addition of methylene chloride (200 ml) the organic phase was
10 extracted with aqueous sodium hydrogen carbonate (3x250 ml), half-sat. aqueous potassium hydrogen sulfate (3x250 ml), and sat. aqueous sodium chloride (250 ml) and dried over magnesium sulfate. Filtration and evaporation to dryness, *in vacuo*, resulted in 2.9 g (99%) yield (oil). $^1\text{H-NMR}$ (250 MHz; CDCl_3):
15 due to limited rotation around the secondary amide several of the signals were doubled; δ 1.43 (s, 9H); 1.88 (s, 3H); 2.63 (t, 1H); 2.74 (t, 1H); 3.25-3.55 (4xt, 8H); 3.65 (2xt, 2H); 3.66 (s, 1.5); 3.72 (s, 1.5); 4.61 (s, 1H); 4.72 (s, 2H); 5.59 (s, 0.5H); 5.96 (s, 0.5H); 7.11 (s, 1H); 10.33 (s, 1H).

20

EXAMPLE 52

N-[(1-Thyminyl)acetyl]-N'-Boc-aminoethyl- β -alanine.

N-[(1-Thyminyl)acetyl]-N'-Boc-aminoethyl- β -alanine methyl ester (3.0 g; 0.0073 mol) was dissolved in 2 M sodium hydroxide (30 ml), the pH adjusted to 2 at 0°C with 4 M hydrochloric acid, and the solution stirred for 2 h. The precipitate was isolated by filtration, washed three times with cold water, and dried over sicapent, *in vacuo*. Yield 2.23 g (77%). M.p. 170-176°C. Anal. for $\text{C}_{17}\text{H}_{26}\text{N}_4\text{O}_7 \cdot \text{H}_2\text{O}$, found
25 (calc.) C: 49.49 (49.03) H: 6.31 (6.78) N: 13.84 (13.45). $^1\text{H-NMR}$ (90 MHz; DMSO-d_6): δ 1.38 (s, 9H); 1.76 (s, 3H); 2.44 and 3.29 (m, 8H); 4.55 (s, 2H); 7.3 (s, 1H); 11.23 (s, 1H). FAB-
30 MS: 399 (M+1).

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EXAMPLE 53

N-[(1-(N'-Z)-cytosyl)acetyl]-N'-Boc-aminoethyl- β -alanine methyl ester.

(N-Boc-amino-ethyl)- β -alanine methyl ester, HCl (3)
5 (2.0 g; 0.0071 mol) and 1-(N-4-Z)-cytosylacetic acid
pentafluorophenyl ester (5) (3.319 g; 0.0071 mol) were
dissolved in DMF (50 ml). Triethyl amine (0.99 ml; 0.0071
mol) was added and the mixture stirred overnight. After
addition of methylene chloride (200 ml), the organic phase was
10 extracted with aqueous sodium hydrogen carbonate (3x250 ml),
half-sat. aqueous potassium hydrogen sulfate (3x250 ml), and
sat. aqueous sodium chloride (250 ml), and dried over
magnesium sulfate. Filtration and evaporation to dryness, *in*
vacuo, resulted in 3.36 g of solid compound which was
15 recrystallized from methanol. Yield 2.42 g (64%). M.p. 158-
161°C. Anal. for C₂₅H₃₃N₅O₈, found (calc.) C: 55.19 (56.49) H:
6.19 (6.26) N: 12.86 (13.18). ¹H-NMR (250 MHz; CDCl₃): due to
limited rotation around the secondary amide several of the
signals were doubled; δ 1.43 (s, 9H); 2.57 (t, 1H); 3.60-3.23
20 (m's, 6H); 3.60 (s, 1.5H); 3.66 (s, 1.5H); 4.80 (s, 1H); 4.88
(s, 1H); 5.20 (s, 2H); 7.80-7.25 (m's, 7H). FAB-MS: 532
(M+1).

EXAMPLE 54

25 N-[(1-(N'-Z)-cytosyl)acetyl]-N'-Boc-aminoethyl- β -alanine.

N-[(1-(N-4-Z)-cytosyl)acetyl]-N'-Boc-aminoethyl- β -
alanine methyl ester (0.621 g; 0.0012 mol) was dissolved in
2 M sodium hydroxide (8.5 ml) and stirred for 2h.
Subsequently, pH was adjusted to 2 at 0°C with 4 M
30 hydrochloric acid and the solution stirred for 2 h. The
precipitate was isolated by filtration, washed three times
with cold water, and dried over sicapent, *in* vacuo. Yield
0.326 g (54%). The white solid was recrystallized from 2-
propanol and washed with petroleum ether. Mp. 163°C (decomp.).
35 Anal. for C₂₄H₃₁N₅O₈, found (calc.) C: 49.49 (49.03) H: 6.31
(6.78) N: 13.84 (13.45). ¹H-NMR (250 MHz; CDCl₃): due to
limited rotation around the secondary amide several of the

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signals were doubled; δ 1.40 (s, 9H); 2.57 (t, 1H); 2.65 (t, 1H); 3.60-3.32 (m's, 6H); 4.85 (s, 1H); 4.98 (s, 1H); 5.21 (s, 2H); 5.71 (s, 1H, broad); 7.99-7.25 (m's, 7H). FAB-MS: 518 (M+1).

5

EXAMPLE 55

Example of a PNA-oligomer with a guanine residue

(a) Solid-Phase Synthesis of H-[Taeg]-[Gaeg]-[Taeg].-Lys-NH₂

10 The protected PNA was assembled onto a Boc-Lys(ClZ) modified MBHA resin with a substitution of approximately 0.15 mmol/g (determined by quantitative Ninhydrin reaction). Capping of uncoupled amino groups was only carried out before the incorporation of the BocGaeg-OH monomer.

15 (b) Stepwise Assembly of H-[Taeg]-[Gaeg]-[Taeg].-Lys-NH₂ (synthetic protocol)

Synthesis was initiated on 102 mg (dry weight) of preswollen (overnight in DCM) and neutralized Boc-Lys(ClZ)-MBHA resin. The steps performed were as follows: (1) Boc-
20 deprotection with TFA/DCM (1:1, v/v), 1 x 2 min and 1 x 1/2 h, 3 ml; (2) washing with DCM, 4 x 20 sec, 3 ml; washing with DMF, 2 x 20 sec, 3 ml; washing with DCM, 2 x 20 sec, 3 ml, and drain for 30 sec; (3) neutralization with DIEA/DCM (1:19 v/v), 2 x 3 min, 3 ml; (4) washing with DCM, 4 x 20 sec, 3 ml, and drain for 1 min.; (5) addition of 4 equiv. diisopropyl carbodiimide (0.06 mmol; 9.7 μ l) and 4 equiv. (0.06 mmol; 24 mg) BocTaeg-OH or (0.06 mmol; 30 mg) BocGaeg-OH dissolved in 0.6 ml DCM/DMF (1:1, v/v) (final concentration of monomer 0.1 M), the coupling reaction was
30 allowed to proceed for 1/2 h shaking at room temperature; (6) drain for 20 sec; (7) washing with DMF, 2 x 20 sec and 1 x 2 min, 3 ml; washing with DCM 4 x 20 sec, 3 ml; (8) neutralization with DIEA/DCM (1:19 v/v), 2 x 3 min, 3 ml; (9) washing with DCM 4 x 20 sec, 3 ml, and drain for 1 min.; (10) qualitative Kaiser test; (11) blocking of unreacted amino groups by acetylation with Ac₂O/pyridine/DCM (1:1:2, v/v), 1 x 1/2 h, 3 ml; and (12) washing with DCM, 4 x 20 sec, 2 x 2

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min and 2 x 20 sec, 3 ml. Steps 1-12 were repeated until the desired sequence was obtained. All qualitative Kaiser tests were negative (straw-yellow colour with no coloration of the beads) indicating near 100% coupling yield. The PNA-oligomer 5 was cleaved and purified by the normal procedure. FAB-MS: 2832.11 [M+1] (calc. 2832.15)

EXAMPLE 56

Solid-Phase Synthesis of H-Taeg-Aaeg-[Taeg],-Lys-NH₂.

- 10 (a) Stepwise Assembly of Boc-Taeg-A(Z)aeg-[Taeg],-Lys(ClZ)-MBHA Resin.

About 0.3 g of wet Boc-[Taeg],-Lys(ClZ)-MBHA resin was placed in a 3 ml SPPS reaction vessel. Boc-Taeg-A(Z)aeg-[Taeg],-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC 15 coupling (single) of the A(Z)aeg residue utilizing 0.19 M of BocA(Z)aeg-OH together with 0.15 M DCC in 2.5 ml 50% DMF/CH₂Cl₂, and a single coupling with 0.15 M BocTaeg-OPfp in neat CH₂Cl₂ ("Synthetic Protocol 5"). The synthesis was monitored by the quantitative ninhydrin reaction, which showed about 50% 20 incorporation of A(Z)aeg and about 96% incorporation of Taeg.

- (b) Cleavage, Purification, and Identification of H-Taeg-Aaeg-[Taeg],-Lys-NH₂.

The protected Boc-Taeg-A(Z)aeg-[Taeg],-Lys(ClZ)-BAH resin was treated as described in Example 40c to yield about 25 15.6 mg of crude material upon HF cleavage of 53.1 mg dry H-Taeg-A(Z)aeg-[Taeg],-Lys(ClZ)-BHA resin. The main peak at 14.4 min accounted for less than 50% of the total absorbance. A 0.5 mg portion of the crude product was purified to give approximately 0.1 mg of H-Taeg-Aaeg-[Taeg],-Lys-NH₂. For 30 (MH⁺)⁺ the calculated m/z value was 2816.16 and the measured m/z value was 2816.28.

- (c) Synthetic Protocol 5

(1) Boc-deprotection with TFA/CH₂Cl₂, (1:1, v/v), 2.5 ml, 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 2.5 ml, 6 35 x 1 min; (3) neutralization with DIEA/CH₂Cl₂, (1: 19, v/v), 2.5 ml, 3 x 2 min; (4) washing with CH₂Cl₂, 2.5 ml, 6 x 1 min, and drain for 1 min; (5) 2-5 mg sample of PNA-resin is taken out

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and dried thoroughly for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 0.47 mmol (0.25 g) BocA(Z)aeg-OH dissolved in 1.25 ml DMF followed by addition of 0.47 mmol (0.1 g) DCC in 1.25 ml CH₂Cl₂, or 0.36 mmol (0.20 5 g) BocTaeg-OPfp in 2.5 ml CH₂Cl₂; the coupling reaction was allowed to proceed for a total of 20-24 hrs shaking; (7) washing with DMF, 2.5 ml, 1 x 2 min; (8) washing with CH₂Cl₂, 2.5 ml, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂, (1: 19, v/v), 2.5 ml, 2 x 2 min; (10) washing with CH₂Cl₂, 2.5 ml, 6 10 x 1 min; (11) 2-5 mg sample of protected PNA-resin is taken out and dried thoroughly for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted amino groups by acetylation with a 25 ml mixture of acetic anhydride/pyridine/CH₂Cl₂, (1:1:2, v/v/v) for 2 h (except 15 after the last cycle); and (13) washing with CH₂Cl₂, 2.5 ml, 6 x 1 min; (14) 2 x 2-5 mg samples of protected PNA-resin are taken out, neutralized with DIEA/CH₂Cl₂, (1: 19, v/v) and washed with CH₂Cl₂ for ninhydrin analyses.

20 EXAMPLE 57

Solid-Phase Synthesis of H-[Taeg],-Aaeg-[Taeg],-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg],-A(Z)aeg-[Taeg],-Lys(ClZ)-MBHA Resin.

About 0.5 g of wet Boc-[Taeg],-Lys(ClZ)-MBHA resin was 25 placed in a 5 ml SPPS reaction vessel. Boc-[Taeg],-A(Z)aeg-[Taeg],-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC coupling of both the A(Z)aeg and the Taeg residues utilising 0.15 M to 0.2 M of protected PNA monomer (free acid) together with an equivalent amount of DCC in 2 ml neat CH₂Cl₂ 30 ("Synthetic Protocol 6"). The synthesis was monitored by the quantitative ninhydrin reaction which showed a total of about 82% incorporation of A(Z)aeg after coupling three times (the first coupling gave about 50% incorporation; a fourth HOBt-mediated coupling in 50% DMF/CH₂Cl₂ did not increase the total 35 coupling yield significantly) and quantitative incorporation (single couplings) of the Taeg residues.

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(b) Cleavage, Purification, and Identification of H-[Taeg]₂-Aaeg-[Taeg]₅-Lys-NH₂.

The protected Boc-[Taeg]₂-A(Z)aeg-[Taeg]₅-Lys(ClZ)-BHA resin was treated as described in Example 40c to yield about 5 16.2 mg of crude material upon HF cleavage of 102.5 mg dry H-[Taeg]₂-A(Z)aeg-[Taeg]₅-Lys(ClZ)-BHA resin. A small portion of the crude product was purified. For (MH⁺)⁺, the calculated m/z value was 2050.85 and the measured m/z value was 2050.90

(c) Synthetic Protocol 6

10 (1) Boc-deprotection with TFA/CH₂Cl₂, (1:1, v/v), 2 ml, 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 2 ml, 6 x 1 min; (3) neutralization with DIEA/CH₂Cl₂, (1: 19, v/v), 2 ml, 3 x 2 min; (4) washing with CH₂Cl₂, 2 ml, 6 x 1 min, and drain for 1 min; (5) 2-5 mg sample of PNA-resin was taken out and 15 dried thoroughly for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 0.44 mmol (0.23 g) BocA(Z)aeg-OH dissolved in 1.5 ml CH₂Cl₂, followed by addition of 0.44 mmol (0.09 g) DCC in 0.5 ml CH₂Cl₂, or 0.33 mmol (0.13 g) BocTaeg-OH in 1.5 ml CH₂Cl₂, followed by addition 20 of 0.33 mmol (0.07 g) DCC in 0.5 ml CH₂Cl₂; the coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking; (7) washing with DMF, 2 ml, 1 x 2 min; (8) washing with CH₂Cl₂, 2 ml, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂, (1: 19, v/v), 2 ml, 2 x 2 min; (10) washing with 25 CH₂Cl₂, 2 ml, 6 x 1 min; (11) 2-5 mg sample of protected PNA-resin is taken out and dried thoroughly for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted amino groups by acetylation with a 25 ml mixture of acetic anhydride/pyridine/CH₂Cl₂, (1:1:2, v/v/v) 30 for 2 h (except after the last cycle); (13) washing with CH₂Cl₂, 2 ml, 6 x 1 min; and (14) 2 x 2-5 mg samples of protected PNA-resin were taken out, neutralized with DIEA/CH₂Cl₂, (1: 19, v/v) and washed with CH₂Cl₂ for ninhydrin analyses.

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EXAMPLE 58

The PNA-oligomer H-T₄C₂TCT-LysNH₂ was prepared as described in Example 93. Hybridization experiments with this sequence should resolve the issue of orientation, since it is truly asymmetrical. Such experiments should also resolve the issues of pH-dependency of the Tm, and the stoichiometry of complexes formed.

Hybridization experiments with the PNA-oligomer H-T₄C₂TCTC-LysNH₂ were performed as follows:

10

Row	Hybridized With	pH	Tm	S
1	5'-(dA), (dG), (dA) (dG) (dA) (dG)	7.2	55.5	2:1
2	5'-(dA), (dG), (dA) (dG) (dA) (dG)	9.0	26.0	2:1
3	5'-(dA), (dG), (dA) (dG) (dA) (dG)	5.0	88.5	2:1
4	5'-(dG) (dA) (dG) (dA) (dG), (dA),	7.2	38.0	2:1
5	5'-(dG) (dA) (dG) (dA) (dG), (dA),	9.0	31.5	-
6	5'-(dG) (dA) (dG) (dA) (dG), (dA),	5.0	52.5	-
7	5'-(dA), (dG) (dT) (dA) (dG) (dA) (dG)	7.2	39.0	-
8	5'-(dA), (dG) (dT) (dA) (dG) (dA) (dG)	9.0	<20	-
9	5'-(dA), (dG) (dT) (dA) (dG) (dA) (dG)	5.0	51.5	-
10	5'-(dA), (dG), (dT) (dG) (dA) (dG)	7.2	31.5	-
11	5'-(dA), (dG), (dT) (dG) (dA) (dG)	5.0	50.5	-
12	5'-(dG) (dA) (dG) (dA) dT) (dG) (dA),	7.2	24.5	-
13	5'-(dG) (dA) (dG) (dA) dT) (dG) (dA),	9.0	<20	-
14	5'-(dG) (dA) (dG) (dA) dT) (dG) (dA),	5.0	57.0	-
15	5'-(dG) (dA) (dG) (dT) (dG), (dA),	7.2	25.0	-
16	5'-(dG) (dA) (dG) (dT) (dG), (dA),	5.0	39.5	-
			52.0	

' = stoichiometry determined by UV-mixing curves

30 - = not determined

These results show that a truly mixed sequence gave rise to well defined melting curves. The PNA-oligomers can actually bind in both orientations (compare row 1 and 4), although there is preference for the N-terminal/5'-

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orientation. Introducing a single mismatch opposite either T or C caused a lowering of T_m by more than 16°C at pH 7.2; at pH 5.0 the T_m -value was lowered more than 27°C . This shows that there is a very high degree a sequence-selectivity which 5 should be a general feature for all PNA C/T sequences.

As indicated above, there is a very strong pH-dependency for the T_m -value, indicating that Hoogsteen basepairing is important for the formation of hybrids. Therefore, it is not surprising that the stoichiometry was 10 found to be 2:1.

The lack of symmetry in the sequence and the very large lowering of T_m when mismatches are present show that the Watson-Crick strand and the Hoogsteen strand are parallel when bound to complementary DNA. This is true for both of the 15 orientations, i.e., 5'/N-terminal and 3'/N-terminal.

EXAMPLE 59

The results of hybridization experiments with H-T₄GT₄-LysNH₂ to were performed as follows:

Row	Deoxyoligonucleotide	T_m
1	5'-(dA)5(dA)(dA)4-3'	55.0
2	5'-(dA)5(dG)(dA)4-3'	47.0
3	5'-(dA)5(dG)(dA)4-3'	56.5
4	5'-(dA)5(dT)(dA)4-3'	46.5
5	5'-(dA)4(dG)(dA)5-3'	48.5
6	5'-(dA)4(dC)(dA)5-3'	55.5
7	5'-(dA)4(dT)(dA)5-3'	47.0

As shown by comparing rows 1, 3, and 6 with rows 2, 4, 30 5, and 7, G can in this mode discriminate between C/A and G/T in the DNA-strand, i.e., sequence discrimination is observed. The complex in row 3 was furthermore determined to be 2 PNA: 1 DNA complex by UV-mixing curves.

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EXAMPLE 60

The masses of some synthesized PNA-oligomers, as determined by FAB mass spectrometry, are as follows:

	SEQUENCE	CALC.	FOUND
5	H-T ₄ C ₂ TCTC-LysNH ₂	2747.15	2746.78
	H-T ₅ GT ₄ -LysNH ₂	2832.15	2832.11
	H-T ₇ -LysNH ₂	2008.84	2540.84
	H-T ₉ -LysNH ₂	2541.04	2540.84
	H-T ₁₀ -LysNH ₂	2807.14	2806.69
	H-T ₂ CT ₅ -LysNH ₂	2259.94	2259.18
	H-T ₃ (L-alalT)T ₄ -LysNH ₂	2287.95	2288.60
	H-T ₄ (Ac)T ₅ -LysNH ₂	2683.12	2683.09

EXAMPLE 61

15 Hybridization data for a PNA-oligomer with a single unit with an extended backbone (the β -alanine modification) is as follows:

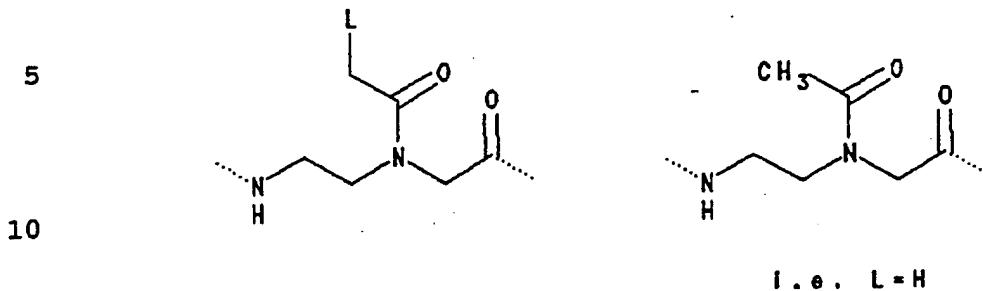
	PNA	DNA	T _m
20	H-T ₁₀ -LysNH ₂	(dA) ₁₀	73 °C
	H-T ₄ (β T)T ₅ -LysNH ₂	(dA) ₁₀	57 °C
	H-T ₄ (β T)T ₅ -LysNH ₂	(dA) ₄ (dG)(dA) ₅	47 °C
	H-T ₄ (β T)T ₅ -LysNH ₂	(dA) ₄ (dT)(dA) ₅	49 °C
	H-T ₄ (β T)T ₅ -LysNH ₂	(dA) ₄ (dT)(dA) ₅	47 °C

25 Although the melting temperature decreases, the data demonstrates that base specific recognition is retained.

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EXAMPLE 62

An example with a "no base" substitution.



15

	PNA	DNA	T _m
20	H-T ₁₀ -LysNH ₂	(dA) ₁₀	73 °C
	H-T ₄ (AC)T ₅ -LysNH ₂	(dA) ₁₀	49 °C
	H-T ₄ (AC)T ₅ -LysNH ₂	(dA) ₄ (dG)(dA) ⁵	37 °C
	H-T ₄ (AC)T ₅ -LysNH ₂	(dA) ₄ (dC)(dA) ⁵	41 °C
	H-T ₄ (AC)T ₅ -LysNH ₂	(dA) ₄ (dT)(dA) ⁵	41 °C
	H-T ₄ (AC)T ₅ -LysNH ₂	(dA) ₅ (dG)(dA) ⁴	36 °C
	H-T ₄ (AC)T ₅ -LysNH ₂	(dA) ₅ (dC)(dA) ⁴	40 °C
	H-T ₄ (AC)T ₅ -LysNH ₂	(dA) ₅ (dT)(dA) ⁴	40 °C

EXAMPLE 63

Iodination Procedure

30 A 5 µg portion of Tyr-PNA-T₁₀-Lys-NH₂ is dissolved in
 40 µl 100 mM Na-phosphate, pH 7.0, and 1 mCi Na¹²⁵I and 2 µl
 chloramine-T (50 mM in CH₃CN) are added. The solution is left
 at 20°C for 10 min and then passed through a 0.5 + 5 cm
 Sephadex G10 column. The first 2 fractions (100 µl each)
 35 containing radioactivity are collected and purified by HPLC:
 reversed phase C-18 using a 0-60% CH₃CN gradient in 0.1%
 CF₃COOH in H₂O. The ¹²⁵I-PNA elutes right after the PNA peak.
 The solvent is removed under reduced pressure.

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EXAMPLE 64

Binding of PNAS-T₁₀/T₉C/T₈C₂ to double stranded DNA targets A₁₀/A₉G/A₈G₂ (Figure 20).

A mixture of 200 cps ³²P-labeled EcoRI-PvuII fragment 5 (the large fragment labeled at the 3'-end of the EcoRI site) of the indicated plasmid, 0.5 µg carrier calf thymus DNA, and 300 ng PNA in 100 µl buffer (200 mM NaCl, 50 mM Na-acetate, pH 4.5, 1 mM ZnSO₄) was incubated at 37°C for 120 min. A 50 unit portion of nuclease S₁ was added and incubated at 20°C 10 for 5 min. The reaction was stopped by addition of 3 µl 0.5 M EDTA and the DNA was precipitated by addition of 250 µl 2% potassium acetate in ethanol. The DNA was analyzed by electrophoresis in 10% polyacrylamide sequencing gels and the radiolabeled DNA bands visualized by autoradiography.

15 The target plasmids were prepared by cloning of the appropriate oligonucleotides into pUC19. Target A₁₀: oligonucleotides GATCCA₁₀G & GATCCT₁₀G cloned into the BamHI site (plasmid designated pT10). Target A₅GA₄: oligonucleotides TCGACT₄CT₅G & TCGACA₅GA₄G cloned into the SalI site (plasmid 20 pT9C). Target A₂GA₂GA₄: oligonucleotides GA₂GA₂GA₄TGCA & GT₄CT₂CT₂CTGCA into the PstI site (plasmid pT8C2). The positions of the targets in the gel are indicated by bars to the left. A/G is an A+G sequence ladder of target P10.

25 **EXAMPLE 65**

Inhibition of restriction enzyme cleavage by PNA (Figure 23).

A 2 µg portion of plasmid pT10 was mixed with the indicated amount of PNA-T₁₀ in 20 µl TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 7.4) and incubated at 37°C for 120 min. 2 30 µl 10 × buffer (10 mM Tris-HCl, pH 7.5, 10 mM, MgCl₂, 50 mM NaCl, 1 mM DTT). PvuII (2 units) and BamHI (2 units) were added and the incubation was continued for 60 min. The DNA was analyzed by gel electrophoresis in 5% polyacrylamide and the DNA was visualized by ethidium bromide staining.

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EXAMPLE 66

Kinetics of PNA-T₁₀ - dsDNA strand displacement complex formation (Figure 21).

A mixture of 200 cps ³²P-labeled EcoRI-PvuII fragment 5 of pT10 (the large fragment labeled at the 3'-end of the EcoRI site), 0.5 µg carrier calf thymus DNA, and 300 ng of PNA-T₁₀-LysNH₂ in 100 µl buffer (200 mM NaCl, 50 mM Na-acetate, pH 4.5, 1 mM ZnSO₄) were incubated at 37°C. At the times indicated, 50 U of S₁ nuclease was added to each of 7 samples 10 and incubation was continued for 5 min at 20°C. The DNA was then precipitated by addition of 250 µl 2% K-acetate in ethanol and analyzed by electrophoresis in a 10% polyacrylamide sequencing gel. The amount of strand displacement complex was calculated from the intensity of the S₁-cleavage 15 at the target sequence, as measured by densitometric scanning of autoradiographs.

EXAMPLE 67

Stability of PNA-dsDNA complexes (Figure 22).

20 A mixture of 200 cps ³²P-pT10 fragment, 0.5 µg calf thymus DNA and 300 ng of the desired PNA (either T₁₀-LysNH₂, T₈-LysNH₂ or T₆-LysNH₂) was incubated in 100 µl 200 mM NaCl, 50 mM Na-acetate, pH 4.5, 1 mM ZnSO₄ for 60 min at 37°C. A 2 µg portion of oligonucleotide GATCCA₁₀G was added and each sample 25 was heated for 10 min at the temperature indicated, cooled in ice for 10 min and warmed to 20°C. A 50 U portion of S₁ nuclease was added and the samples treated and analyzed and the results quantified.

30 **EXAMPLE 68**

Inhibition of Transcription by PNA

A mixture of 100 ng plasmid DNA (cleaved with restriction enzyme PvuII (see below) and 100 ng of PNA in 15 µl 10 mM Tris-HCl, 1 mM EDTA, pH 7.4 was incubated at 37°C for 35 60 min. Subsequently, 4 µl 5 × concentrated buffer (0.2 M Tris-HCl (pH 8.0), 40 mM MgCl₂, 10 mM spermidine, 125 mM NaCl) were mixed with 1 µl NTP-mix (10 mM ATP, 10 mM CTP, 10 mM GTP,

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1 mM UTP, 0.1 μ Ci/ μ l 32 P-UTP, 5 mM DTT, 2 μ g/ml tRNA, 1 μ g/ml heparin) and 3 units RNA polymerase. Incubation was continued for 10 min at 37°C. The RNA was then precipitated by addition of 60 μ l 2% postassium acetate in 96% ethanol at -20°C and 5 analyzed by electrophoresis in 8% polyacrylamide sequencing gels. RNA transcripts were visualized by autoradiography. The following plasmids were used: pT8C2-KS/pA8G2-KS: oligonucleotides GA₂GA₂GA₄GTGAC & GT₄CT₂CT₂CTGCA cloned into the PstI site of pBluescript-KS⁺; pT10-KS/pA10-KS (both 10 orientations of the insert were obtained). pT10UV5: oligonucleotides GATCCA₁₀G & GATCCT₁₀G cloned into the BamHI site of a pUC18 derivative in which the lac UV5 E.coli promoter had been cloned into the EcoRI site (Jeppesen, et al., Nucleic Acids Res., 1988, 16, 9545).

15 Using T₃-RNA polymerase, transcription elongation arrest was obtained with PNA-T₈C₂-LysNH₂ and the pA8G2-KS plasmid having the PNA recognition sequence on the template strand, but not with pT8C2-KS having the PNA recognition sequence on the non-template strand. Similar results were 20 obtained with PNA-T10-LysNH₂ and the plasmids pA10-KS and pT10-KS. (see, Figure 25) Using E.coli RNA polymerase and the pT10UV5 plasmid (A₁₀-sequence on the template strand) transcription elongation arrest was obtained with PNA-T₁₀-LysNH₂.

25

EXAMPLE 69

Biological stability of PNA

A mixture of PNA-T₅ (10 μ g) and a control, "normal" peptide (10 μ g) in 40 μ l 50 mM Tris-HCl, pH 7.4 was treated 30 with varying amounts of peptidase from porcine intestinal mucosa or protease from Streptomyces caespitiosus for 10 min at 37°C. The amount of PNA and peptide was determined by HPLC analysis (reversed phase C-18 column: 0-60% acetonitrile, 0.1% trifluoroacetic acid).

35 At peptidase/protease concentrations where complete degradation of the peptide was observed (no HPLC peak) the PNA was still intact.

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EXAMPLE 70

Inhibition of Gene Expression

A preferred assay to test the ability of peptide nucleic acids to inhibit expression of the E2 mRNA of 5 papillomavirus is based on the well-documented transactivation properties of E2. Spalholz, et al., *J. Virol.*, 1987, 61, 2128-2137. A reporter plasmid (E2RECAT) was constructed to contain the E2 responsive element, which functions as an E2 dependent enhancer. E2RECAT also contains the SV40 early 10 promoter, an early polyadenylation signal, and the chloramphenicol acetyl transferase gene (CAT). Within the context of this plasmid, CAT expression is dependent upon expression of E2. The dependence of CAT expression on the presence of E2 has been tested by transfection of this plasmid 15 into C127 cells transformed by BPV-1, uninfected C127 cells and C127 cells cotransfected with E2RECAT and an E2 expression vector.

A. Inhibition of BPV-1 E2 Expression

BPV-1 transformed C127 cells are plated in 12 well 20 plates. Twenty four hours prior to transfection with E2RE1, cells are pretreated by addition of antisense PNAs to the growth medium at final concentrations of 5, 15 and 30 mM. The next day cells are transfected with 10 µg of E2RE1CAT by calcium phosphate precipitation. Ten micrograms of E2RE1CAT 25 and 10 µg of carrier DNA (PUC 19) are mixed with 62 µl of 2 M CaCl₂ in a final volume of 250 µl of H₂O, followed by addition of 250 µl of 2X HBSP (1.5 mM Na₂PO₄, 10 mM KCl, 280 mM NaCl, 12 mM glucose and 50 mM HEPES, pH 7.0) and incubated at room temperature for 30 minutes. One hundred microliters 30 of this solution is added to each test well and allowed to incubate for 4 hours at 37°C. After incubation, cells are glycerol shocked for 1 minute at room temperature with 15% glycerol in 0.75 mM Na₂PO₄, 5 mM KCl, 140 mM NaCl, 6 mM glucose and 25 mM HEPES, pH 7.0. After shocking, cells are 35 washed 2 times with serum free DMEM and refed with DMEM containing 10% fetal bovine serum and antisense oligonucleotide at the original concentration. Forty eight

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hours after transfection cells are harvested and assayed for CAT activity.

For determination of CAT activity, cells are washed 2 times with phosphate buffered saline and collected by scraping. Cells are resuspended in 100 μ l of 250 mM Tris-HCl, pH 8.0 and disrupted by freeze-thawing 3 times. Twenty four microliters of cell extract is used for each assay. For each assay the following are mixed together in an 1.5 ml Eppendorf tube and incubated at 37°C for one hour: 25 μ l of cell extract, 5 μ l of 4 mM acetyl coenzyme A, 18 μ l H₂O and 1 μ l ¹⁴C-chloramphenicol, 40-60 mCi/mM. After incubation, chloramphenicol (acetylated and nonacetylated forms) is extracted with ethyl acetate and evaporated to dryness. Samples are resuspended in 25 μ l of ethyl acetate, spotted onto a TLC plate and chromatographed in chloroform:methanol (19:1). Chromatographs are analyzed by autoradiography. Spots corresponding to acetylated and nonacetylated ¹⁴C-chloramphenicol are excised from the TLC plate and counted by liquid scintillation for quantitation of CAT activity. Peptide nucleic acids that depress CAT activity in a dose dependent fashion are considered positives.

B. Inhibition of HPV E2 Expression

The assay for inhibition of human papillomavirus (HPV) E2 by peptide nucleic acids is essentially the same as that for BPV-1 E2. For HPV assays appropriate HPVs are co-transfected into either CV-1 or A431 cells with PSV2NEO using the calcium phosphate method described above. Cells which take up DNA are selected for by culturing in media containing the antibiotic G418. G418-resistant cells are then analyzed for HPV DNA and RNA. Cells expressing E2 are used as target cells for antisense studies. For each PNA, cells are pretreated as above, transfected with E2RE1CAT, and analyzed for CAT activity as above. Peptide nucleic acids are considered to have a positive effect if they can depress CAT activity in a dose dependent fashion.

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EXAMPLE 71

Synthesis of PNA 15-mer Containing Four Naturally Occurring Nucleobases; H-[Taeg]-[Aaeg]-[Gaeg]-[Taeg]-[Taeg]-[Aaeg]-[Taeg]-[Caeg]-[Taeg]-[Caeg]-[Taeg]-[Aaeg]-[Taeg]-[Caeg]-
5 [Taeg]-LYS-NH₂.

The protected PNA was assembled onto a Boc-Lys(Clz) modified MBHA resin with a substitution of approximately 0.145 mmol/g. Capping of uncoupled amino groups was only carried out before the incorporation of the BocGaeg-OH monomer.

10 Synthesis was initiated on 100 mg (dry weight) of neutralised Boc-Lys(ClA)-MBHA resin that had been preswollen overnight in DCM. The incorporation of the monomers followed the protocol of Example 32, except at step 5 for the incorporation of the BocAaeg-OH monomer. Step 5 for the
15 present synthesis involved addition of 4 equiv. diisopropyl carbodiimide (0.06 ml; 9.7 µl) and 4 equiv. BocAaeg-OH (0.06 mmol; 32 mg) dissolved in 0.6 ml DCM/DMF (1:1, v/v) (final concentration of monomer 0.1M). The coupling reaction was allowed to proceed for 1 x 15 min and 1 x 60 min.
20 (recoupling).

All qualitative Kaiser tests were negative (straw-yellow color with no coloration of the beads). The PNA-oligomer was cleaved and purified by the standard procedure. FAB-MS average mass found(calc.) (M+H) 4145.1 (4146.1).

25

EXAMPLE 72

Hybridization of H-TAGTTATCTCTATCT-LysNH₂

DNA -target	pH	Tm
5'----3'	5	60.5
30 5'----3'	7.2	43.0
5'----3'	9	38.5
3'----5'	5	64.5/49.0
3'----5'	7.2	53.5
3'----5'	9	51.5

35

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The fact that there is almost no loss in T_m in going from pH 7.2 to 9.0 indicates that Hoogsteen basepairing is not involved. The increase in T_m in going from 7.2 to 5 is large for the parallel orientation and is probably due to the formation of a 2:1 complex. It is believed that the most favorable orientation in the Watson-Crick binding motif is the 3'/N-orientation and that in the Hoogsteen motif the 5'/N-orientation is the most stable. Thus, it may be the case that the most stable complex is with the two PNA's strands anti parallel.

There is apparently a very strong preference for a parallel orientation of the Hoogsteen strand. This seems to explain why even at pH 9 a 2:1 complex is seen with the 5'/N-orientation. Furthermore, it explains the small loss in going from pH 7.2 to 9 in the 3'/N, as this is probably a 1:1 complex.

EXAMPLE 73

Solid-Phase Synthesis of H-[Taeg]₂-Aaeg-Taeg-Caeg-Aaeg-Taeg-
20 Caeg-Taeg-Caeg-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]2-A(Z)aeg-Taeg-C(Z)aeg-A(Z)aeg-Taeg-C(Z)aeg-Taeg-C(Z)aeg-Lys(C1Z)-MBHA Resin.

About 1 g of wet Boc-Lys(C1Z)-MBHA (0.28 mmol Lys/g) resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]2-A(Z)aeg-Taeg-C(Z)aeg-A(Z)aeg-Taeg-C(Z)aeg-Taeg-C(Z)aeg-Lys(C1Z)-MBHA resin was assembled by *in situ* DCC coupling of the five first residues utilizing 0.16 M of BocC[Z]-OH, BocTaeg-OH or BocA(Z)aeg-OH, together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9") and by analogous *in situ* DIC coupling of the five last residues ("Synthetic Protocol 10"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored by the ninhydrin reaction, which showed nearly quantitative incorporation of all residues except of the first A(Z)aeg residue, which had to be coupled twice. The total

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coupling yield was about 96% (first coupling, about 89% efficiency).

(b) Cleavage, Purification, and Identification of H-[Taeg]2-Aaeg-Taeg-Caeg-Aaeg-Taeg-Caeg-Taeg-Caeg-Lys-NH₂.

5 The protected Boc-[Taeg]2-A(Z)aeg-Taeg-C(Z)aeg-A(Z)aeg-Taeg-C(Z)aeg-Taeg-C(Z)aeg-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 53.4 mg of crude material upon HF cleavage of 166.1 mg dry Boc-[Taeg]2-A(Z)aeg-Taeg-C(Z)aeg-A(Z)aeg-Taeg-C(Z)aeg-Taeg-C(Z)aeg-Lys(ClZ)-MBHA 10 resin. The crude product (53.4 mg) was purified to give 18.3 mg of H-[Taeg]2-Aaeg-Taeg-Caeg-Aaeg-Taeg-Caeg-Taeg-Caeg-Lys-NH₂. For (M+H)⁺, the calculated m/z value = 2780.17 and the measured m/z value = 2780.07.

15 EXAMPLE 74

Hybridization properties of H-TTA TCA TCT C-Lys-NH₂.

The title compound hybridized with the following oligonucleotides:

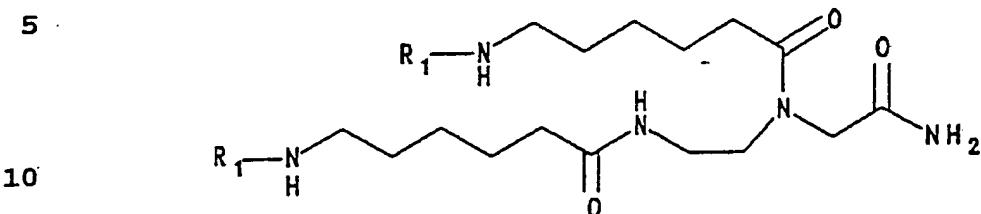
Oligodeoxynucleotide	pH	T _m (°C)
20 5'-AAT AGT AGT G-3'	5	31.5†
5'-ATT AGT AGT G-3'	7.2	28.5†
5'-AAT AGT AGT G-3"	9	28.0†
5'-GTG ATG ATA A-3'	7.2	30.5
25 5'-GTG ATG ATA A-3'	9	28.0

25 †Low hypochromicity

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EXAMPLE 75

Synthesis of a PNA With Two Parallel Strings Tied Together



A 375 mg portion of MBHA resin (loading 0.6 mmol/g) was
15 allowed to swell over night in dichloromethane (DCM). After
an hour in DMF/DCM, the resin was neutralized by washing 2
times with 5% diisopropylethylamine in DCM (2 min.), followed
by washing with DCM (2ml; 6 x 1 min.) N,N'-di-Boc-aminoethyl
glycine (41,9 mg; 0,132 mmol) dissolved in 2 ml DMF was added
20 to the resin, followed by DCC (64,9 mg; 0,315 mmol) dissolved
in 1 ml of DCM. After 2.5 hours, the resin was washed with
DMF 3 times (1 min.) and once with DCM (1 min.). The
unreacted amino groups were then capped by treatment with
acetic anhydride/DCM/pyridine (1 ml\2 ml\2 ml) for 72 hours.
25 After washing with DCM (2 ml; 4 x 1 min), a Kaiser test showed
no amino groups were present. The resin was deprotected and
washed as described above. This was followed by reaction with
6-(Bocamino)-hexanoic acid DHBT ester (255.8 mg; 67 mmol)
dissolved in DMF/DCM 1:1 (4 ml) overnight. After washing and
30 neutralization, a Kaiser test and an isatin test were
performed. Both were negative. After capping, the
elongation of the PNA-chains was performed according to
standard procedures for DCC couplings. All Kaiser tests
performed after the coupling reactions were negative (Yellow).
35 Qualitative Kaiser tests were done after deprotection of PNA
units number 1, 2, 4, and 6. Each test was blue. The PNA
oligomers were cleaved and purified by standard procedures.

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The amount of monomer and DCC used for each coupling was as follows (total volume 4.5 ml):

	Coupling	Monomer (T)	DCC
5	1.	173 mg	95 mg
	2.	176 mg	101 mg
	3.	174 mg	97 mg
	4.	174 mg	103 mg
	5.	178 mg	97 mg
	6.	173 mg	99 mg
	7.	174 mg	95 mg
	8.	175 mg	96 mg

15 For the PNA having the Structure (70) where $R_{70} = T_6$, there was 24.5 mg of crude product, which resulted in 6.9 mg. after purification. For the PNA where $R_1 = T_8$, there was 28.8 mg of crude product, which resulted in 2.8 mg. after purification. The products had a high tendency of
20 aggregation, as indicated by a complex HPLC chromatogram after a few hours at room temperature in concentration above 1 mg/ml. The PNA-(T_6)₂ and PNA-(T_8)₂ were hybridised to (dA)₆ and (dA)₈, respectively, with recorded T_m of 42°C and 59°C, respectively.

25

EXAMPLE 76

Solid-Phase Synthesis of H-[Taeg]_n-Lys(ClZ)-MBHA Resin

The PNA oligomer was assembled onto 500 mg (dry weight) of MBHA resin that had been preswollen overnight in DCM. The
30 resin was initially substituted with approximately 0.15 mmol/g Boc-Lys(ClZ) as determined by quantitative ninhydrin reaction. The stepwise synthesis of the oligomer followed the synthetic protocol described in Example 32 employing 0.077 g (0.2 mmol) BocTaeg-OH and 31.3 μ l (0.2 mmol) diisopropyl carbodiimide in
35 2.0 ml 50% DMF/CH₂Cl₂ in each coupling. Capping of uncoupled amino groups was carried out before deprotection in each step. All qualitative Kaiser tests were negative indicating near 100% coupling yield.

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EXAMPLE 77

Solid-Phase Synthesis of H-[Taeg]₄-[apgT]-[Taeg]₂Lys-NH₂

Synthesis was initiated on approximately 1/4 of the wet H-[Taeg]₅-Lys(ClZ)-MBHA resin from Example 76. *In situ* diisopropyl carbodiimide (DIC) couplings of both Boc-(apgT)-OH and BocTaeg-OH were carried out in 1.2 ml 50% DMF/CH₂Cl₂ using 0.048 g (0.12 mmol) and 0.046 g (0.12 mmol) monomer, respectively, and 18.7 μ l (0.12 mmol) diisopropyl carbodiimide in each coupling. All qualitative Kaiser tests were negative, indicating near 100% coupling yield. The PNA oligomer was cleaved and purified by standard procedures. For (M+H)⁺, the calculated m/z value was 2820.15 and the measured m/z value was 2820.92.

15 EXAMPLE 78

Solid-Phase Synthesis of H-[Taeg]₄-[proT]-[Taeg]₂-Lys-NH₂

Synthesis was initiated on approximately 1/4 of the wet H-[Taeg]₅-Lys(ClZ)-MBHA resin from Example 76. *In situ* diisopropyl carbodiimide couplings of BocTaeg-OH were carried out in 1.2 ml 50% DMF/CH₂Cl₂ using 0.046 g (0.12 mmol) monomer and 18.7 μ l (0.12 mmol) diisopropyl carbodiimide in each coupling. Due to solubility problems, Boc-(proT)-OH 0.048 g (0.12 mmol) was suspended in 2.5 ml 50% DMF/DMSO prior to coupling, the suspension filtered, and approximately 2 ml of the filtrate used in the overnight coupling. All qualitative Kaiser tests were negative, indicating near 100% coupling yield. The PNA oligomer was cleaved and purified by standard procedures.

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EXAMPLE 79

Hybridization properties of H-[Taeg]-[proT]-[Taeg]s.Lys-NH₂

	Oligodeoxynucleotide	Tm (°C)
5	5'-AAA AAA AAA A	53.5
	5'-AAA AGA AAA A	44.0
	5'-AAA AAG AAA A	43.5
	5'-AAA ACA AAA A	46.5
	5'-AAA AAC AAA A	46.5
	5'-AAA ATA AAA A	46.5
10	5'-AAA AAT AAA A	46.0

EXAMPLE 80

Solid-Phase Synthesis of H-[Taeg]-[bC]-[Taeg]-Lys-NH₂

15 The PNA oligomer was assembled onto 100 mg (dry weight) MBHA resin that had been preswollen overnight in DCM. The resin was initially substituted with approximately 0.25 mmol/g Boc-Lys(ClZ) as determined by quantitative ninhydrin reaction. The stepwise synthesis of the oligomer followed synthetic
 20 Protocol 9 employing 0.023 g (0.06 mmol) BocTaeg-OH, 0.062 g (0.12 mmol) BocbC(Z)-OH and 0.012 g (0.06 mmol) DCC in 1.2 ml 50% DMF/CH₂Cl₂ in each coupling. Capping of uncoupled amino groups was carried out before deprotection in each step. All qualitative Kaiser tests were negative, indicating near 100%
 25 coupling yield. The PNA-oligomer was cleaved and purified by standard procedures.

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EXAMPLE 81

Hybridization properties of H-T,bCT,-Lys-NH₂

5	Oligodeoxynucleotide	Tm (°C)
	5'-AAA AAA AAA A	43.5
	5'-AAA AGA AAA A	58.0
	5'-AAA AAG AAA A	60.0
	5'-AAA ACA AAA A	34.5
10	5'-AAA AAC AAA A	34.5
	5'-AAA ATA AAA A	34.0
	5'-AAA AAT AAA A	36.0

EXAMPLE 82

15 Stepwise Assembly of H-[Taeg]-[Taeg]-[Taeg]-[Taeg]-[Aaeg]-[Taeg]-[Taeg]-[Taeg]-[Taeg]-LYS-NH₂.

Synthesis was initiated on a Boc-[Taeg],-Lys(ClZ)-MBHA resin (from example 76) that had been preswollen overnight in DCM. The resin resembled approximately 100 mg (dry weight) of Boc-Lys(ClZ)-MBHA resin (loading 0.15 mmol/g). The incorporation of the monomers followed the protocol of example 55, except for step 5 (incorporation of the BocA(Z)aeg-OH monomer). New step 5 (incorporation of A(Z)aeg) involved addition of 4 equiv. diisopropyl carbodiimide (0.06 mmol; 9.7 μ l) and 4 equiv. BocA(Z)aeg-OH (0.06 mmol; 32 mg) dissolved in 0.6 ml DCM/DMF (1:1, v/v) (final concentration of monomer 0.1 M). The coupling reaction was allowed to proceed for 1 x 15 min. and 1 x 60 min. (recoupling).

Capping of uncoupled amino groups was only carried out before the incorporation of the BocA(Z)aeg-OH monomer. The coupling reaction was monitored by qualitative ninhydrin reaction (Kaiser test). All qualitative Kaiser tests were negative (straw-yellow color with no coloration of the beads). The PNA oligomer was cleaved and purified by standard procedures.

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EXAMPLE 84

Hybridization properties of H-T₄AT₅-LysNH₂

	Oligodeoxynucleotide	Tm (°C)
5	5'-AAA AAA AAA A	59.5
	5'-AAA AGA AAA A	45.0
	5'-AAA AAG AAA A	45.5
	5'-AAA ACA AAA A	48.0
	5'-AAA AAC AAA A	48.0
	5'-AAA ATA AAA A	52.0
10	5'-AAA AAT AAA A	52.5

EXAMPLE 85

Stepwise Assembly of H-[Taeg]-[Taeg]-[Taeg]-[Taeg]-[Gaeg]-
15 [Gaeg]-[Taeg]-[Gaeg]-[Taeg]-[Gaeg]-LYS-NH₂.

The protected PNA was assembled onto a Boc-Lys(C1Z) modified MBHA resin with a substitution of 0.15 mmol/g. The incorporation of the monomers followed the protocol of example 32, except that the capping step 11 and the washing step 12 were omitted. After the incorporation and deprotection of the first, second, and fourth G(Bz1)aeg-monomer there were some difficulties getting the resin to swell properly. Three hours of shaking in neat DCM gave acceptable swelling. For the incorporation of residues Taeg-4, G(Bz1)aeg-6, and Taeg-7 to 25 Taeg-10, recoupling was necessary to obtain near quantitative coupling yields. Taeg₄ (2 x in 50% DMF/DCM), Gaeg₆ (2 x in 50% DMF/DCM), Taeg₇ (2 x in 50% DMF/DCM, 1 x in 50% NMP/DCM and 1 x in neat DCM), Taeg₈ (1 x in 50% DMF/DCM and 2 x in neat DCM), Taeg₉ (2 x in 50% DMF/DCM), Taeg₁₀ (2 x in 50% DMF/DCM). 30 All qualitative Kaiser tests were negative (straw-yellow color with no coloration of the beads). The PNA oligomer was cleaved and purified by standard procedures

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EXAMPLE 86

Hybridization properties of crude (approx. 50%) H-T₄G₂TGTG-LysNH₂

	Oligodeoxynucleotide	Tm
5	5'-A4C2ACAC	38
	5'-CACAC2A4	55

EXAMPLE 87

10 Large Scale Solid-Phase Synthesis of H-[Taeg]_n-Lys-NH₂, H-[Taeg]_n-Lys-NH₂, H-[Taeg]_n-Lys-NH₂, H-[Taeg]_n-Lys-NH₂, and H-[Taeg]_n-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]_n-Lys(ClZ)-MBHA Resin and Shorter Fragments.

15 About 9 g of wet Boc-[Taeg]_n-Lys(ClZ)-MBHA (see, Example 19b) resin was placed in a 60 ml SPPS reaction vessel. Boc-[Taeg]_n-Lys(ClZ)-MBHA resin was assembled by single coupling of both residues with 0.15 M of BocTaeg-OPfp in 10 ml neat CH₂Cl₂ ("Synthetic Protocol 8"). Both coupling reactions were allowed to proceed overnight. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of both residues. After deprotection of the N-terminal Boc group, about 4.5 g of H-[Taeg]_n-Lys(ClZ)-MBHA was placed in a 20 ml SPPS reaction vessel and elongated to Boc-[Taeg]_n-Lys(ClZ)-MBHA by single *in situ* DCC coupling of all residues (close to quantitative, except for residue number eight) overnight with 0.2 M of BocTaeg-OH together with 0.2 M DCC in 7.5 ml neat CH₂Cl₂ ("Synthetic Protocol 9"). Before coupling of Taeg residues number seven and eight, respectively, small portions of H-[Taeg]_n-Lys(ClZ)-MBHA and H-[Taeg]_n-Lys(ClZ)-MBHA, respectively, were taken out for HF cleavage.

30 Taeg residue number eight was coupled twice (overnight) to give close to quantitative incorporation. After 35 deprotection of the N-terminal Boc group, a large portion of H-[Taeg]_n-Lys(ClZ)-MBHA was taken out for HF cleavage. Boc-

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[Taeg]₁₀-Lys(ClZ)-MBHA resin was assembled by double *in situ* DCC coupling of 0.16 M BocTaeg-OH, together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂, ("Synthetic Protocol" 9). Before coupling of the final residue, a small portion of H-[Taeg]₅-Lys(ClZ)-MBHA was taken out for HF cleavage.

(b) Cleavage, Purification, and Identification of H-[Taeg]₅-Lys-NH₂.

The protected Boc-[Taeg]₅-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 14.0 mg of crude material upon HF cleavage of 52.4 mg dry H-Taeg]₅-Lys(ClZ)-MBHA resin. The crude product was not purified (about 99% purity).

(c) Cleavage, Purification, and Identification of H-[Taeg]₁₀-Lys-NH₂.

15 The protected Boc-[Taeg]₁₀-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 5.2 mg of crude material upon HF cleavage of 58.4 mg dry H-Taeg]₁₀-Lys(ClZ)-MBHA resin.

(d) Cleavage, Purification, and Identification of H-[Taeg]₁₅-Lys-NH₂.

The protected Boc-[Taeg]₁₅-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 114 mg of crude material upon HF cleavage of about 604 mg dry H-Taeg]₁₅-Lys(ClZ)-MBHA resin.

25 (e) Cleavage, Purification, and Identification of H-[Taeg]₂₀-Lys-NH₂.

The protected Boc-[Taeg]₂₀-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 19.3 mg of crude material upon HF cleavage of 81.0 mg dry H-Taeg]₂₀-Lys(ClZ)-MBHA resin.

(f) Cleavage, Purification, and Identification of H-[Taeg]₂₅-Lys-NH₂.

The protected Boc-[Taeg]₂₅-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 141 mg of crude material upon HF cleavage of about 417 mg dry H-Taeg]₂₅-Lys(ClZ)-MBHA resin.

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(g) Synthetic Pr tocol 8 (General Pr t col)

(1) Boc-deprotection with TFA/CH₂Cl₂, (1:1, v/v), 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 6 x 1 min; (3) neutralization with DIEA/CH₂Cl₂, (1: 19, v/v), 3 x 2 min; (4) 5 washing with CH₂Cl₂, 6 x 1 min, and drain for 1 min; (5) at some stages of the synthesis, 2-5 mg sample of PNA-resin is taken out and dried thoroughly for a ninhydrin analysis to determine the substitution; (6) addition of Boc-protected PNA monomer (Pfp ester); the coupling reaction was allowed to 10 proceed for a total of X hrs shaking; (7) washing with DMF, 1 x 2 min; (8) washing with CH₂Cl₂, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂, (1: 19, v/v), 2 x 2 min; (10) washing with CH₂Cl₂, 6 x 1 min; (11) occasionally, 2-5 mg 15 sample of protected PNA-resin is taken out and dried thoroughly for a ninhydrin analysis to determine the extent of coupling; (12) at some stages of the synthesis, unreacted amino groups are blocked by acetylation with a mixture of acetic anhydride/pyridine/CH₂Cl₂, (1:1:2, v/v/v) for 2 h followed by washing with CH₂Cl₂, 6 x 1 min, and, occasionally, 20 ninhydrin analysis.

EXAMPLE 88

Solid-Phase Synthesis of H-[Taeg]4-Caeg-[Taeg]5-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]4-C[Z]aeg-[Taeg]5-25 Lys(ClZ)-MBHA Resin.

About 1 g of wet Boc-[Taeg]5-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]4-C[Z]aeg-[Taeg]5-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC coupling of all residues utilizing 0.16 M of BocC[Z]aeg-OH 30 together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ or 0.16 M BocTaeg-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored by the ninhydrin reaction, which 35 showed about 98% incorporation of C[Z]aeg and close to quantitative incorporation of all the Taeg residues.

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(b) Cl avag , Purificati n, and Identification of H-[Taeg]4-C[Z]aeg-[Taeg]5-Lys-NH₂.

The protected Boc-[Taeg]4-C[Z]aeg-[Taeg]5-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 5 22.5 mg of crude material upon HF cleavage of 128.2 mg dry H-[Taeg]4-C[Z]aeg-[Taeg]5-Lys(ClZ)-MBHA resin. Crude product (5.8 mg) was purified to give 3.1 mg of H-[Taeg]4-Caeg-[Taeg]5-Lys-NH₂.

(c) Synthetic Protocol 9 (General Protocol)

10 (1) Boc-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 3 x 1 min and 1 x 30 min; (2) washing with CH₂Cl₂, 6 x 1 min; (3) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 3 x 2 min; (4) washing with CH₂Cl₂, 6 x 1 min, and drain for 1 min; (5) at some stages of the synthesis, 2-5 mg sample of PNA-resin is 15 taken out and dried thoroughly for a ninhydrin analysis to determine the substitution; (6) addition of Boc-protected PNA monomer (free acid) in X ml DMF followed by addition of DCC in X ml CH₂Cl₂; the coupling reaction was allowed to proceed for a total of Y hrs shaking; (7) washing with DMF, 1 x 2 min; 20 (8) washing with CH₂Cl₂, 4 x 1 min; (9) neutralization with DIEA/CH₂Cl₂ (1: 19, v/v), 2 x 2 min; (10) washing with CH₂Cl₂, 6 x 1 min; (11) occasionally, 2-5 mg sample of protected PNA-resin is taken out and dried thoroughly for a ninhydrin analysis to determine the extent of coupling; (12) at some 25 stages of the synthesis, unreacted amino groups are blocked by acetylation with a mixture of acetic anhydride/pyridine/CH₂Cl₂ (1:1:2, v/v/v) for 2 h followed by washing with CH₂Cl₂, 6 x 1 min, and, occasionally, ninhydrin analysis.

30

EXAMPLE 89

Solid-Phase Synthesis of H-[Taeg]4-(NBaeg)-[Taeg]5-Lys-NH₂.
(NB = COCH₃)

(a) Stepwis Assembly of Boc-[Taeg]4-(NBaeg)-[Taeg]5-35 Lys(ClZ)-MBHA Resin.

About 1 g of wet Boc-[Taeg]5-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]4-(NBaeg)-

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[Taeg]5-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC coupling utilizing 0.16 M of Boc(NBaeg)-OH together with 0.16 M DCC in 2.0 ml neat CH₂Cl₂ or 0.16 M BocTaeg-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9").
5 Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The NBaeg residue was coupled three times and the Taeg residues were all coupled once. The synthesis was monitored by the ninhydrin reaction which showed >99% total incorporation of NBaeg (about 88% after the first
10 coupling and about 93% after the second coupling) and close to quantitative incorporation of all the Taeg residues.

(b) Cleavage, Purification, and Identification of H-[Taeg]4-(NBaeg)-[Taeg]5-Lys-NH₂.

The protected Boc-[Taeg]4-(NBaeg)-[Taeg]5-Lys(ClZ)-MBHA
15 resin was treated as described in Example 17c to yield about 33.6 mg of crude material upon HF cleavage of 108.9 mg dry H-[Taeg]4-(NBaeg)-[Taeg]5-Lys(ClZ)-MBHA resin. Crude product (20.6 mg) was purified to give 4.6 mg of H-[Taeg]4-(NBaeg)-[Taeg]5-Lys-NH₂. For (M+H)⁺, the calculated m/z value was
20 2683.12 and the measured m/z value was 2683.09.

EXAMPLE 90

Solid-Phase Synthesis of H-[Taeg]4-aeg-[Taeg]5-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]4-aeg-[Taeg]5-Lys(ClZ)-MBHA Resin.

About 1 g of wet Boc-[Taeg]5-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]4-aeg-[Taeg]5-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC single coupling of all residues utilizing: (1) 0.16 M of BocTaeg-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ or (2) 0.16 M BocTaeg-OH together with (2) 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored by the ninhydrin
35 reaction, which showed close to quantitative incorporation of all the residues.

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(b) Cleavage, Purification, and Identification of H-[Taeg]4-aeg-[Taeg]5-Lys-NH₂.

The protected Boc-[Taeg]4-aeg-[Taeg]5-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 5 22.2 mg of crude material upon HF cleavage of 126.0 mg dry H-[Taeg]4-aeg-[Taeg]5-Lys(ClZ)-MBHA resin. Crude product (22.2 mg) was purified to give 7.6 mg of H-[Taeg]4-aeg-[Taeg]5-Lys-NH₂. For (M+H)⁺, the calculated m/z value was 2641.11 and the measured m/z value was 2641.16.

10

EXAMPLE 91

Solid-Phase Synthesis of H-[Taeg]4-Gly-[Taeg]5-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]4-Gly-[Taeg]5-Lys(ClZ)-MBHA Resin.

15 About 1 g of wet Boc-[Taeg]5-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]4-Gly-[Taeg]5-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC single coupling of all residues utilizing: (1) 0.16 M of BocGly-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ or 20 (2) 0.16 M BocTaeg-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of all the 25 residues.

(b) Cleavage, Purification, and Identification of H-[Taeg]4-Gly-[Taeg]5-Lys-NH₂.

The protected Boc-[Taeg]4-Gly-[Taeg]5-Lys(ClZ)-MBHA resin was treated as described in Example 18c to yield about 30 45.0 mg of crude material upon HF cleavage of 124.1 mg dry H-[Taeg]4-Gly-[Taeg]5-Lys(ClZ)-MBHA resin. Crude product (40.4 mg) was purified to give 8.2 mg of H-[Taeg]4-Gly-[Taeg]5-Lys-NH₂.

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EXAMPLE 92

Solid-Phase Synthesis of H-[Taeg]4-Gly2-[Taeg]5-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]4-Gly2-[Taeg]5-Lys(ClZ)-MBHA Resin.

5 About 1 g of wet Boc-[Taeg]5-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]4-[C[Z]aeg]2-Taeg-C[Z]aeg-Taeg-C[Z]aeg-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC single coupling of all residues utilizing: (1) 0.16 M of BocGly-OH together with 0.16 M DCC 10 in 2.0 ml 50% DMF/CH₂Cl₂ or (2) 0.16 M BocTaeg-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative 15 incorporation of all the residues.

(b) Cleavage, Purification, and Identification of H-[Taeg]4-Gly2-[Taeg]5-Lys-NH₂.

The protected Boc-[Taeg]4-Gly2-[Taeg]5-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 20 32.6 mg of crude material upon HF cleavage of 156.6 mg dry H-[Taeg]4-Gly2-[Taeg]5-Lys(ClZ)-MBHA resin. Crude product (30 mg) was purified to give 7.8 mg of H-[Taeg]4-Gly2-[Taeg]5-Lys-NH₂. For (M+H)⁺, the calculated m/z value was 2655.09 and the measured m/z value was 2655.37.

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EXAMPLE 93

Solid-Phase Synthesis of H-[Taeg]4-[Caeg]2-Ta g-Caeg-Taeg-Caeg-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]4-[C[Z]aeg]2-Taeg-5 C[Z]aeg-Taeg-C[Z]aeg-Lys(ClZ)-MBHA Resin.

About 1.5 g of wet Boc-Lys(ClZ)-MBHA (0.28 mmol.Lys/g) resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]4-[C[Z]aeg]2-Taeg-C[Z]aeg-Taeg-C[Z]aeg-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC single coupling of all residues 10 utilizing: (1) 0.16 M of BocC[Z]-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ or (2) 0.16 M BocTaeg-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored 15 by the ninhydrin reaction, which showed close to quantitative incorporation of all the residues.

(b) Cleavage, Purification, and Identification of H-[Taeg],-[Caeg],-Taeg-Caeg-Taeg-Caeg-Lys-NH₂.

The protected Boc-[Taeg]4-[C[Z]aeg]2-Taeg-C[Z]aeg-Taeg-20 C[Z]aeg-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 52.1 mg of crude material upon HF cleavage of 216.7 mg dry H-[Taeg]4-[C[Z]aeg]2-Taeg-C[Z]aeg-Taeg-C[Z]aeg-Lys(ClZ)-MBHA resin. Crude product (30.6 mg) was purified to give 6.2 mg of H-[Taeg]4-[Caeg]2-Taeg-Caeg-25 Taeg-Caeg-Lys-NH₂. For (M+H)⁺ the calculated m/z value was 2747.15 and the measured m/z value was 2746.78.

EXAMPLE 94

Solid-Phase Synthesis of H-Caeg-Taeg-Caeg-Taeg-[Caeg]3-Taeg-30 Caeg-Taeg-Lys-NH₂.

(a) Stepwise Assembly of Boc-C[Z]aeg-Taeg-C[Z]aeg-Taeg-[C[Z]aeg]3-Taeg-C[Z]aeg-Taeg-Lys(ClZ)-MBHA Resin.

About 1.5 g of wet Boc-Lys(ClZ)-MBHA (0.28 mmol Lys/g) resin was placed in a 5 ml SPPS reaction vessel. Boc-C[Z]aeg-Taeg-C[Z]aeg-Taeg-[C[Z]aeg]3-Taeg-C[Z]aeg-Taeg-Lys(ClZ)-MBHA 35 resin was assembled by *in situ* DCC single coupling of all residues utilizing: (1) 0.16 M of BocC[Z]-OH together with

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0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ or (2) 0.16 M BocTaeg-OH together with 0.16 M DCC in 2.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of all the residues.

(b) Cleavage, Purification, and Identification of H-Caeg-Taeg-Caeg-Taeg-[Caeg]3-Taeg-Caeg-Taeg-Lys-NH₂.

The protected Boc-C(Z)aeg-Taeg-C(Z)aeg-Taeg-[C(Z)aeg]3-Taeg-C(Z)aeg-TaegLys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 56.1 mg of crude material upon HF cleavage of 255.0 mg dry H-C(Z)aeg-Taeg-C(Z)aeg-Taeg-[C(Z)aeg]3-Taeg-C(Z)aeg-TaegLys(ClZ)-MBHA resin. Crude product (85.8 mg) was purified to give 46.2 mg of H-Caeg-Taeg-Caeg-Taeg-[Caeg]3-Taeg-Caeg-Taeg-LysNH₂. For (M+H)⁺ the calculated m/z value was 2717.15 and the measured m/z value was 2716.93.

EXAMPLE 95

20 Solid-Phase Synthesis of H-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-Lys-NH₂, H-Caeg-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-Lys-NH₂, and H-Tyr-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA Resin, Boc-Caeg-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA Resin, and Boc-Tyr(BrZ)-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA Resin.

About 3 g of wet Boc-Lys(ClZ)-MBHA (0.28 mmol Lys/g) resin was placed in a 20 ml SPPS reaction vessel. Boc-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA resin was assembled by *in situ* DCC single coupling of all residues utilizing: (1) 0.16 M of BocC(Z)-OH together with 0.16 M DCC in 3.0 ml 50% DMF/CH₂Cl₂ or (2) 0.16 M BocTaeg-OH together with 0.16 M DCC in 3.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative

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incorporation of all the residues. After deprotection of the N-terminal Boc group, half of the PNA-resin was coupled quantitatively onto Tyr(BrZ)-OH and a small portion was coupled quantitatively onto one more Caeg residue. Both 5 couplings employed the above-mentioned synthetic protocol.

(b) Cleavage, Purification, and Identification of H-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-Lys-NH₂.

The protected Boc-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA resin was treated as described in 10 Example 17c to yield about 50.9 mg of crude material upon HF cleavage of 182.5 mg dry H-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA resin. Crude product (50.9) mg was purified to give 13.7 mg of H-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-LysNH₂. For (M+H)⁺ the calculated m/z value was 2466.04; the 15 m/z value was not measured.

(c) Cleavage, Purification, and Identification of H-Tyr-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-Lys-NH₂.

The protected Boc-Tyr(BrZ)-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA resin was treated as described in 20 Example 17c to yield about 60.8 mg of crude material upon HF cleavage of 188.8 mg dry H-Tyr(BrZ)-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA resin. Crude product (60.8 mg) was purified to give 20.7 mg of H-Tyr-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-LysNH₂. For (M+H)⁺ the calculated m/z value 25 was 2629.11 and the measured m/z value was 2629.11.

(d) Cleavage, Purification, and Identification of H-Caeg-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-Lys-NH₂.

The protected Boc-C(Z)aeg-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA resin was treated as described in 30 Example 17c to yield about 11.7 mg of crude material upon HF cleavage of 42.0 mg dry H-C(Z)aeg-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(ClZ)-MBHA resin. Crude product (11.6 mg) was purified to give 3.1 mg of H-Caeg-[Taeg]2-[Caeg]3-[Taeg]2-[Caeg]2-LysNH₂. For (M+H)⁺ the calculated m/z value was 35 2717.15; the m/z value was not measured.

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EXAMPLE 96

Solid-Phase Synthesis of H-[Caeg]2-[Taeg]2-[Caeg]3-[Taeg]2-Lys-NH₂,
H-Taeg-[Caeg]2-[Taeg]2-[Caeg]3-[Taeg]2-Lys-NH₂, and H-Tyr-
5 **[Caeg]2-[Taeg]2-[Caeg]3-[Taeg]2-Lys-NH₂.**

(a) Stepwise Assembly of Boc-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(Clz)-MBHA Resin, Boc-Taeg-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(Clz)-MBHA Resin, and Boc-Tyr(Brz)-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(Clz)-MBHA Resin.

About 3 g of wet Boc-Lys(Clz)-MBHA (0.28 mmol Lys/g) resin was placed in a 20 ml SPPS reaction vessel. Boc-[Taeg]2-[C(Z)aeg]3-[Taeg]2-[C(Z)aeg]2-Lys(Clz)-MBHA resin was assembled by *in situ* DCC single coupling of all residues 15 utilizing: (1) 0.16 M of BocC[Z]-OH together with 0.16 M DCC in 3.0 ml 50% DMF/CH₂Cl₂ or (2) 0.16 M BocTaeg-OH together with 0.16 M DCC in 3.0 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed for a total of 20-24 hrs with shaking. The synthesis was monitored 20 by the ninhydrin reaction, which showed close to quantitative incorporation of all the residues. After deprotection of the N-terminal Boc group, half of the PNA-resin was coupled quantitatively onto Tyr(Brz)-OH and a small portion was coupled quantitatively onto one more Taeg residue. Both 25 couplings employed the above-mentioned synthetic protocol.

(b) Cleavage, Purification, and Identification of H-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys-NH₂.

The protected Boc-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(Clz)-MBHA resin was treated as described in 30 Example 17c to yield about 57.6 mg of crude material upon HF cleavage of 172.7 mg dry H-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(Clz)-MBHA resin. Crude product (57.6 mg) was purified to give 26.3 mg of H-[Caeg]2-[Taeg]2-[Caeg]3-[Taeg]2-Lys-NH₂. For (M+H)⁺ the calculated m/z value was 2466.04; the 35 m/z value was not measured.

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(c) Cleavage, Purification, and Identification of H-Tyr-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys-NH₂.

The protected Boc-Tyr(BrZ)-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 57.6 mg of crude material upon HF cleavage of 172.7 mg dry H-Tyr(BrZ)-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(ClZ)-MBHA resin. Crude product (47.1 mg) was purified to give 13.4 mg of H-Tyr-[Caeg]2-[Taeg]2-[Caeg]3-[Taeg]2-Lys-NH₂. For (M+H)⁺ the calculated m/z value was 2629.11 and the measured m/z value was 2629.11.

(d) Cleavage, Purification, and Identification of H-Taeg-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys-NH₂.

The protected Boc-Taeg-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 53.4 mg of crude material upon HF cleavage of 42.4 mg dry H-Taeg-[C(Z)aeg]2-[Taeg]2-[C(Z)aeg]3-[Taeg]2-Lys(ClZ)-MBHA resin. Crude product (11.9 mg) was purified to give 4.3 mg of H-Taeg-[Caeg]2-[Taeg]2-[Caeg]3-[Taeg]2-Lys-NH₂. For (M+H)⁺ the calculated m/z value was 2732.15; the m/z value was not measured.

(c) Synthetic Protocol 10 (General Protocol)

Same protocol as "Synthetic Protocol 9", except that DCC has been replaced with DIC.

25

EXAMPLE 97

SYNTHESIS OF THE BACKBONE MOIETY FOR SCALE UP BY REDUCTIVE AMINATION

(a) Preparation of (bocamino)acetaldehyde.

30 3-Amino-1,2-propanediol(80.0 g; 0.88 mol) was dissolved in water (1500 ml) and the solution was cooled to 4°C, whereafter Boc anhydride (230 g; 1.05 mol) was added at once. The solution was gently heated to room temperature with a water bath. The pH was kept at 10.5 by the dropwise addition 35 of sodium hydroxide. Over the course of the reaction a total of 70.2 g NaOH, dissolved in 480 ml water, was added. After stirring overnight, ethyl acetate (1000 ml) was added and the

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mixture was cooled to 0°C and the pH was adjusted to 2.5 by the addition of 4 M hydrochloric acid. The ethyl acetate layer was removed and the acidic aqueous solution was extracted with more ethyl acetate (8x500 ml). The combined 5 ethyl acetate solution was reduced to a volume of 1500 ml using a rotary evaporator. The resulting solution was washed with half saturated potassium hydrogen sulphate (1500 ml) and then with saturated sodium chloride. It then was dried over magnesium sulphate and evaporated to dryness, in vacuo.

10 Yield. 145.3 g (86%)

3-Bocamino-1,2-propanediol (144.7 g; 0.757 mol) was suspended in water (750 ml) and potassium periodate (191.5 g; 0.833 mol) was added. The mixture was stirred under nitrogen for 2.5 h and the precipitated potassium iodate was removed 15 by filtration and washed once with water (100 ml). The aqueous phase was extracted with chloroform (6x400 ml). The chloroform extracts were dried and evaporated to dryness, in vacuo. Yield 102 g (93%) of an oil. The (bocamino)acetaldehyde was purified by kugelrohr distillation 20 at 84°C and 0.3 mmHg in two portions. The yield 79 g (77%) of a colorless oil.

(b) Preparation of (N'-bocaminoethyl)glycine methyl ester

Palladium on carbon (10%; 2.00 g) was added to a 25 solution of (bocamino)acetaldehyde (10.0 g; 68.9 mmol) in methanol (150 ml) at 0°C. Sodium acetate (11.3 g; 138 mmol) in methanol (150 ml), and glycine methyl ester hydrochloride (8.65 g; 68.9 mmol) in methanol (75 ml) then were added. The mixture was hydrogenated at atmospheric pressure for 2.5 h, 30 then filtered through celite and evaporated to dryness, in vacuo. The material was redissolved in water (150 ml) and the pH was adjusted to 8.0 with 0.5 N NaOH. The aqueous solution was extracted with methylene chloride (5 x 150 ml). The combined extracts were dried over sodium sulphate and 35 evaporated to dryness, in vacuo. This resulted in 14.1 g (88%) of (N'-bocaminoethyl)glycine methyl ester. The crude material was purified by kugelrohr destination at 120°C and

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0.5 mmHg to give 11.3 g (70%) of a colorless oil. The product had a purity that was higher than the material produced in example 26 according to tlc-analysis (10% methanol in methylene chloride).

5 Alternatively, sodium cyanoborohydride can be used as reducing agent instead of hydrogen (with Pd(C) as catalyst), although the yield (42%) was lower.

(c) Preparation of (N'-bocaminoethyl)glycine ethyl ester.

10 The title compound was prepared by the above procedure with glycine ethyl ester hydrochloride substituted for glycine methyl ester hydrochloride. Also, the solvent used was ethanol. The yield was 78%.

15 EXAMPLE 98

Solid-Phase Synthesis of H-Tyr-[Taeg]₁₀-Lys-NH₂

(a) Stepwise Assembly of Boc-Tyr(BrZ)-[Taeg]₁₀-Lys(ClZ)-MBHA Resin.

About 0.2 g of wet Boc-[Taeg]₁₀-Lys(ClZ)-MBHA resin was 20 placed in a 5 ml SPPS reaction vessel. Boc-Tyr(BrZ)-[Taeg]₁₀-Lys(ClZ)-MBHA resin was assembled by standard *in situ* DCC coupling utilizing 0.32 M of BocTyr(BrZ)-OH together with 0.32 M DCC in 3.0 ml neat CH₂Cl₂ overnight. The ninhydrin reaction showed about 97% incorporation of BocTyr(BrZ).

25 (b) Cleavage, Purification, and Identification of H-Tyr-[Taeg]₁₀-Lys-NH₂.

The protected Boc-Tyr(BrZ)-[Taeg]₁₀-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 5.5 mg of crude material upon HF cleavage of 20.7 mg dry H-Tyr(BrZ)-30 [Taeg]₁₀-Lys(ClZ)-MBHA resin. The crude product was purified to give 2.5 mg of H-Tyr-[Taeg]₁₀-Lys-NH₂.

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EXAMPLE 99

Solid-Phase Synthesis of Dansyl-[Taeg]₁₀-Lys-NH₂

(a) Stepwise Assembly of Dansyl-[Taeg]₁₀-Lys(ClZ)-MBHA Resin.

5 About 0.3 g of wet Boc-[Taeg]₁₀-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Dansyl-[Taeg]₁₀-Lys(ClZ)-MBHA resin was assembled by coupling of 0.5 M dansyl-Cl in 2.0 ml neat pyridine overnight. The ninhydrin reaction showed about 95% incorporation of dansyl.

10 **(b) Cleavage, Purification, and Identification of Dansyl-[Taeg]₁₀-Lys-NH₂.**

The protected dansyl-[Taeg]₁₀-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 12 mg of crude material upon HF cleavage of 71.3 mg dry dansyl-[Taeg]₁₀-Lys(ClZ)-MBHA resin. The crude product was purified to give 15 5.4 mg of dansyl-[Taeg]₁₀-Lys-NH₂.

EXAMPLE 100

Solid-Phase Synthesis of Gly-Gly-His-[Taeg]₁₀-Lys-NH₂

20 **(a) Stepwise Assembly of Boc-Gly-Gly-His(Tos)-[Taeg]₁₀-Lys(ClZ)-MBHA Resin.**

About 0.05 g of Boc-[Taeg]₁₀-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Boc-Gly-Gly-His(Tos)-[Taeg]₁₀-Lys(ClZ)-MBHA resin was assembled by standard double 25 *in situ* DCC coupling of Boc-protected amino acid (0.1 M) in 2.5 ml 25% DMF/CH₂Cl₂, except for the first coupling of BocHis(Tos), which was done by using a preformed symmetrical anhydride (0.1M) in 25% DMF/CH₂Cl₂. All couplings were performed overnight and ninhydrin reactions were not carried 30 out.

(b) Cleavage, Purification, and Identification of Gly-Gly-His-[Taeg]₁₀-Lys-NH₂.

The protected Boc-Gly-Gly-His(Tos)-[Taeg]₁₀-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield 35 about 10.3 mg of crude material (about 40% purity) upon HF cleavage of 34.5 mg dry Boc-Gly-Gly-His(Tos)-[Taeg]₁₀-Lys(ClZ)-

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MBHA resin. A small portion of the crude product (taken out before lyophilization) was purified to give 0.1 mg of Gly-Gly-His-[Taeg]₁₀-Lys-NH₂.

5 EXAMPLE 101

Solid-Phase Synthesis of H-[Taeg],-[Caeg],-NH₂.

(a) Stepwise Assembly of Boc-[Taeg],-[C(Z)aeg],-MBHA Resin.

About 0.2 g of MBHA resin was placed in a 3 ml SPPS reaction vessel and neutralized. The loading was determined to be about 0.64 mmol/g. BocC(Z)aeg-OPfp was coupled onto the resin using a concentration of 0.13 M in 2.5 ml 25% phenol//CH₂Cl₂. The ninhydrin analysis showed a coupling yield of about 40%. The remaining free amino groups were acetylated as usual. Boc-[Taeg],-[C(Z)aeg],-MBHA resin was assembled by single *in situ* DCC coupling of the next residue utilizing 0.11 M of BocC(Z)aeg-OH together with 0.11 M DCC in 2.5 ml 50% DMF/CH₂Cl₂, and by coupling with 0.13 M BocTaeg-OPfp in neat CH₂Cl₂, for the remaining residues ("Synthetic Protocol 8"). Each coupling reaction was allowed to proceed with shaking overnight. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of all the residues.

(b) Cleavage, Purification, and Identification of H-[Taeg],-[Caeg],-NH₂.

The protected Boc-[Taeg],-[C(Z)aeg],-MBHA resin was treated as described in Example 17c to yield about 21.7 mg of crude material (>80% purity) upon HF cleavage of 94.8 mg dry H-[Taeg],-[C(Z)aeg],-MBHA resin. Crude product (7.4 mg) was purified to give 2.0 mg of H-[Taeg],-[Caeg],-NH₂ (>99% purity).

EXAMPLE 102

Solid-Phase Synthesis of H-[Taeg],-Caeg-[Taeg],-NH₂.

(a) Stepwise Assembly of Boc-[Taeg],-C(Z)aeg-[Taeg],-MBHA Resin.

About 0.2 g of the above-mentioned MBHA resin was placed in a 5 ml SPPS reaction vessel and neutralized. Boc-

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[Taeg],-C(Z)aeg-[Taeg],-MBHA resin was assembled by single *in situ* DCC coupling of the C(Z)aeg residue utilizing 0.13 M of BocC[Z]aeg-OH together with 0.13 M DCC in 2.5 ml 50% DMF/CH₂Cl₂, and by coupling the Taeg residues with 0.13 M BocTaeg-OPfp in 5 2.5 ml neat CH₂Cl₂. Each coupling reaction was allowed to proceed with shaking overnight. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of all the residues.

(b) Cleavage, Purification, and Identification of H-
10 [Taeg],-Caeg-[Taeg],-NH₂.

The protected Boc-[Taeg],-C(Z)aeg-[Taeg],-MBHA resin was treated as described in Example 17c to yield about 44.4 mg of crude material upon HF cleavage of about 123 mg dry H-[Taeg],-C(Z)aeg-[Taeg],-MBHA resin. Crude product (11.0 mg) was 15 purified to give 3.6 mg of H-[Taeg],-Caeg-[Taeg],-NH₂.

EXAMPLE 103

Solid-Phase Synthesis of H-Taeg-Caeg-[Taeg],-LysNH₂.

(a) Stepwise Assembly of Boc-Taeg-C(Z)aeg-[Taeg],-
20 Lys(ClZ)-MBHA Resin.

About 0.3 g of wet Boc-[Taeg],-Lys(ClZ)-MBHA resin was placed in a 3 ml SPPS reaction vessel. Boc-Taeg-C(Z)aeg-[Taeg],-Lys(ClZ)-MBHA resin was assembled by single *in situ* DCC coupling overnight of the C(Z)aeg residue ("Synthetic 25 Protocol" 9) utilizing 0.2 M of BocC[Z]aeg-OH together with 0.2 M DCC in 2.5 ml 50% DMF/CH₂Cl₂, (incorporation was about 80% as judged by ninhydrin analysis; remaining free amino groups were acetylated) and by overnight coupling the Taeg residue with 0.15 M BocTaeg-OPfp in 2.5 ml neat CH₂Cl₂, (nearly 30 quantitatively).

(b) Cleavage, Purification, and Identification of H-Taeg-Caeg-[Taeg],-LysNH₂.

The protected Boc-Taeg-C(Z)aeg-[Taeg],-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 35 22.3 mg of crude material upon HF cleavage of about 76.5 mg dry H-Taeg-C(Z)aeg-[Taeg],-Lys(ClZ)-MBHA resin. Crude product

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(6.7 mg) was purified to give 2.6 mg of H-Taeg-Caeg-[Taeg]_s-LysNH₂. For (M+H)⁺ the calculated m/z value was 2792.15 and the measured m/z value was 2792.21.

5 EXAMPLE 104

Solid-Phase Synthesis of H-Caeg-[Taeg]_s-Lys-NH₂ and H-[Taeg]_s-Caeg-[Taeg]_s-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]_s-C(Z)aeg-[Taeg]_s-Lys(ClZ)-MBHA Resin.

10 About 0.5 g of wet Boc-[Taeg]_s-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. Boc-[Taeg]_s-C(Z)aeg-[Taeg]_s-Lys(ClZ)-MBHA resin was assembled by single *in situ* DCC coupling of all residues utilizing: (1) 0.12 M of BocC[Z]aeg-OH together with 0.12 M DCC in 3.0 ml 50% DMF/CH₂Cl₂, 15 or (2) 0.12 M BocTaeg-OH together with 0.12 M DCC in 3.0 ml 50% DMF/CH₂Cl₂, ("Synthetic Protocol 9"). Each coupling reaction was allowed to proceed overnight with shaking. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of all the 20 residues. During the synthesis, a small portion of H-C(Z)aeg-[Taeg]_s-Lys(ClZ)-MBHA resin was taken out for HF cleavage.

(b) Cleavage, Purification, and Identification of H-Caeg-[Taeg]_s-Lys-NH₂.

The protected Boc-C[Z]aeg-[Taeg]_s-Lys(ClZ)-MBHA resin 25 was treated as described in Example 17c to yield about 3.0 mg of crude material upon HF cleavage of 37.5 mg dry H-C[Z]aeg-[Taeg]_s-Lys(ClZ)-MBHA resin. About 0.7 mg of the crude product was purified to give about 0.5 mg of H-Caeg-[Taeg]_s-Lys-NH₂.

30

(c) Cleavage, Purification, and Identification of H-[Taeg]_s-Caeg-[Taeg]_s-Lys-NH₂.

The protected Boc-[Taeg]_s-C[Z]aeg-[Taeg]_s-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 35 37.7 mg of crude material upon HF cleavage of 118.6 mg dry H-[Taeg]_s-C[Z]aeg-[Taeg]_s-Lys(ClZ)-MBHA resin.

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EXAMPLE 105

Solid-Phase Synthesis of H-[Caeg]₅-Lys-NH₂, H-[Caeg]₆-Lys-NH₂, H-[Caeg]₇-Lys-NH₂, and H-[Caeg]₈-Lys-NH₂

(a) Stepwise Assembly of Boc-[C(Z)aeg]₁₀-Lys(ClZ)-MBHA

5 Resin and Shorter Fragments.

About 5 g of wet Boc-Lys(ClZ)-MBHA resin (substitution = 0.3 mmol Lys/g) was placed in a 30 ml SPPS reaction vessel. Boc-[C(Z)aeg]₁₀-Lys(ClZ)-MBHA resin was assembled by single *in situ* DCC coupling of the first three residues with 0.1 M of 10 BocC(Z)aeg-OH together with 0.1 M DCC in 10 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 9") and by single *in situ* DIC coupling of the remaining seven residues with 0.1 M of BocC(Z)aeg-OH together with 0.1 M DIC in 10 ml 50% DMF/CH₂Cl₂ ("Synthetic Protocol 10"). All the coupling reactions were allowed to 15 proceed overnight. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of all residues. During the synthesis, portions of the shorter fragments H-[C(Z)aeg]₅-Lys(ClZ)-MBHA resin, H-[C(Z)aeg]₆-Lys(ClZ)-MBHA resin, H-[C(Z)aeg]₇-Lys(ClZ)-MBHA 20 resin, H-[C(Z)aeg]₈-Lys(ClZ)-MBHA resin, and H-[C(Z)aeg]₉-Lys(ClZ)-MBHA resin were taken out for HF cleavage.

(b) Cleavage, Purification, and Identification of H-[Caeg]₅-Lys-NH₂.

The protected Boc-[C(Z)aeg]₅-Lys(ClZ)-MBHA resin was treated 25 as described in Example 17c to yield about 10.8 mg of crude material upon HF cleavage of 60.1 mg dry H-[C(Z)aeg]₅-Lys(ClZ)-MBHA resin.

(c) Cleavage, Purification, and Identification of H-[Caeg]₆-Lys-NH₂.

30 The protected Boc-[C(Z)aeg]₆-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 13.4 mg of crude material upon HF cleavage of 56.2 mg dry H-[C(Z)aeg]₆-Lys(ClZ)-MBHA resin.

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(d) Cleavage, Purification, and Identification of H-[Caeg]₈-Lys-NH₂.

The protected Boc-[C(Z)aeg]₈-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 16.8 mg of crude material upon HF cleavage of 65.6 mg dry H-[C(Z)aeg]₈-Lys(ClZ)-MBHA resin.

(e) Cleavage, Purification, and Identification of H-[Caeg]₁₀-Lys-NH₂.

The protected Boc-[C(Z)aeg]₁₀-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 142.4 mg of crude material upon HF cleavage of 441 mg dry H-[C(Z)aeg]₁₀-Lys(ClZ)-MBHA resin.

EXAMPLE 106

15 Solid-Phase Synthesis of H-[Taeg]₂-Caeg-[Taeg]₂-Caeg-[Taeg]₂-Lys-NH₂.

(a) Stepwise Assembly of Boc-[Taeg]₂-C(Z)aeg-[Taeg]₂-C(Z)aeg-[Taeg]₂-Lys(ClZ)-MBHA Resin.

About 0.3 g of wet H-[Taeg]₂-C(Z)aeg-[Taeg]₂-Lys(ClZ)-MBHA resin from the earlier synthesis of Boc-[Taeg]₂-C(Z)aeg-[Taeg]₂-Lys(ClZ)-MBHA resin was placed in a 5 ml SPPS reaction vessel. After coupling of the next residue five times, a total incorporation of BocC(Z)aeg of 87% was obtained. The five repeated couplings were carried out with 0.18 M BocC(Z)aeg-OPfp in 2 ml of TFE/CH₂Cl₂, (1:2, v/v), 2 ml of TFE/CH₂Cl₂, (1:2, v/v), 2 ml of TFE/CH₂Cl₂, (1:2, v/v) with two drops of dioxane and two drops of DIEA (this condition gave only a few per cent coupling yield), 2 ml of TFE/CH₂Cl₂, (1:2, v/v) plus 0.5 g phenol, and 1 ml of CH₂Cl₂ plus 0.4 g of phenol, respectively. The two final Taeg residues were incorporated close to quantitatively by double couplings with 0.25 M BocTaeg-OPfp in 25% phenol/CH₂Cl₂. All couplings were allowed to proceed overnight.

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(b) Cleavage, Purification, and Identification of H-[Taeg]₂-Caeg-[Taeg]₂-Caeg-[Taeg]₂-Lys-NH₂

The protected Boc-[Taeg]₂-C(Z)aeg-[Taeg]₂-C(Z)aeg-[Taeg]₂-Lys(ClZ)-MBHA resin was treated as described in Example 17c to yield about 7 mg of crude material upon HF cleavage of 80.7 mg dry H-[Taeg]₂-C(Z)aeg-[Taeg]₂-C(Z)aeg-[Taeg]₂-Lys(ClZ)-MBHA resin. The crude product was purified to give 1.2 mg of H-[Taeg]₂-Caeg-[Taeg]₂-Caeg-[Taeg]₂-Lys-NH₂ (>99.9% purity).

10 EXAMPLE 107

SYNTHESIS OF A PNA WITH TWO ANTI PARALLEL STRANDS TIED TOGETHER

synthesis of H-[Taeg]-[Taeg]-[Taeg]-[Gaeg]-[Taeg]-[Taeg]-[Taeg]-[6-AHA]-[aeg]-[6-AHA]-[Taeg]-[Taeg]-[Taeg]-[Aaeg]-[Taeg]-[Taeg]-[Taeg]-LYS-NH₂. (6-AHA = 6-aminohexanoic acid) (Figure 26)

The protected PNA was assembled onto a Boc-Lys(ClZ) modified MBHA resin with a substitution of approximately 0.30 mmol/g. Capping of uncoupled amino groups was only carried out before the incorporation of the BocGaeg-OH monomer. Synthesis was initiated on 1.00 g (dry weight) of preswollen (overnight in DCM) and neutralized Boc-Lys(ClZ)-MBHA resin. The incorporation of the monomers followed the protocol of Example 32 and Example 71. The coupling reaction was monitored by qualitative ninhydrin reaction (kaiser test). In case of a positive Kaiser test, the coupling reaction was repeated until the test showed no coloration of the beads. Final deprotection, cleavage from support, and purification were performed according to standard procedures.

30

EXAMPLE 108

Alternative protecting group strategy for PNA-synthesis (Figure 27).

(a) Synthesis of target compounds.

35 2-amino-6-O-benzyl purine. To a solution of 2.5 g (0.109 mol) of sodium in 100 ml of benzyl alcohol was added 10.75 g (0.063 mol) of 2-amino-6-chloropurine. The mixture

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was stirred for 12 h at 120 0°C. The solution was cooled to room temperature and neutralized with acetic acid and extracted with 10 portions of 50 ml of 0.2 N sodium hydroxide. The collected sodium hydroxide phases were washed with 100 ml 5 of diethyl ether and neutralized with acetic acid, whereby precipitation starts. The solution was cooled to 0°C and the yellow precipitate was collected by filtration. Recrystallization from ethanol gave 14.2 g 92% of pure white crystals of the target compound. 1H-NMR (250 MHz--DMSO-d6) 10 d ppm: 8-H, 7.92; benzyl aromatic, 7.60-7.40; 2NH₂, 6.36; benzyl CH₂, 5.57.

(2-amino-6-O-benzyl purinyl)methylethanoate. A mixture of 5 g (0.0207 mol) of 2-amino-6-O-benzyl-purine, 30 ml of DMF and 2.9 g (0.021 mol) of potassium carbonate was stirred at 15 room temperature. Methyl bromoacetate (3.2 g; 1.9 ml; 0.0209 mol) was added dropwise. The solution was filtrated after 4 h and the solvent was removed under reduced pressure (4 mmHg, 40°C). The residue was recrystallized two times from ethyl acetate to give 3.7 g (57%) of the target compound. 1H-NMR 20 (250 MHz, DMSO-d6) d ppm: 8-H, 7.93; benzyl aromatic 7.4-7.6; 2-NH₂, 6.61; benzyl CH₂, 5.03; CH₂, 5.59; OCH₃, 3.78.

(2N-p-Toluene sulfonamido-6-O-benzyl purinyl) methyl ethanoate. To a solution of 0.5 g (1.6 mmol) of (2-amino-6-O-benzyl purinyl) methyl ethanoate in 25 ml methylene chloride 25 was added 0.53 g (1.62 mmol) of p-toluenesulfonic anhydride and 0.22 g (1.62 mmol) of potassium carbonate. The mixture was stirred at room temperature. The mixture was filtered and the solvent was removed at reduced pressure (15 mmHg, 40°C). Diethyl ether was added to the oily residue. The resulting 30 solution was stirred overnight, whereby the target compound (0.415 mg; 55%) precipitated and was collected by filtration. 1H-NMR (250 MHz, DMSO-d6) d ppm: 8-H, 8.97; aromatic 7.2-7.8; benzyl CH₂, 5.01; CH₂, 4.24; OCH₃, 3.73; CH₃, 2.43.

(b) Stability of the tosyl protected base-residue in 35 TFA and HF.

The material was subjected to the standard deprotection conditions (TFA-deprotection) and the final cleavage

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conditions with HF. The products were then subjected to HPLC-analysis using a 4 μ RCM 8x10 Nova pack column and solvents A (0.1% TFA in water) and B (0.1% TFA in acetonitrile) according to the following time gradient with a flow of 2 ml/min.

	Time	% A	% B
	0	100	0
	5	100	0
	35	0	100
10	37	0	100
	39	100	0

The following retention times were found: (a) Compound 1: 30.77 min; (b) compound 2: 24.22 min; and (c) compound 3: 11.75 min. The analysis showed that the O6-benzyl group was removed both by TFA and HF, whereas there was no cleavage of the tosyl group in TFA, but quantitative removal in HF under the standard cleavage conditions.

EXAMPLE 109

20 5-Bromouracil-N¹-methyl acetate

5-Bromouracil (5.00 g; 26.2 mmol) and potassium carbonate (7.23 g; 52.3 mmol) were suspended in DMF (75 ml). Methyl bromoacetate (2.48 ml; 26.1 mmol) was added over a period of 5 min. The suspension was stirred for 2 h at room temperature, and then filtered. The solid residue was washed twice with DMF, and the combined filtrates were evaporated to dryness, *in vacuo*. The residue was an oil containing the title compound, DMF and some unidentified impurities. It is not necessary to purify the title compound before hydrolysis.

30 ¹H-NMR (DMSO-d₆, 250 MHz); 8.55 (impurity); 8.27 (CBr=CHN); 8.02 (impurity); 4.76 (impurity); 4.70 (impurity); 4.62 (NCH₂COOCH₃); 3.78 (COOCH₃); 2.96 (DMF); 2.80 (DMF). ¹³C-NMR (DMSO-d₆, 250 MHz); 168.8 (COOCH₃); 172.5 (CH=CBr=CON); 161.6 (DMF); 151.9 (NCON); 145.0 (CO-CBr=CHN); 95.6 (COCBr=CHN); 35 52.6 (impurity); 52.5 (OCH₃); 49.7 (impurity); 48.8 (NCH₂COOMe); 43.0 (impurity); 36.0 (DMF). UV(Methanol; _{max}nm); 226; 278. IR (KBr; cm⁻¹); 3158s (NH); 1743vs (C=O, COOMe); 1701vs (C=O, CONH); 1438vs (δ CH, CH₃O); 1223vs (C-O,

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COOMe); 864 m (δ CH, Br=C-H). FAB-MS m/z (assignment): 265/263 (M+H).

EXAMPLE 110

5 (5-Bromouracil)acetic acid

Water (30 ml) was added to the oil of the crude product from Example 109 and the mixture was dissolved by adding sodium hydroxide (2M, 60 ml). After stirring at 0°C for 10 min, hydrochloric acid (4M, 45 ml) was added to pH=2 and the title compound precipitated. After 50 min, the solid residue was isolated by filtration, washed once with cold water, and then dried in vacuo over sicapent. Yield: 2.46 g (38%). Mp, 250°-251°C. Anal. for C₆H₅BrN₂O₄. Found (calc.): C: 28.78 (28.94); H: 2.00 (2.02); Br: 32.18 (32.09); N: 11.29 (11.25). ¹H-NMR (DMSO-d₆, 250 MHz): 12.55 (1H,s,COOH); 11.97 (1H,s,NH); 8.30 (1H,s,C=C-H); 4.49 (2H,s,NCH₂COOH). ¹³C-NMR (DMSO-d₆, 250 MHz); 169.4 (COOH); 159.8 (NHCOBr=CH); 150.04 (NCON); 145.8 (COBr=CHN); 94.6 (COBr=CHN); 48.8 (NCH₂COOH). UV (Methanol; _{max}nm); 226; 278. IR (KBr; cm⁻¹); 3187s (NH); 1708vs (C=O,COOH); 1687vs; 1654VS (C=O, CONH); 1192s (C-O, COOH); 842 m (δ CH, Br-C=C-H). FAB-MS m/z (assignment, relative intensity); 251/249 (M + H, 5).

EXAMPLE 111

25 N-(Boc-aminoethyl)-N-(5-bromouracil)methylenecarbonoylglycine ethyl ester

Boc-aminoethylglycine ethyl ester (1.80 g; 7.30 mmol) was dissolved in DMF (10 ml). Dhbt-OH (1.31 g; 8.03 mmol) was added, whereby a precipitate was formed. DMF (2 x 10 ml) was added until the precipitate was dissolved. The product of Example 110 (2.00 g; 8.03 mmol) was added slowly to avoid precipitation. Methylene chloride (30 ml) was added, and the mixture was cooled to 0°C and then filtered. The precipitate, DCU, was washed twice with methylene chloride. To the combined filtrate was added methylene chloride (100 ml). The mixture was washed with half saturated NaHCO₃-solution (3 x 100 ml, H₂O:saturated NaHCO₃-solution 1:1 v/v), then with

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dilute KHSO_4 -solution ($2 \times 100 \text{ ml}$, H_2O :saturated KHSO_4 -solution 4:1 v/v), and finally with saturated NaCl -solution ($1 \times 100 \text{ ml}$). The organic phase was dried over magnesium sulphate, filtered, and evaporated to dryness *in vacuo* (about 15 mmHg
5 and then about 1 mmHg). The residue was suspended in methylene chloride (35 ml), stirred for 45 min at room temperature, and filtered (the precipitate was DCU). Petroleum ether (2 volumes) was added dropwise to the filtrate at 0°C , whereby an oil precipitated. The liquor was decanted
10 and the remaining oil dissolved in methylene chloride (20-50 ml). Precipitated was effected by the addition of petroleum ether (2 volumes). This procedure was repeated 5 times until an impurity was removed. The impurity can be seen at TLC with 10% $\text{MeOH}/\text{CH}_2\text{Cl}_2$ as the developing solvent. The resulting oil
15 was dissolved in methylene chloride (25 ml) and evaporated to dryness *in vacuo*, which caused solidification of the title compound. Yield: 2.03 g ((58%). Mp. $87^\circ\text{--}90^\circ\text{C}$. Anal. for $\text{C}_{17}\text{H}_{25}\text{BrN}_4\text{O}_7$. Found (calc.): C: 42.33 (42.78); H: 5.15 (5.28); Br: 17.20 (16.74); N: 1.69 (11.74). $^1\text{H-NMR}$ (DMSO-d_6 , 250 MHz,
20 J in Hz): 1.93 & 11.92 (1H, s, C=ONHC=O); 8.09 & 8.07 (1H, s, C=C-H); 7.00 & 6.80 (1H, t, b, BocNH); 4.80 & 4.62 (2H, s, NCH_2CON); 4.35 & 4.24 (2H, s, NCH_2COOEt); 4.27-4.15 (2H, m's, $\text{COOCH}_2\text{CH}_3\text{O}$); 3.47-3.43 (2H, m's, $\text{BocNHCH}_2\text{CH}_2\text{N}$); 3.28-
3.25 & 3.12-3.09 (2H, m's, $\text{BocNHCH}_2\text{CH}_2\text{N}$): 1.46 & 1.45
25 (9H, s, $t\text{-Bu}$); 1.26 & 1.32 (3H, t, J=7.1, $\text{COOCH}_2\text{CH}_3$). $^{13}\text{C-NMR}$ (DMSO-d_6 , 250 MHz); 169.3 & 169.0 ($t\text{-BuOC=O}$); 167.4 & 167.1 (COOEt); 159.8 (C=C-CON); 155.9 (NCH_2CON); 150.4 (NCON); 145.9 ($\text{COCB}-\text{CHN}$); 94.5 ($\text{COCB}=CHN$); 78.2 (Me_3C); 61.3 & 60.7 (COCH_2CH_3); 49.1 & 48.0 (NCH_2COOH); 48.0 & 47.0 (NCH_2CON); 38.6
30 (BocNHCH₂CH₂N); 38.2 (BocNHCH₂CH₂N); 26.3 ($\text{C}(\text{CH}_3)_3$); 14.1 (COCH_2CH_3). UV (Methanol; λ_{max} NM): 226; 280. IR (KBr, cm^{-1}): 3200ms, broad (NH); 168vs, vbroad (C=O , COOH , CONH); 1250s (C-O , COOEt); 1170s (C-O , COO^tBu); 859m ($\delta \text{ CH}$, Br-C=C-H). FAB-MS m/z (assignment, relative intensity): 479/477 ($\text{M} + \text{H}$, 5); 423/421 ($\text{M} + 2\text{H} - t\text{-Bu}$, 8); 379/377 ($\text{M} + 2\text{H} - \text{Boc}$, 100); 233/231 ($\text{M} - \text{backbone}$, 20).

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EXAMPLE 112

N-(Boc-aminoethyl)-N-(5-bromouracyl-N'-methylenecarbonoyl)-glycine

The product of Example 111 (1.96 g; 4.11 mmol) was dissolved in methanol (30 ml) by heating, and then cooled to 0°C. Sodium hydroxide (2M, 30 ml) was added, and the mixture stirred for 30 min. HCl (1M, 70 ml) was added to pH = 2.0. The water phase was extracted with ethyl acetate (3 x 65 ml + 7 x 40 ml). The combined ethyl acetate extractions were washed with saturated NaCl-solution (500 ml). The ethyl acetate phase was dried over magnesium sulphate, filtered and evaporated to dryness in vacuo. Yield: 1.77 g (96%). Mp. 92°-97°C. Anal. for C₁₅H₂₁BrN₄O₇. Found (calc.): C: 40.79 (40.10); H: 5.15 (4.71); Br: 14.64 (17.70); N: 11.35 (12.47). ¹H-NMR (DMSO-d₆, 250 MHz, J in Hz): 12.83 (1H, s, COOH); 11.93 & 11.91 (1H, s, C=ONHC=O); 8.10 & 8.07 (1H, s, C=C-H); 7.00 & 6.81 (1H, t, b, BocNH); 4.79 & 4.61 (2H, s, NCH₂CON); 4.37 & 4.25 (2H, s, NCH₂COOH); 3.46-3.39 (2H, m's, BocNHCH₂CH₂N); 3.26-3.23 & 3.12-3.09 (2H, m's, BocNHCH₂CH₂N); 1.46 (9H, s, ^tBu). ¹³C-NMR (9DMSO-d₆, 250 MHz); 170.4 (^tBuOC=O); 166.9(COOH); 159.7 (C=C-CON); 155.8 (NCH₂CON); 150.4 (NCON); 145.9 (COCB_r=CHN); 94.4 (CO_CBr=CHN); 78.1 (Me₃C); 49.1 & 48.0 (NCH₂COOH); 47.7 & 47.8 (NCH₂CON); 38.6 (BocNHC₂CH₂N); 38.1 (Boc NHCH₂CH₂N); 28.2 (C(CH₃)₃). UV (Methanol; _{max} nm); 226; 278. IR (KBr, cm⁻¹): 3194ms, broad (_NH); 1686vs, vbroad (_C=O COOH, CONH); 1250s (_C-O, COOH); 1170s (_C-O, COO^tBu); 863m (δ CH, Br-C=C-H). FAB-MS m/z (assignment, relative intensity): 449/451 (M + H, 70); 349/351 (M + 2H -Boc, 100); 231/233 (M - backbone, 20).

30 EXAMPLE 113

Uracil-N¹-methyl acetate

Uracil (10.0 g; 89.2 mmol) and potassium carbonate (24.7 g; 178 mmol) were suspended in DMF (250 ml). Methyl bromoacetate (8.45 ml; 89.2 mmol) was added over a period of 5 min. The suspension was stirred overnight under nitrogen at room temperature, and then filtered. TLC (10% methanol in ethylene chloride) indicated incomplete conversion of uracil.

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The solid residue was washed twice with DMF, and the combined filtrates were evaporated to dryness in vacuo. The precipitate was suspended in water (60 ml) and HCl (2.5 ml, 4M) was added to pH = 2. The suspension was stirred for 30 min at 0°C, and then filtered. The precipitated title compound was washed with water and dried, in vacuo, over sicapent. Yield: 9.91 g (60%). Mp. 182° - 183°C. Anal. for C₆H₈N₂O₄. Found (calc.): C: 45.38 (45.66); H: 4.29 (4.38); N: 15.00 (15.21). ¹H-NMR (DMSO-d₆, 250 MHz, J in Hz): 1.47 (1H, s, NH); 7.68 (1H, d, J_{H-C=C-H}=7.9), CH=CHN; 5.69 (1H, d, J_{H-C=C-H}=7.9), CH=CHN; 4.59 (2H, s, NCH₂COOMe); 3.76 (3H, s, COOCH₃). ¹³C-NMR (DMSO-d₆, 250 MHz); 168.8 (COOMe); 164.0 (C=C-CON); 151.1 (NCON); 146.1 (COCH=CHN); 101.3 (COCH=CHN); 52.5 (COOCH₃); 48.7 (NCH₂COOMe). UV (Methanol; _{max}nm): 226; 261. IR (KBr; cm⁻¹): 3164s (NH); 1748vs (C=O, COOMe); 1733vs (C=O, CONH); 1450vs (δ CH, CH₃O); 1243VS (C-O, COOMe); 701m (δ CH, H-C=C-H). FAB-MS m/z (assignment); 185 (M+H).

EXAMPLE 114

20 Uracilacetic acid

Water (90 ml) was added to the product of Example 113 (8.76 g; 47.5 mmol), followed by sodium hydroxide (2M, 40 ml). The mixture was heated for 40 min, until all the methyl ester has reacted. After stirring at 0°C for 15 min, hydrochloric acid (4M, 25 ml) was added to pH=2. The title compound precipitated and the mixture was filtered after 2-3 h. The precipitate was washed once with the mother liquor and twice with cold water and dried in vacuo over sicapent. Yield: 6.66 g (82%). Mp. 288°-289°C. Anal. for C₆H₆N₂O₄. Found 30 (calc.): C: 42.10 (42.36), H: 3.43 (3.55); N: 16.25 (16.47)/ ¹H-NMR (DMSO-d₆, 250 MHz, J in Hz): 13.19 (1H, s, COOH); 11.41 (1H, s, NH); 7.69 (1H, d, J_{H-C=C-H}=7.8, J_{H-C-C-N-H}=2.0, COCH=CHN); 4.49 (2H, s, NCH₂COOH). ¹³C-NMR (DMSO-d₆, 2509 MHz); 169.9 (COOH); 163.9 (CH=CHCON); 151.1 (NCON); 146.1 (COCH=CHN); 100.9 35 (COCH=CHN); 48.7 NCH₂COOH. UV (Methanol; _{max}nm): 246; 263. IR (KBr; cm⁻¹): 3122s (NH); 1703vs (C=O, COOH); 1698vs,

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1692vs ($\text{C}=\text{O}$, CONH); 1205s ($\text{C}-\text{O}, \text{COOH}$); 676 (δ CH, H-C=C-H).
FAB-MS m/z (assignment): 171 (M + H).

EXAMPLE 115

5 N-(Bocaminoethyl)-N-(uracil-N¹-methylenecarbonoyl)glycine ethyl ester

(Bocaminoethyl)glycine ethyl ester (2.00 g; 8.12 mmol) was dissolved in DMF (10 ml). Dhbt-OH (1.46 g; 8.93 mmol) was added and a precipitate was formed. DMF (2 x 10 ml) was added 10 until all was dissolved. The product of Example 114 (1.52 g; 8.93 mmol) was added slowly to avoid precipitation. Methylene chloride (30 ml) was added, and the mixture was cooled to 0°C, whereafter DDC (2.01g; 9.74 mmol) was added. The mixture was stirred for 1 h at 0°C, at 2 h at room temperature, and then 15 filtered. The precipitated DCU was washed twice with methylene chloride. To combined filtrate was added methylene chloride (100 ml), and the solution washed with half-saturated NaHCO₃-solution (3 x 100 ml, H₂O:saturated NaHCO₃-solution 1:1 v/v), then with dilute KHSO₄-solution (2 x 100 ml, 20 H₂O:saturated KHSO₄-solution 4:1 v/v) and finally with saturated NaCl-solution (1 x 100 ml). The organic phase was dried over magnesium sulphate, filtered and evaporated to dryness in vacuo (about 15 mmHg and then about 1mmHg). The residue was suspended in methylene chloride (32 ml), and 25 stirred for 35 min at room temperature, and 30 min at 0°C, and then filtered. The precipitate (DCU) was washed with methylene chloride. Petroleum ether (2 volumes) was added dropwise to the combined filtrate at 0°C, which caused separation of an oil. The mixture was decanted, the remaining 30 oil was then dissolved in methylene chloride (20 ml), and then again precipitated by addition of petroleum ether (2 volumes). This procedure was repeated 5 times until an impurity was removed. The impurity can be seen by TLC with 10% MeOH/CH₂Cl₂ as the developing solvent. The resulting oil was dissolved 35 in methylene chloride (20 ml) and evaporated to dryness in vacuo, which caused solidification of the title compound. Yield: 1.71 g (53%). Mp. 68.5° - 75.7°C. Anal for C₁₇H₂₆N₄O₇.

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Found (calc.): C: 50.61 (51.25); H: 6.48 (6.58); N: 13.33 (14.06). $^1\text{H-NMR}$ (DMSO-d₆, 250 MHz, J in Hz): 11.36 (1H, s, C=ONHC=O); 7.51 & 7.47 (1H, d, J_{H-C=C-H} 6.1; COCH=X-H); 7.00 & 6.80 (1H, t, b, BocNH); 5.83 & 5.66 (1H, d, J_{H-C=C-H} = 5.7, 5 COCH=CH); 4.78 & 4.60 (2H, s, NCH₂CON); 4.37 & 4.12 (2H, s, NCH₂COOEt); 4.30 - 4.15 (2H, m's, COOCH₂CH₃); 3.49-3.46 (2H, m's, BocNHCH₂CH₂n); 3.27 3.23 & 3.11-3.09 (2H, m's, BocNHCH₂CH₂N); 1.46 (9H, s, ^tBu); 1.39-1.23 (3H, m's, COOCH₂CH₃). $^{13}\text{C-NMR}$ (DMSO-d₆, 250 MHz): 169.4 & 169.0 (^tBuOC=O); 167.6 & 10 167.3 (COOEt); 163.8 (CH=CHCON); 155.8 (NCH₂CON); 151.0 (NCON); 146.3 (COCH=CHN); 100.8 (COCH=CHN); 78.1 (Me₃C); 61.2 & 60.6 (COOCH₂CH₃); 49.1 (NCH₂COOEt); 47.8 & 47.0 (NCH₂CON); 38.6 (BocNHCH₂CH₂N); 38.1 & 37.7 (BocNHCH₂N); 28.2 (C(CH₃)₃); 14.1 (CO-OCH₂CH₃). UV (Methanol; _{max} nm); 226; 264. IR (KBr; cm⁻¹): 3053m (NH); 1685vs, v broad (C=O, COOH, CONH); 1253s (C-O, COOEt); 1172s (C-O, COO^tBu); 718w (δ CH, C-C-C-H), FAB-MS m/z (assignment, relative intensity); 399 (M + H, 35); 343 (M + 2H - ^tBu, 100); 299 (M + 2H - Boc, 100); 153 (M-backbone, 30).

20

EXAMPLE 116

N-(Bocaminoethyl)-N-(uracilmethylene carbonoyl)glycine

The product of Example 115 (1.56 g; 3.91 mmol) was dissolved in methanol (20 ml) and then cooled to 0°C. Sodium 25 hydroxide (2M, 20 ml) was added, and the mixture was stirred for 75 min at 0°C. Hydrochloric acid (1M, 46 ml) was added to pH = 2.0. The water phase was extracted with ethyl acetate (3 X 50 ml + 7 x 30 ml). The combined ethyl acetate extractions were washed with saturated NaCl solution (360 ml). 30 The ethyl acetate phase was dried over magnesium sulphate, filtered, and evaporated to dryness, *in vacuo*. The residue was dissolved in methanol and evaporated to dryness, *in vacuo*. Yield: 0.55 g (38%). Mp 164° - 170°C. Anal. for C₁₅H₂₂N₄O₇. Found (calc.): C: 46.68 (48.65); H: 6.03 (5.99); N: 1461 (15.13). $^1\text{H-NMR}$ (DMSO-d₆, 250 MHz, J in Hz); 12.83 (1H, s, COOH); 11.36 (1H, s, C=ONHC=O); 7.52-7.45 (1H, m's, COCH=CHN); 7.00 & 6.82 (1H, t, b, BocNH); 5.67-5.62 (1H, m's, COCH=CHN);

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4.76 & 4.58 (2H, s, NCH₂CON); 4.26 & 4.05 (2H, s, NCH₂COOH); 3.46-3.39 (2H, m's, BocNHCH₂CH₂N); 3.25-3.23 & 3.15-3.09 (2H, m's, BocNHCH₂CH₂N); 1.46 (9H, s, ^tBu). ¹³C-NMR (DMSO-d₆, 250 MHz); 170.5 (^tBuOC=O); 167.2 (COOH); 163.9 (C=C-CON); 155.8 5 (NCH₂CON); 151.1 (NCON); 146.4 (COCH=CHN); 100.8 (COCH=CHN); 78.1 (Me₃C); 49.1 & 47.8 (NCH₂ COOH); 47.6 & 46.9 (NCH₂CON); 38.6 (BocNHCH₂CH₂N); 38.1 & 37.6 (BocNHCH₂CH₂N); 28.2 (C(CH₃)₃). UV (Methanol; _{max} nm); 226; 264. IR (KBr; cm⁻¹); 3190 (NH); 1685vs, vbroad (C=O, COOH, CONH); 1253s (C-O, COOH); 1171s 10 (C-O, COO^tBu); 682w (δ CH, H-C=C-H). FAB-MS m/z (assignment, relative intensity): 371 (M + H, 25); 271 (M + H -Boc, 100).

EXAMPLE 117

H-U10-LysNH₂

Synthesis of the title compound was accomplished by 15 using "synthetic Protocol 10". The synthesis was initiated on approximately 100 mg Lys (ClZ)-MHBA-resin. The crude product (12 mg) was pure enough for hybridization studies. The hybrid between 5'-(dA)10 and H-U10 had Tm of 67.5°C.

EXAMPLE 118

20 Deprotection and Cleavage of H-[Cacg]₁₀-Lys-NH₂ by Trifluoromethansulfonic Acid (TFMSA). An Alternative Method to Deprotection and Cleavage by Hydrogen Fluoride (HF).

(a) Deprotection of Side-Chain Protecting Groups by a "Low-Acidity" TFMSA-TFA-DMS Procedure

25 A portion of ca. 0.4 g wet Boc-[Cacg]₁₀-Lys(ClZ)-MBHA resin (prepared in one of the previous examples) was placed in a 5 ml solid-phase reaction vessel. The n-Terminal Boc group was removed by the following protocol: (1) 50% TFA/CH₂Cl₂, 2 x 1 min and 1 x 30 min; (2) 100% TFA, 2 x 1 min 30 and drain. In order to deprotect the benzyl-based side-chain protecting groups a so-called "low-acidity" TFMSA procedure was carried out as follows: A stock solution (A) containing 5 ml of TFA-DMS-m-cresol (2:6:2, v/v/v) and a stock solution (B) containing TFA-TFMSA (8:2, v/v) were prepared. Next, the 35 following steps were carried out: (3) 1 ml of stock solution (A) is added to the PNA-resin in the reaction vessel with shaking for 2 min. No drain; (4) 1 ml of stock solution (B) (cooled with ice/water) is added in portions of 200 μ l every

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10th minute over a period of 40 min, and shaking is continued for another 50 min; (5) drain and washing with 100% TFA, 5 x 1 min. and drain.

(b) Cleavage from the Resin by a "High-Acidity"
TFMSA-TFA Procedure

5 In order to cleave the above-mentioned deprotected PNA from the resin a so-called "high acidity" TFMSA procedure was carried out as follows: A stock solution (C) containing m-cresol-TFA (2:8, v/v) was prepared. Next, the following steps 10 were carried out: (6) 1 ml of stock solution (C) was added to the deprotected PNA-resin in the SPPS vessel with shaking for 2 min; (7) 1 ml of stock solution (B) (cooled with ice/water) is added in portions of 200 μ l over a period of 30 min and shaking is continued for another 150 min; (8) the 2 ml solution 15 in the reaction vessel is "blown out" through the filter into a 20 ml solution of diethylether cooled with dry ice/isopropanol. In order to complete the precipitation process, 200 μ l of anhydrous pyridine is added dropwise to the acid-ether mixture; (9) centrifugation at 3000 rpm for 5 20 min; (9) the supernatant is decanted and the precipitate is washed three times with cold diethylether, dried, dissolved in water, and lyophilized.

(c) Purification and Identification of H-[Caeg]₁₀-Lys-NH₂

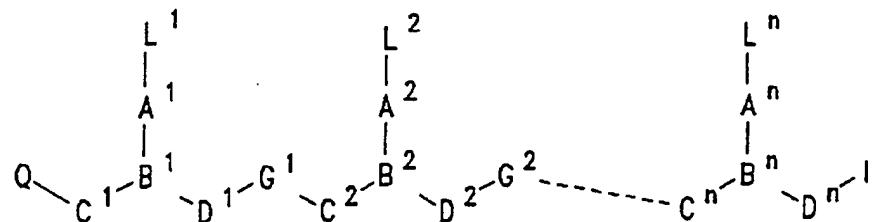
25 An analytical HPLC chromatogram showed a nice crude product of good purity and a profile almost identical to that obtained from the HF cleavage of H-[Caeg]₁₀-Lys-NH₂, except that an additional peak, of course, arising from pyridine TFMSA salt elutes early in the chromatogram. Purification and 30 identification was carried out by the usual procedures.

Those skilled in the art will appreciate that numerous changes and modifications may be made to the preferred embodiments of the invention and that such changes and modifications may be made without departing from the spirit 35 of the invention. It is therefore intended that the appended claims cover all such equivalent variations as fall within the true spirit and scope of the invention.

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WHAT IS CLAIMED IS:

1. A compound comprising a polyamide backbone bearing a plurality of ligands that are individually bound to aza nitrogen atoms located within said backbone, at least one of said ligands being a naturally occurring nucleobase, a non-naturally occurring nucleobase, a DNA intercalator, or a nucleobase-binding group.
2. The compound of claim 1 wherein said aza nitrogen atoms are separated from one another in said backbone by from 4 to 6 intervening atoms.
3. A compound having the formula:

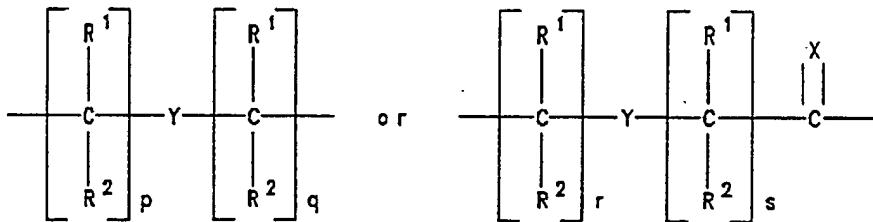


wherein:

n is at least 2,

each of L¹-Lⁿ is independently selected from the group consisting of hydrogen, hydroxy, (C₁-C₄) alkanoyl, naturally occurring nucleobases, non-naturally occurring nucleobases, aromatic moieties, DNA intercalators, nucleobase-binding groups, heterocyclic moieties, and reporter ligands, at least one of L¹-Lⁿ being a naturally occurring nucleobase, a non-naturally occurring nucleobase, a DNA intercalator, or a nucleobase-binding group;

each of A¹-Aⁿ is a single bond, a methylene group or a group of formula:



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where:

X is O, S, Se, NR³, CH₂ or C(CH₃)₂;

Y is a single bond, O, S or NR⁴;

each of p and q is zero or an integer from 1 to 5, the sum p+q being not more than 10;

each of r and s is zero or an integer from 1 to 5, the sum r+s being not more than 10;

each R¹ and R² is independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl which may be hydroxy- or alkoxy- or alkylthio-substituted, hydroxy, alkoxy, alkylthio, amino and halogen; and

each R³ and R⁴ is independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl, hydroxy- or alkoxy- or alkylthio-substituted (C₁-C₄)alkyl, hydroxy, alkoxy, alkylthio and amino;

each of B¹-Bⁿ is N or R³N⁺, where R³ is as defined above;

each of C¹-Cⁿ is CR⁶R⁷, CHR⁶CHR⁷ or CR⁶R⁷CH₂, where R⁶ is hydrogen and R⁷ is selected from the group consisting of the side chains of naturally occurring alpha amino acids, or R⁶ and R⁷ are independently selected from the group consisting of hydrogen, (C₂-C₆)alkyl, aryl, aralkyl, heteroaryl, hydroxy, (C₁-C₆)alkoxy, (C₁-C₆)alkylthio, NR³R⁴ and SR⁵, where R³ and R⁴ are as defined above, and R⁵ is hydrogen, (C₁-C₆)alkyl, hydroxy-, alkoxy-, or alkylthio- substituted (C₁-C₆)alkyl, or R⁶ and R⁷ taken together complete an alicyclic or heterocyclic system;

each of D¹-Dⁿ is CR⁶R⁷, CH₂CR⁶R⁷ or CHR⁶CHR⁷, where R⁶ and R⁷ are as defined above;

each of G¹-Gⁿ⁻¹ is -NR³CO-, -NR³CS-, -NR³SO- or -NR³SO₂-, in either orientation, where R³ is as defined above;

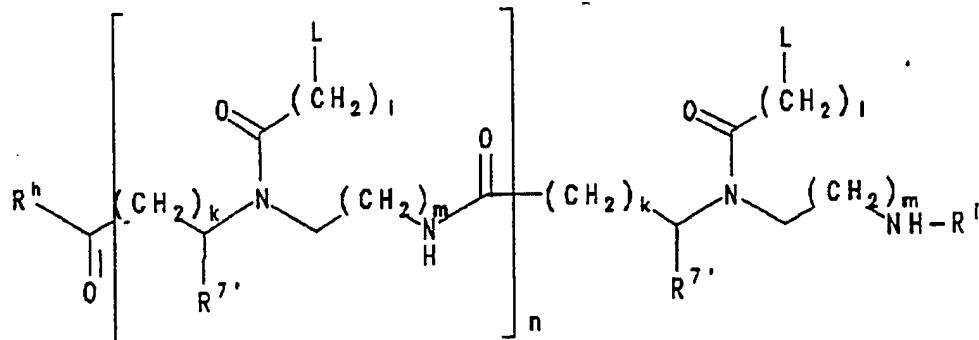
Q is -CO₂H, -CONR'R'', -SO₃H or -SO₂NR'R'' or an activated derivative of -CO₂H or -SO₃H; and

I is -NHR'''R'''' or -NR'''C(O)R''''', where R', R'', R''' and R'''' are independently selected from the group consisting of hydrogen, alkyl, amino protecting groups, reporter ligands, intercalators, chelators, peptides,

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proteins, carbohydrates, lipids, steroids, oligonucleotides and soluble and non-soluble polymers.

4. The compound of claim 3 having the formula:



wherein:

each L is independently selected from the group consisting of hydrogen, phenyl, heterocyclic moieties, naturally occurring nucleobases, and non-naturally occurring nucleobases;

each R' is independently selected from the group consisting of hydrogen and the side chains of naturally occurring alpha amino acids;

n is an integer from 1 to 60,

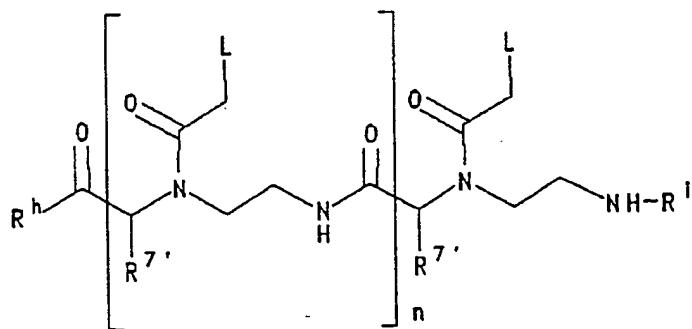
each k and m is, independently, zero or 1;

each l is zero or an integer from 1 to 5;

R^h is OH, NH₂ or -NHLysNH₂; and

Rⁱ is H or COCH₃.

5. The compound of claim 4 having formula:



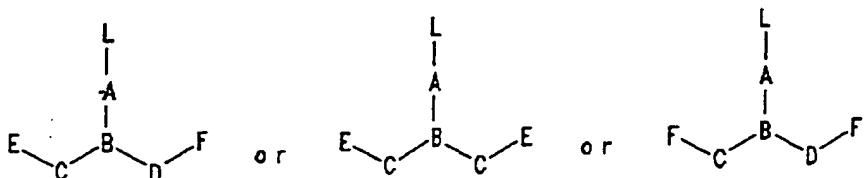
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each L is independently selected from the group consisting of the nucleobases thymine, adenine, cytosine, guanine, and uracil;

each R⁷ is hydrogen; and

n is an integer from 1 to 30.

6. A compound having one of the following formulas:



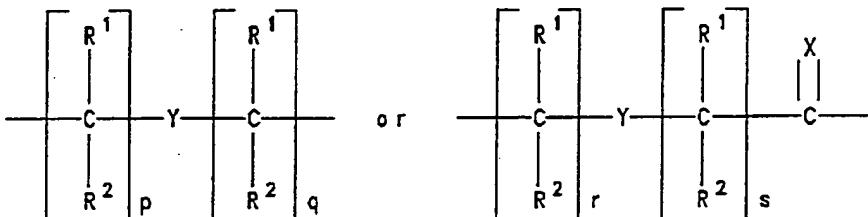
wherein:

L is selected from the group consisting of hydrogen, hydroxy, (C₁-C₄) alkanoyl, naturally occurring nucleobases, non-naturally occurring nucleobases, aromatic moieties, DNA intercalators, nucleobase-binding groups, and heterocyclic moieties, reporter ligands, wherein:

at least one of L¹-Lⁿ is a naturally occurring nucleobase, a non-naturally occurring nucleobase, a DNA intercalator, or a nucleobase-binding group; and

amino groups are, optionally, protected by amino protecting groups;

A is a single bond or a group of the formula:



where:

X is O, S, Se, NR³, CH₂ or C(CH₃)₂;

Y is a single bond, O, S or NR⁴;

each of p and q is zero or an integer from 1 to 5, the sum p+q being not more than 10;

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each of r and s is zero or an integer from 1 to 5, the sum r+s being not more than 10;

each R¹ and R² is independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl, hydroxy- or alkoxy- or alkylthio-substituted (C₁-C₄)alkyl, hydroxy, alkoxy, alkylthio, amino and halogen; and

each R³ and R⁴ is independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl, hydroxy- or alkoxy- or alkylthio-substituted (C₁-C₄)alkyl, hydroxy, alkoxy, alkylthio and amino;

B is N or R³N⁺, where R³ is as defined above;

each C is CR⁶R⁷, CHR⁶CHR⁷ or CR⁶R⁷CH₂, where R⁶ is hydrogen and R⁷ is selected from the group consisting of the side chains of naturally occurring alpha amino acids, or R⁶ and R⁷ are independently selected from the group consisting of hydrogen, (C₂-C₆)alkyl, aryl, aralkyl, heteroaryl, hydroxy, (C₁-C₆)alkoxy, (C₁-C₆)alkylthio, NR³R⁴ and SR⁵, where R³ and R⁴ are as defined above, and R⁵ is hydrogen or (C₁-C₆)alkyl, hydroxy-, alkoxy-, or alkylthio- substituted (C₁-C₆)alkyl, or R⁶ and R⁷ taken together complete an alicyclic or heterocyclic system;

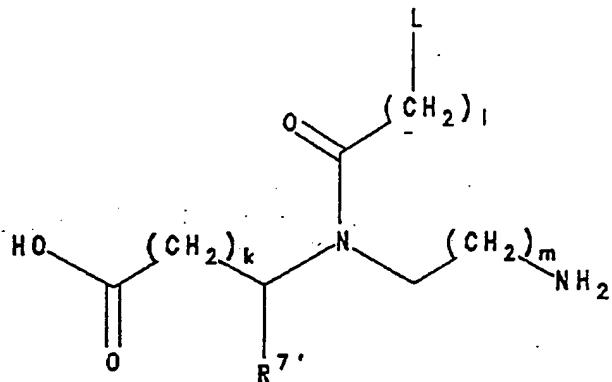
each D is CR⁶R⁷, CH₂CR⁶R⁷ or CHR⁶CHR⁷, where R⁶ and R⁷ are as defined above;

each E is COOH, CSOH, SOOH, SO₂OH or an activated or protected derivative thereof; and

each F is NHR³ or NPgR³, where R³ is as defined above, and Pg is an amino protecting group.

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7. The compound of claim 6 having the formula:



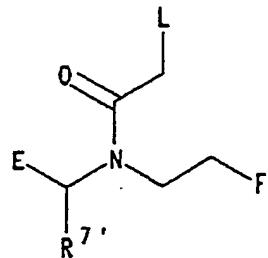
wherein:

each L is independently selected from the group consisting of hydrogen, phenyl, heterocyclic moieties, naturally occurring nucleobases, and non-naturally occurring nucleobases;

each R^{7'} is independently selected from the group consisting of hydrogen and the side chains of naturally occurring alpha amino acids; and

each k, l, and m is, independently, zero or an integer from 1 to 5.

8. The compound of claim 7 having formula:



wherein:

L is selected from the group consisting of the nucleobases thymine, adenine, cytosine, guanine, uracil, 5-methylcytosine, 6-thioguanine and 5-bromouracil, and protected derivatives thereof;

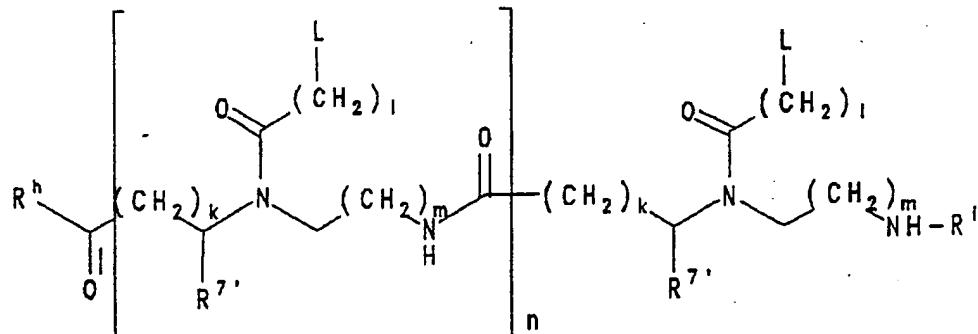
R^{7'} is hydrogen;

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E is COOH or an activated or protected derivative thereof; and

F is NH₂ or NHPg, where Pg is an amino protecting group.

9. A compound having the formula:



wherein:

each L is independently selected from the group consisting of hydrogen, phenyl, heterocyclic moieties, naturally occurring nucleobases, and non-naturally occurring nucleobases;

each R' is independently selected from the group consisting of hydrogen and the side chains of naturally occurring alpha amino acids;

n is an integer from 1 to 60,

each k , l , and m is, independently, zero or an integer from 1 to 5;

R^h is OH, NH₂ or -NHLysNH₂; and

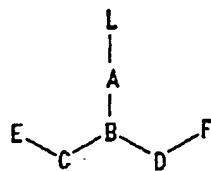
R^i is H or COCH_3 .

10. A process for preparing a compound according to claim 1, comprising the steps of:

A) providing a polymer substrate, said polymer being functionalized with a chemical group capable of forming an anchoring linkage with an amino acid;

B) coupling said polymer with a first amino acid through said anchoring linkage, said first amino acid having formula (IV):

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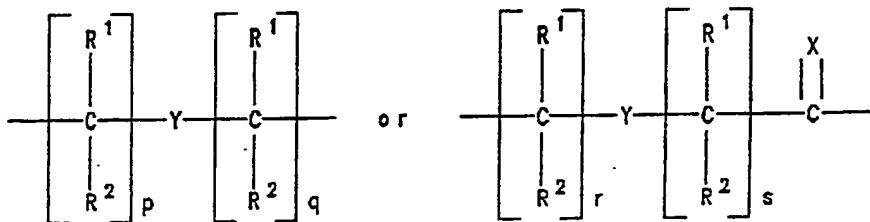


(IV)

wherein:

L is selected from the group consisting of naturally occurring nucleobases, non-naturally occurring nucleobases, aromatic moieties, DNA intercalators, nucleobase-binding groups, heterocyclic moieties, and reporter ligands, wherein amino groups are, optionally, protected by amino protecting groups;

A is a single bond or a group of the formula:



where:

X is O, S, Se, NR³, CH₂ or C(CH₃)₂;

Y is a single bond, O, S or NR⁴;

p and q are zero or integers from 1 to 5, the sum p+q being not more than 10;

r and s are zero or integers from 1 to 5, the sum r+s being not more than 10;

R¹ and R² are independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl, hydroxy- or alkoxy- or alkylthio-substituted (C₁-C₄)alkyl, hydroxy, alkoxy, alkylthio, amino and halogen; and

R³ and R⁴ are independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl, hydroxy- or alkoxy- or alkylthio-substituted (C₁-C₄)alkyl, hydroxy, alkoxy, alkylthio and amino;

B is N or R³N⁺, where R³ is as defined above;

C is CR⁶R⁷, CHR⁶CHR⁷ or CR⁶R⁷CH₂, where R⁶ is hydrogen and R⁷ is selected from the group consisting

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of the side chains of naturally occurring alpha amino acids, or R⁶ and R⁷ are independently selected from the group consisting of hydrogen, (C₂-C₆)alkyl, aryl, aralkyl, heteroaryl, hydroxy, (C₁-C₆)alkoxy, (C₁-C₆)alkylthio, NR³R⁴ and SR⁵, where R³ and R⁴ are as defined above, and R⁵ is hydrogen or (C₁-C₆)alkyl, hydroxy-, alkoxy-, or alkylthio- substituted (C₁-C₆)alkyl, or R⁶ and R⁷ taken together complete an alicyclic or heterocyclic system;

D is CR⁶R⁷, CH₂CR⁶R⁷ or CHR⁶CHR⁷, where R⁶ and R⁷ are as defined above;

E is COOH or an activated or protected derivative thereof; and

F is NPgR³ where R³ is as defined above and Pg is an amino protecting group;

C) removing said amino protecting group from said coupled first amino acid to generate a free amino group; and

D) reacting said free amino group with a second amino acid having formula (IV) to form a peptide chain.

11. The process of claim 10 further comprising the steps of:

E) removing said amino protecting group from said second amino acid to generate a terminal free amino group on said peptide chain; and

F) reacting said free amino group on said peptide chain with a further amino acid having formula (IV) to lengthen said peptide chain.

12. The process of claim 11 wherein steps E and F are performed a plurality of times.

13. The process of claim 11 further comprising removing at least one protecting group remaining on the amino acid moieties of the peptide chain.

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14. The process of claim 10 further comprising cleaving said anchoring linkage without substantially degrading said peptide chain.

15. The process of claim 10 wherein the polymer substrate contains polystyrene, polyacrylamide, silica, a composite material, cotton, or a derivative thereof.

16. The process of claim 10 wherein the chemical group capable of forming said anchoring linkage is chloro-, bromo- and iodo-substituted alkyl, amino-substituted alkyl, amino and aryl-substituted alkyl, amino- and alkylaryl-substituted alkyl, hydroxy-substituted alkyl, or a derivative thereof having a spacer group that can be cleaved substantially without degradation of said polypeptide.

17. The process of claim 16 wherein chloro-substituted alkyl is chloromethyl, amino-substituted alkyl is aminomethyl, amino- and alkyl-substituted aryl is α -aminobenzyl, amino- and alkylaryl-substituted alkyl is selected from the group consisting of α -amino-3- and α -amino-4-methylbenzyl, and hydroxy-substituted alkyl is hydroxymethyl.

18. The process of claim 16 wherein:

the chemical group is derived from an amino-containing moiety selected from amino-substituted alkyl, amino- and aryl substituted alkyl, and amino- and alkylaryl-substituted alkyl; and

the chemical group includes a spacer group derived from the group consisting of 4-(haloalkyl)aryl-lower alkanoic acids, Boc-aminoacyl-4-(oxymethyl)aryl-lower alkanoic acids, N-Boc-p-acylbenzhydrylamines, N-Boc-4'-(lower alkyl)-p-acylbenzhydrylamines, N-Boc-4'-(lower alkoxy)-p-acylbenzhydrylamines, and 4-hydroxymethylphenoxy-lower alkanoic acids.

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19. A process for sequence-specific recognition of a double-stranded polynucleotide, comprising contacting said polynucleotide with a compound that is different from natural RNA and that binds to one strand of the polynucleotide, thereby displacing the other strand.

20. The process of claim 19 wherein said compound is an oligomer comprising a homogenous or heterogeneous backbone to which are linked naturally occurring nucleobases, non-naturally occurring nucleobases or other ligands that individually bind by hydrogen to at least one natural nucleobase in said bound polynucleotide strand.

21. The process of claim 20 wherein said compound is the compound of claim 1.

22. The process of claim 20 wherein said compound is the compound of claim 4.

23. A process for modulating the expression of a gene in an organism, comprising administering to said organism a compound according to claim 1 that specifically binds to DNA or RNA deriving from said gene.

24. The process of claim 23 wherein said compound is the compound of claim 1.

25. The process of claim 23 wherein said compound is the compound of claim 4.

26. The process of claim 23 wherein said modulation includes inhibiting transcription of said gene.

27. The process of claim 23 wherein said modulation includes inhibiting replication of said gene.

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28. A process for treating conditions associated with undesired protein production in an organism, comprising contacting said organism with an effective amount of a compound according to claim 1 that specifically binds with DNA or RNA deriving from a gene controlling said protein production.

29. The process of claim 28 wherein said compound is the compound of claim 1.

30. The process of claim 28 wherein said compound is the compound of claim 4.

31. A process for inducing degradation of DNA or RNA in cells of an organism, comprising administering to said organism a compound according to claim 1 that specifically binds to said DNA or RNA.

32. A process for killing cells or virus, comprising contacting said cells or virus with a compound according to claim 1 that specifically binds to a portion of the genome of said cells or virus.

33. A pharmaceutical composition comprising a compound according to claim 1 and at least one pharmaceutically effective carrier, binder, thickener, diluent, buffer, preservative, or surface active agent.

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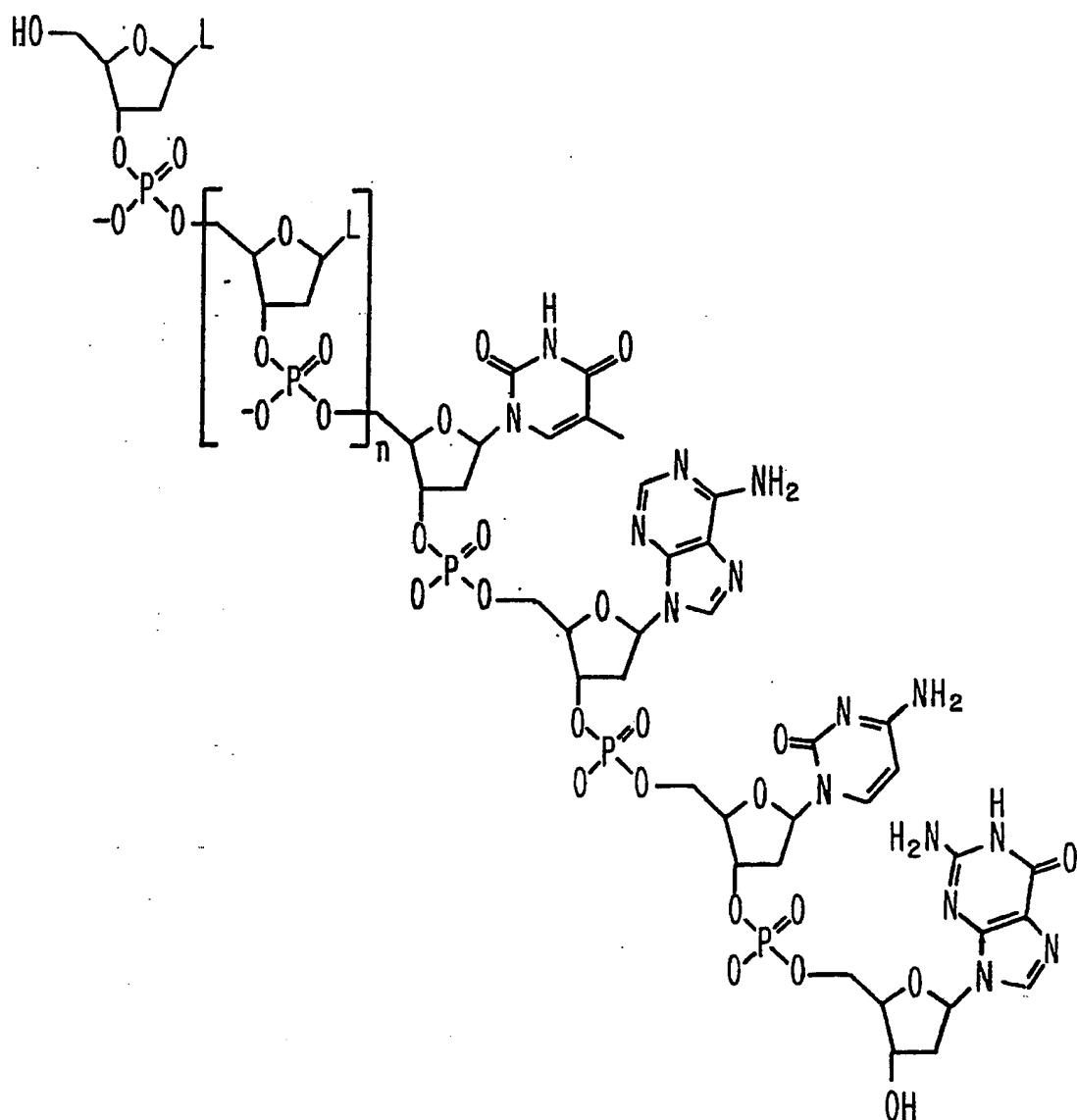


Fig. 1(A)

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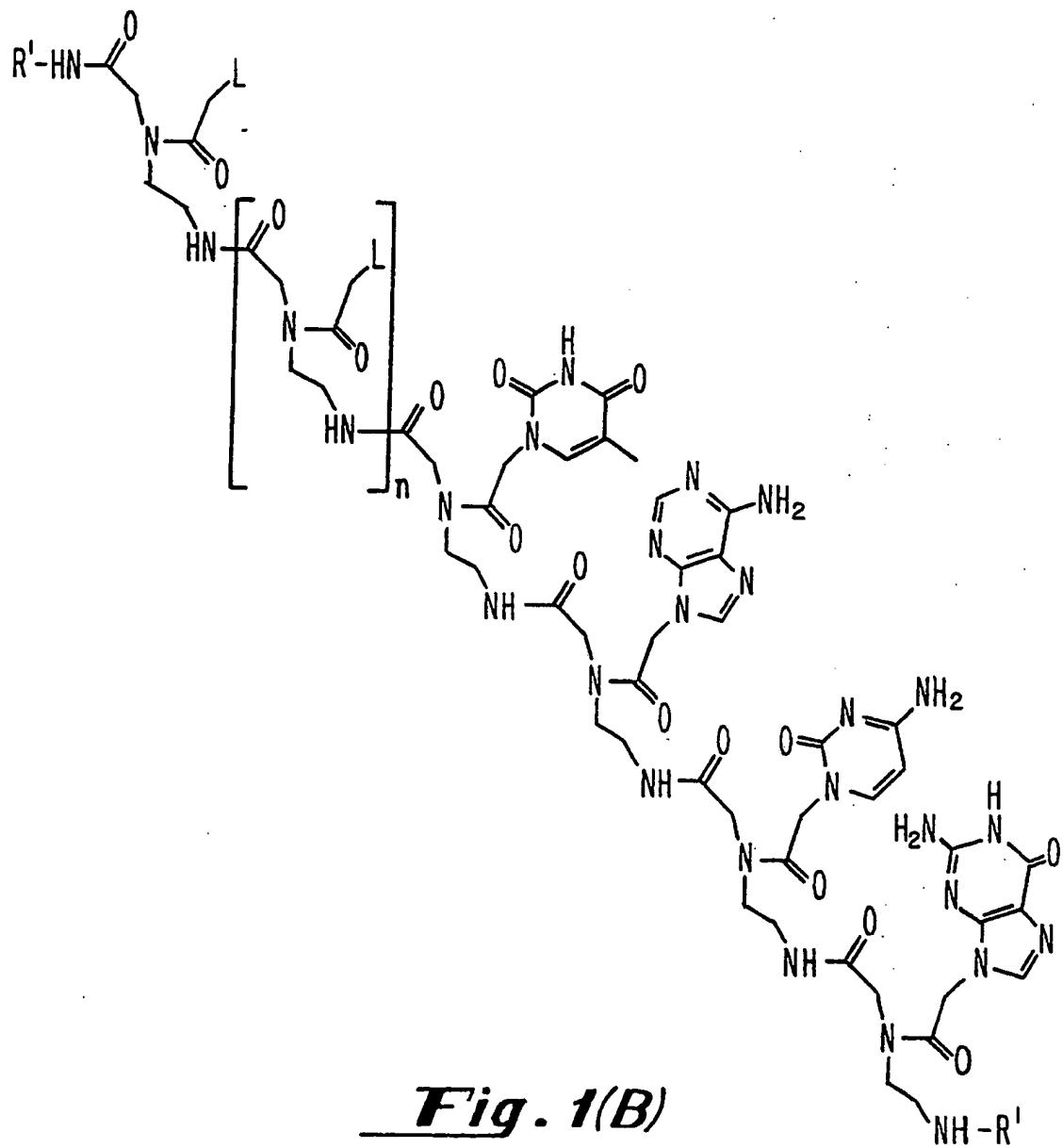
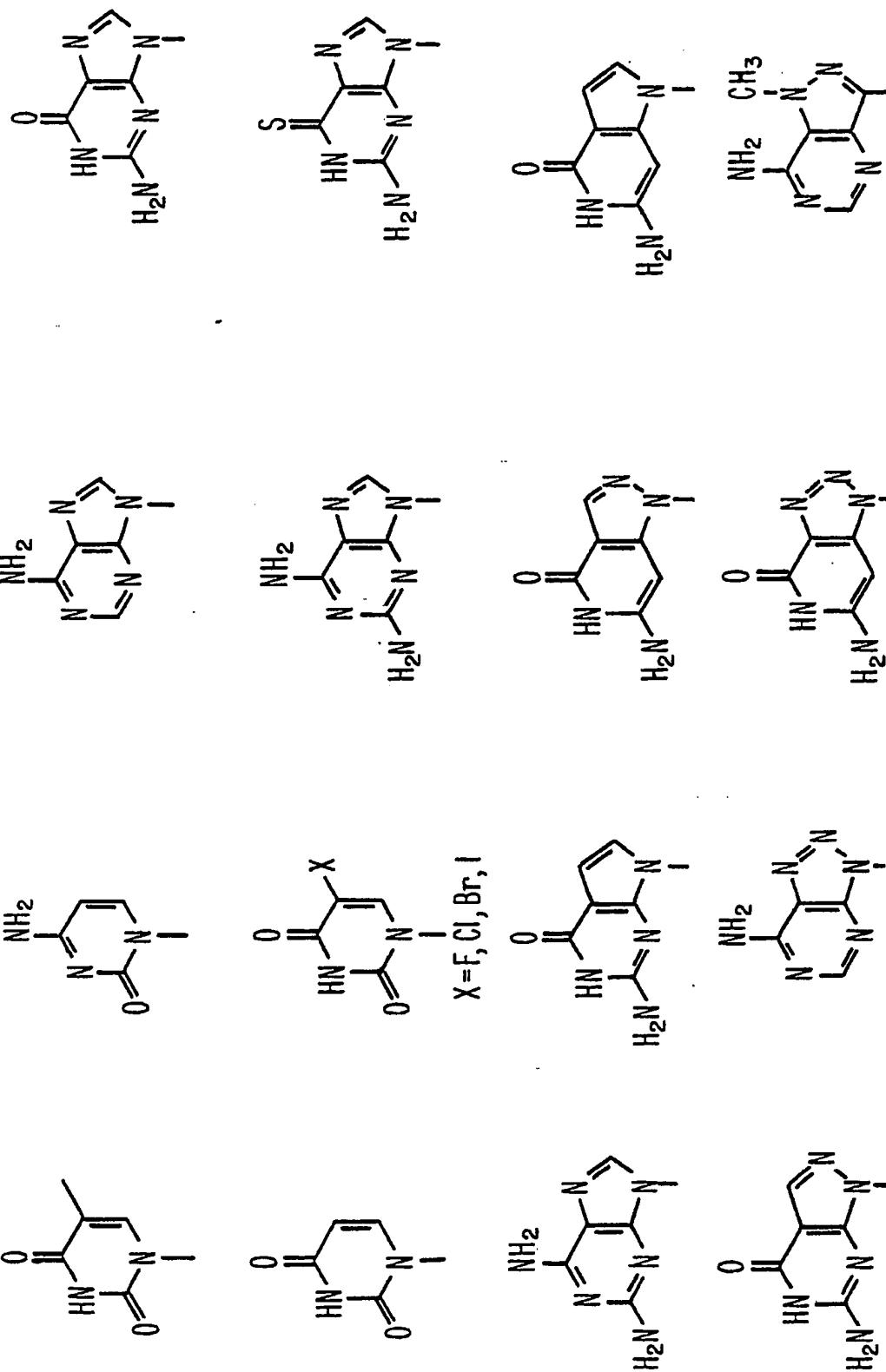
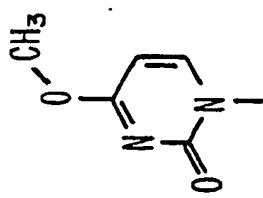
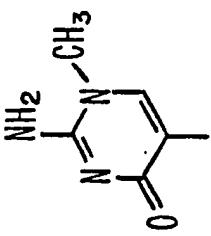
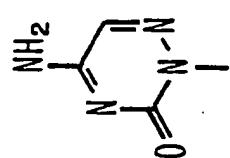
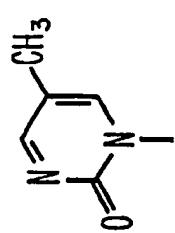
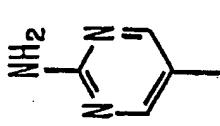
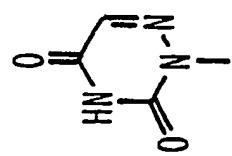
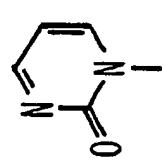
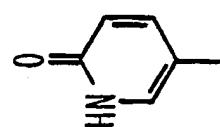
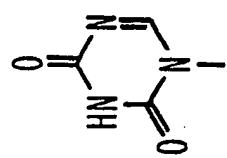
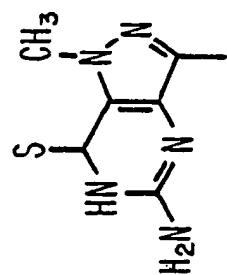
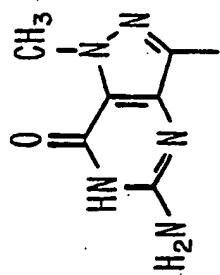
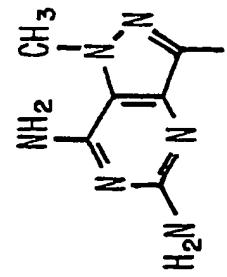


Fig. 1(B)

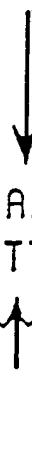
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***Fig. 2(A)***

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***Fig. 2(B)***

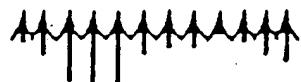
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300nm RADIATION
PHOTOCLEAVAGE

Fig. 3(a)

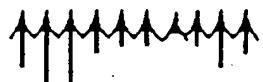
5'-GATCCAAAAAAAGGATC
3'-CTAGGGTTTTTTTCCTAG



DIAZOACRIDINE
PHOTOFOOTPRINT

Fig. 3(b)

5'-GATCCAAAAAAAGGATC
3'-CTAGGGTTTTTTTCCTAG



KMnO₄ CLEAVAGE

Fig. 3(c)

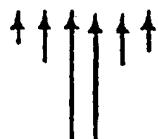
5'-GATCCAAAAAAAGGATC
3'-CTAGGGTTTTTTTCCTAG



S₁- NUCLEASE
CLEAVAGE

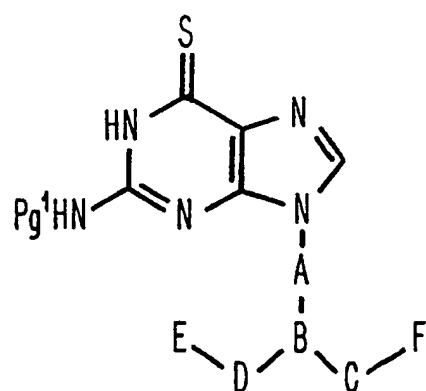
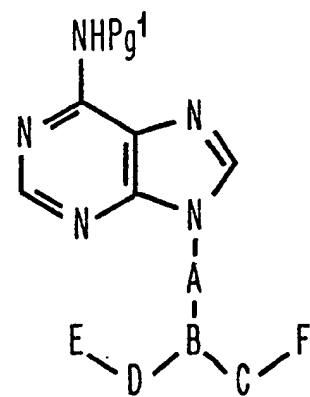
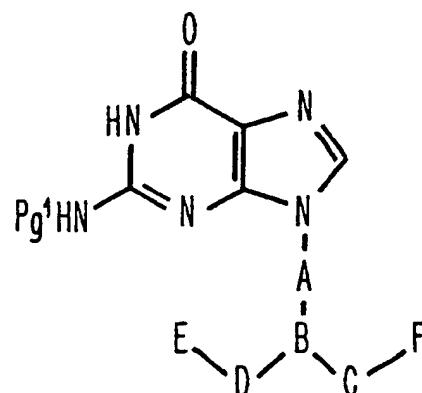
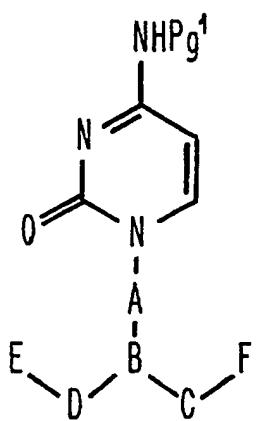
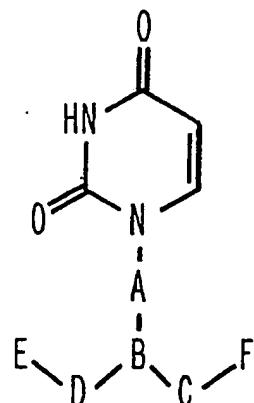
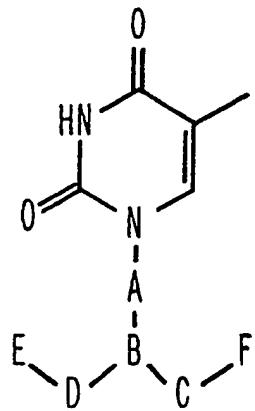
Fig. 3(d)

5'-GATCCAAAAAAAGGATC
3'-CTAGGGTTTTTTTCCTAG



MICROCOCCUS NUCLEASE
ENHANCEMENT

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***Fig. 4***

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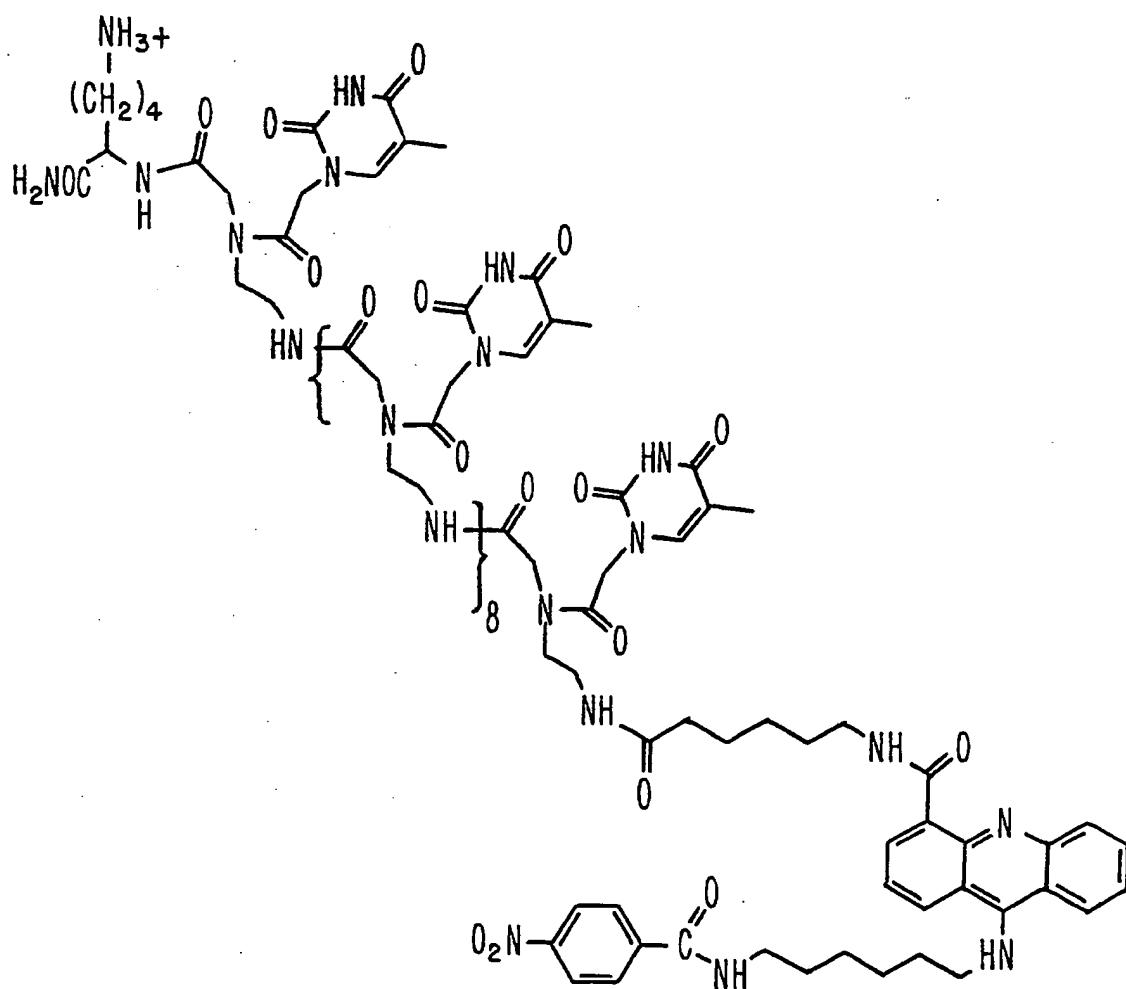
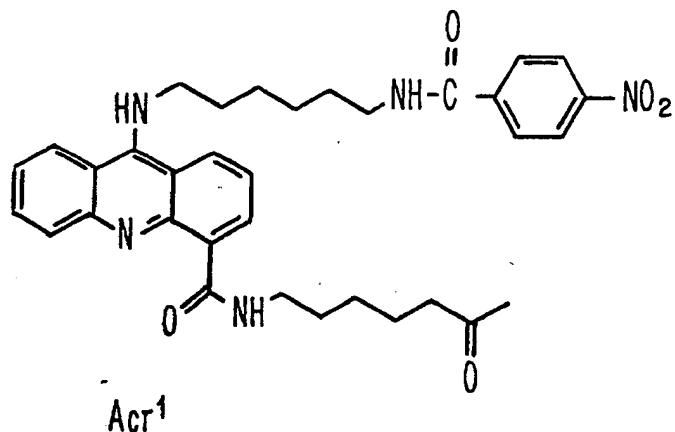
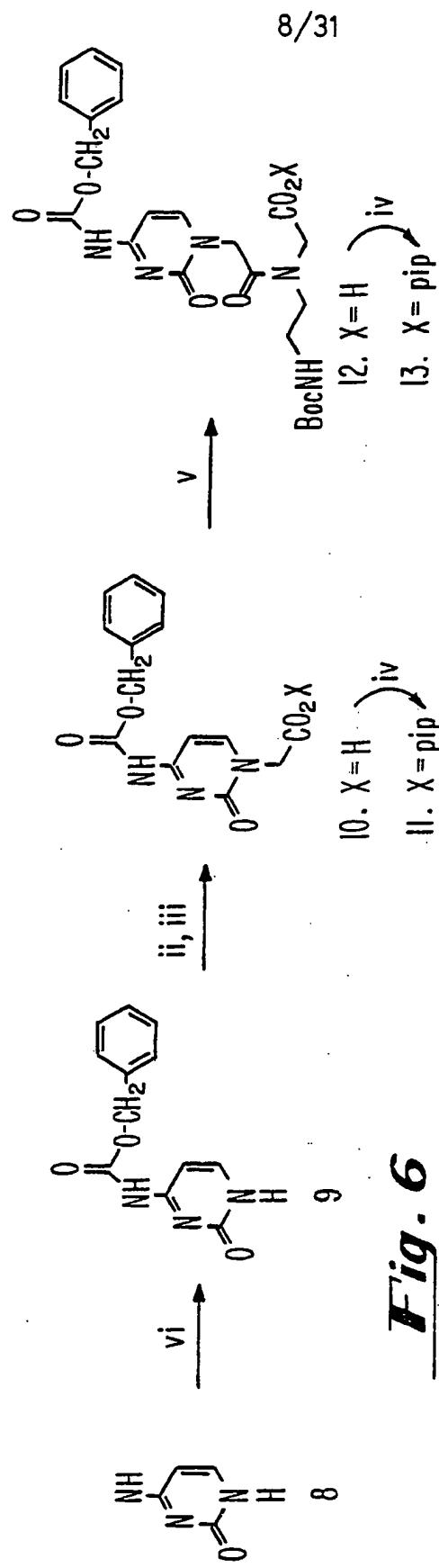
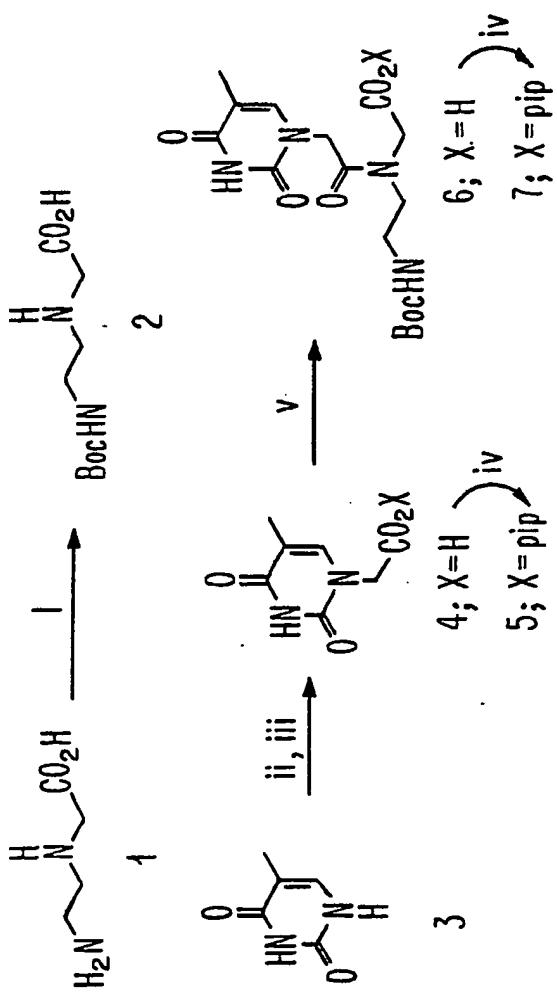
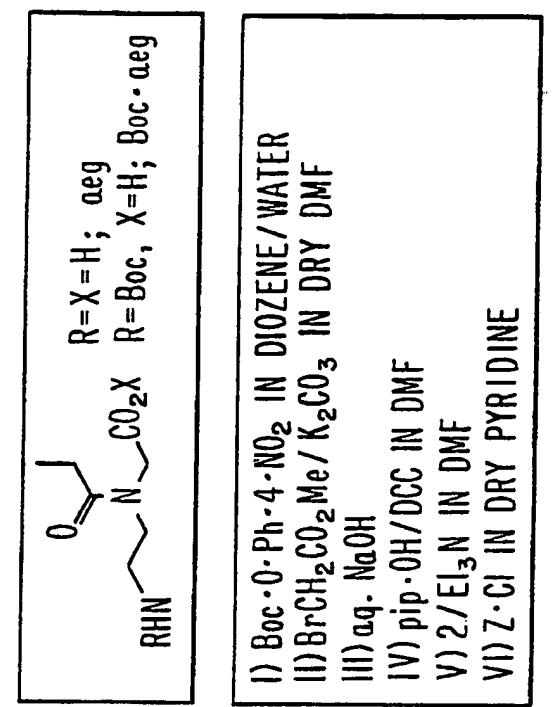
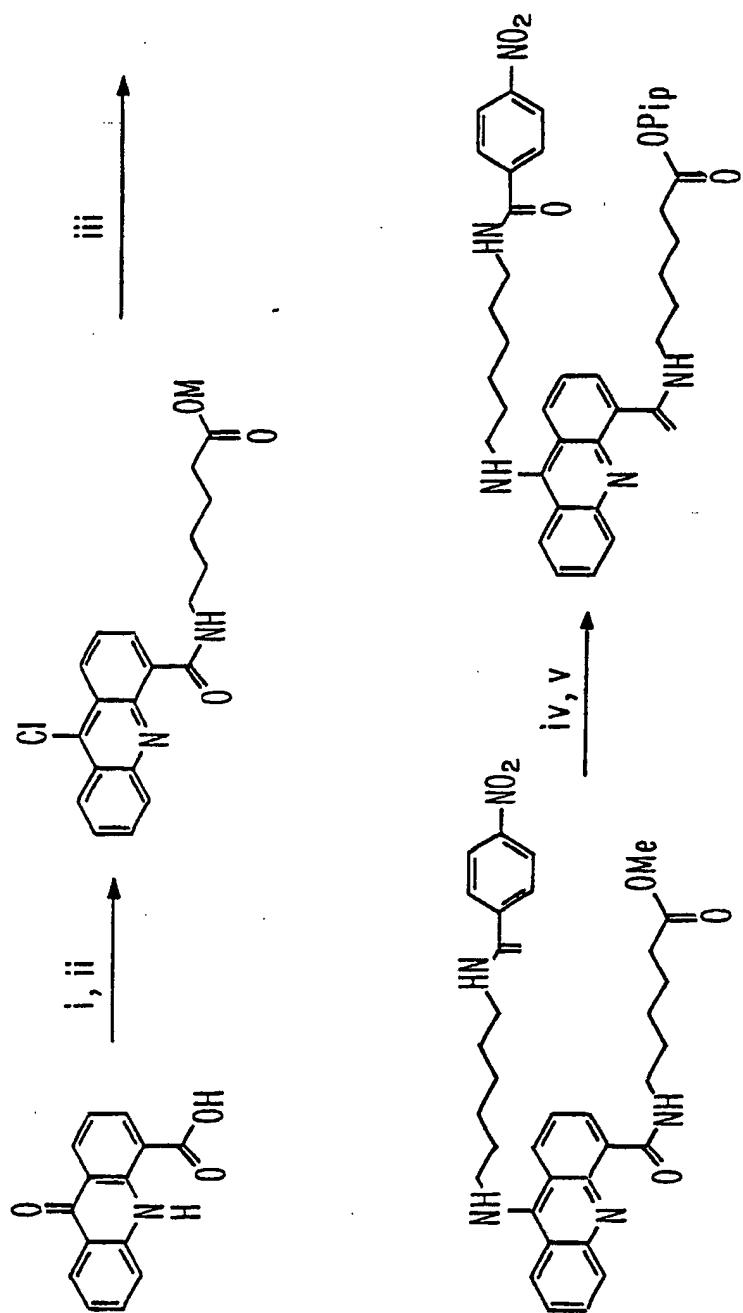


Fig. 5

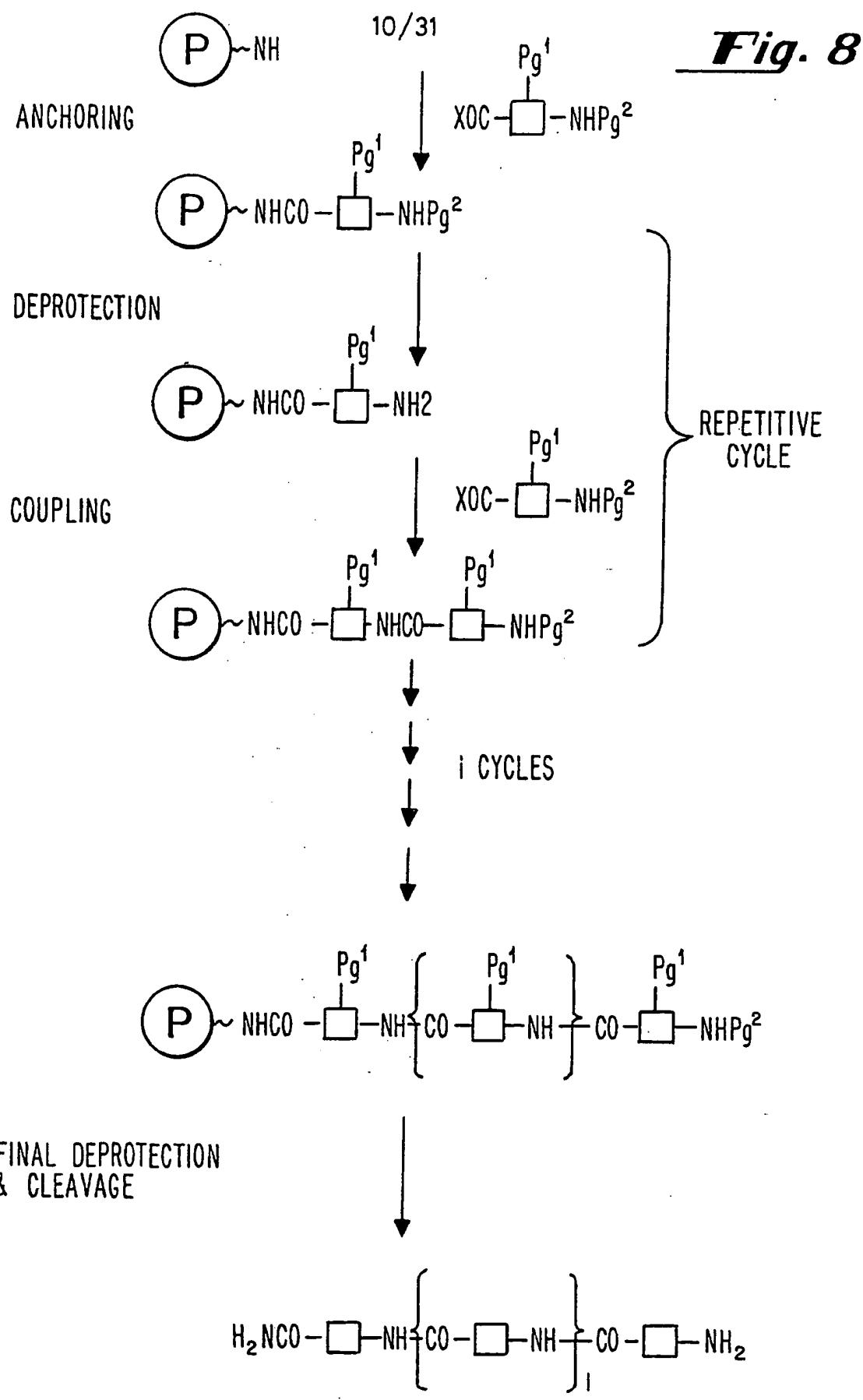
***Fig. 6***

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- I) SOCl_2/DMF (cat.) REFLUX
- II) $^+\text{H}_2\text{N}(\text{CH}_2)_5\text{CO}_2\text{CH}_3/\text{Et}_3\text{N}/\text{CH}_2\text{Cl}_2$ AT 0°C
- III) $\text{PhOH}/4\text{-NO}_2\text{Ph}-\text{CONH}(\text{CH}_2)_6\text{NH}_2$ AT 120°C
- IV) $\text{DMF}/\text{aq. NaOH}$
- V) $\text{DMF}/\text{CH}_2\text{Cl}_2/\text{PipOH/DCC}$

Fig. 7



SUBSTITUTE SHEET

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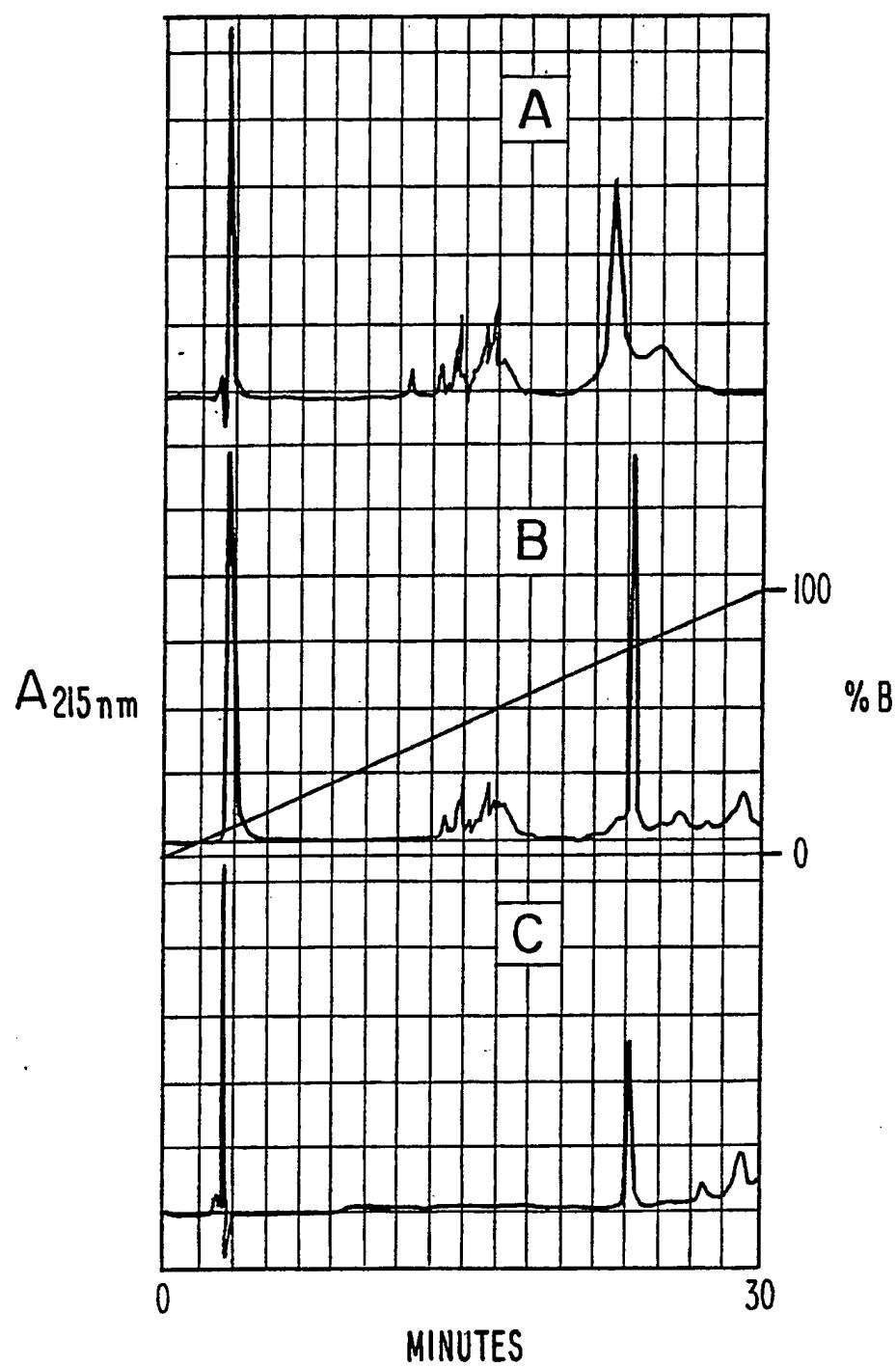


Fig. 9

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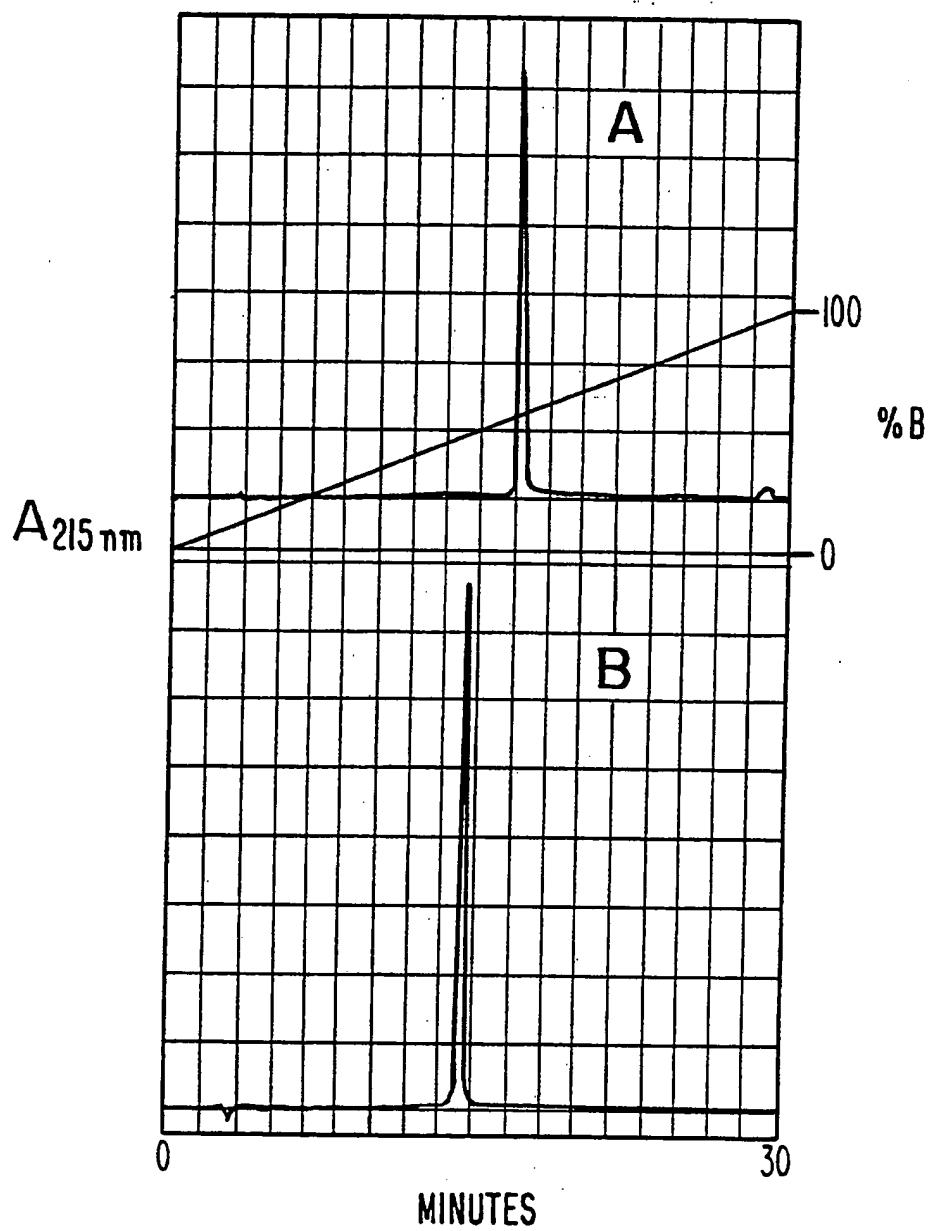


Fig. 10

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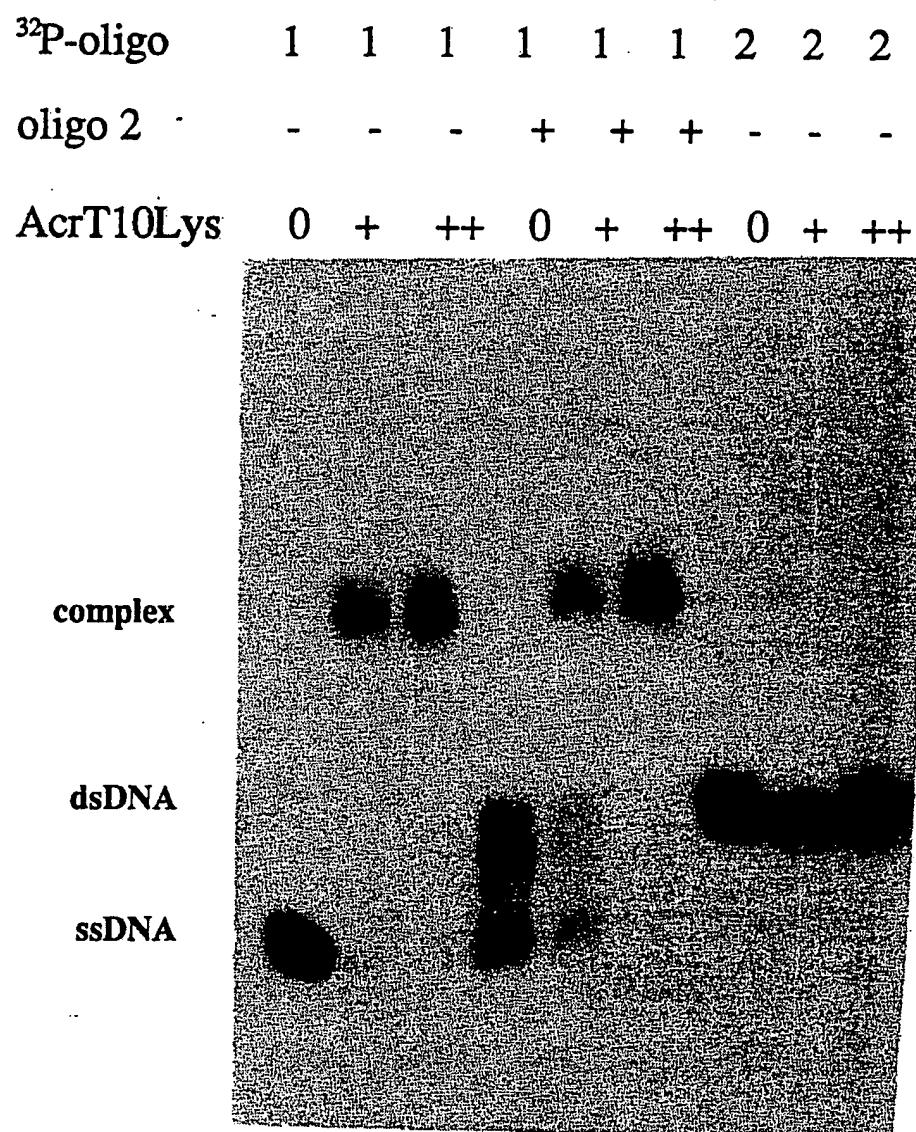


FIG. IIIA

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³² P-oligo	1	1	1	1	1	1	2	2	2
oligo 2	-	-	-	+	+	+	-	-	-
AcrT10Lys	0	+	++	0	+	++	0	+	++

complex

ssDNA

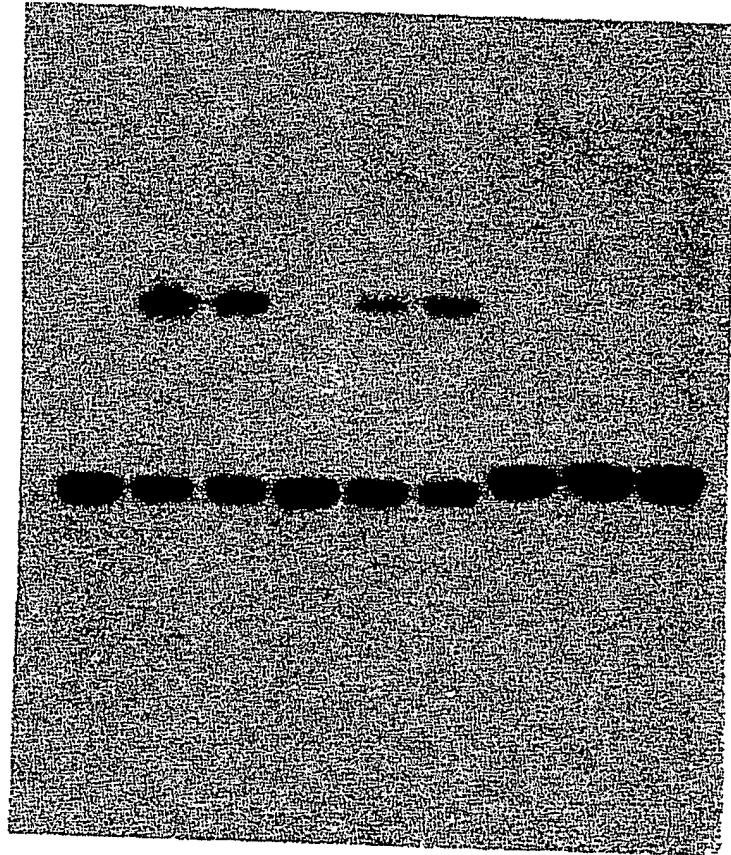


FIG. II B

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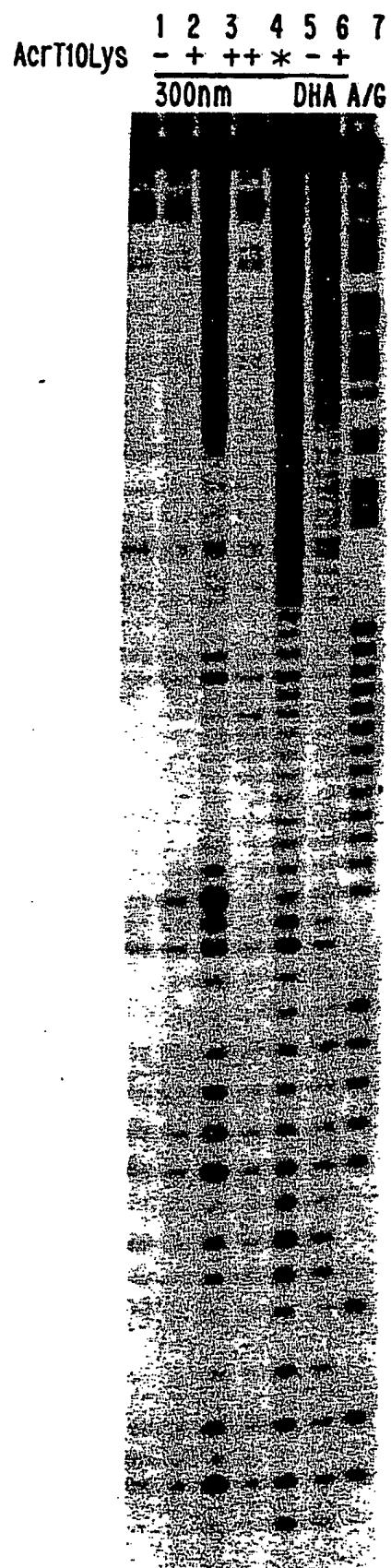


FIG. 12A

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AcrT1OLys 1 2 3 4 5 6 7 8 9 10 11
- + ++ - + ++ - - + +
300nm KMnO₄ c staph A/G

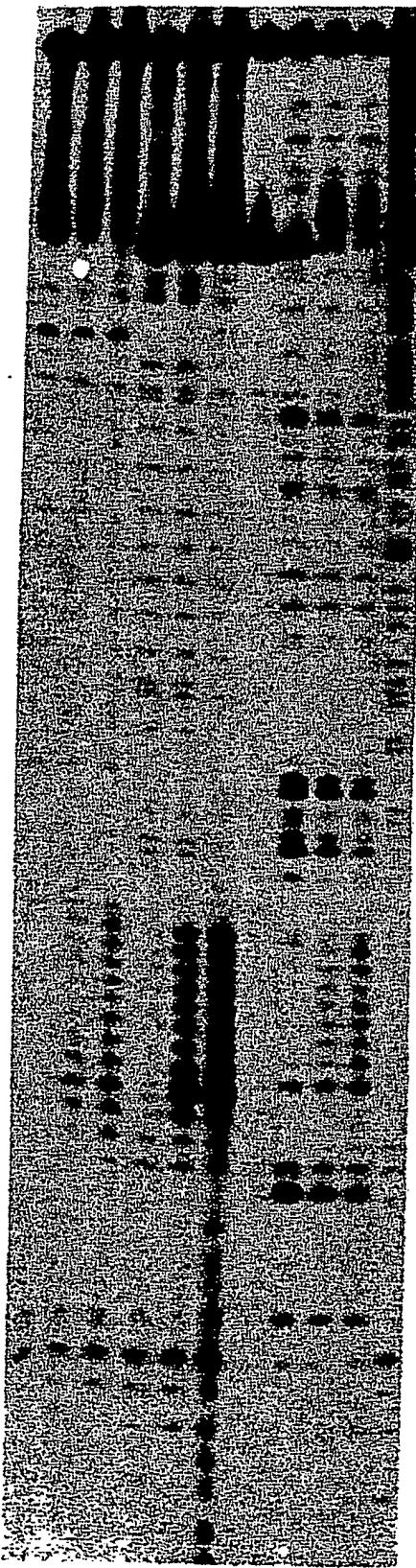


FIG. 12B

T₁₀

S₁-nuclease 0.1 1 10 0.1 1 10

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AcrT10Lys - - - + + +

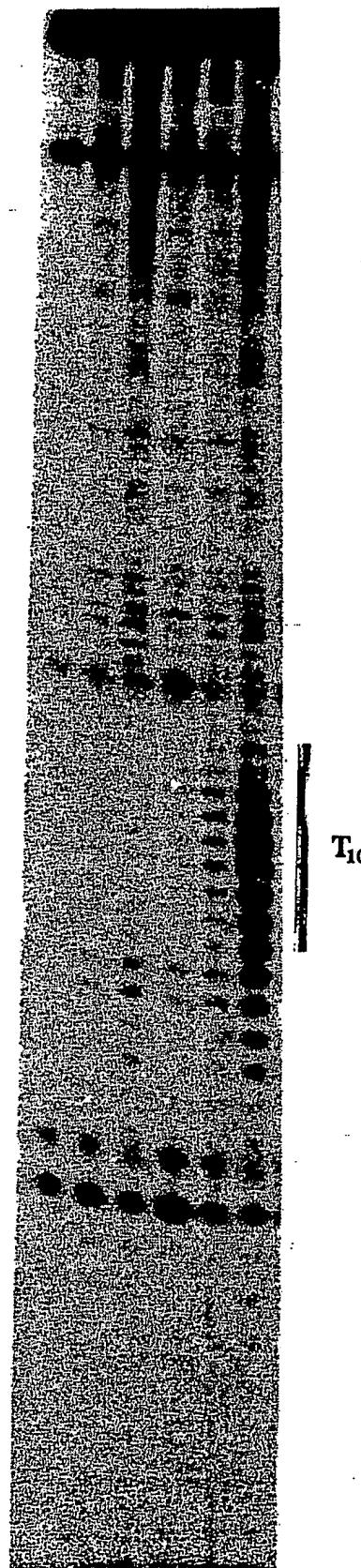
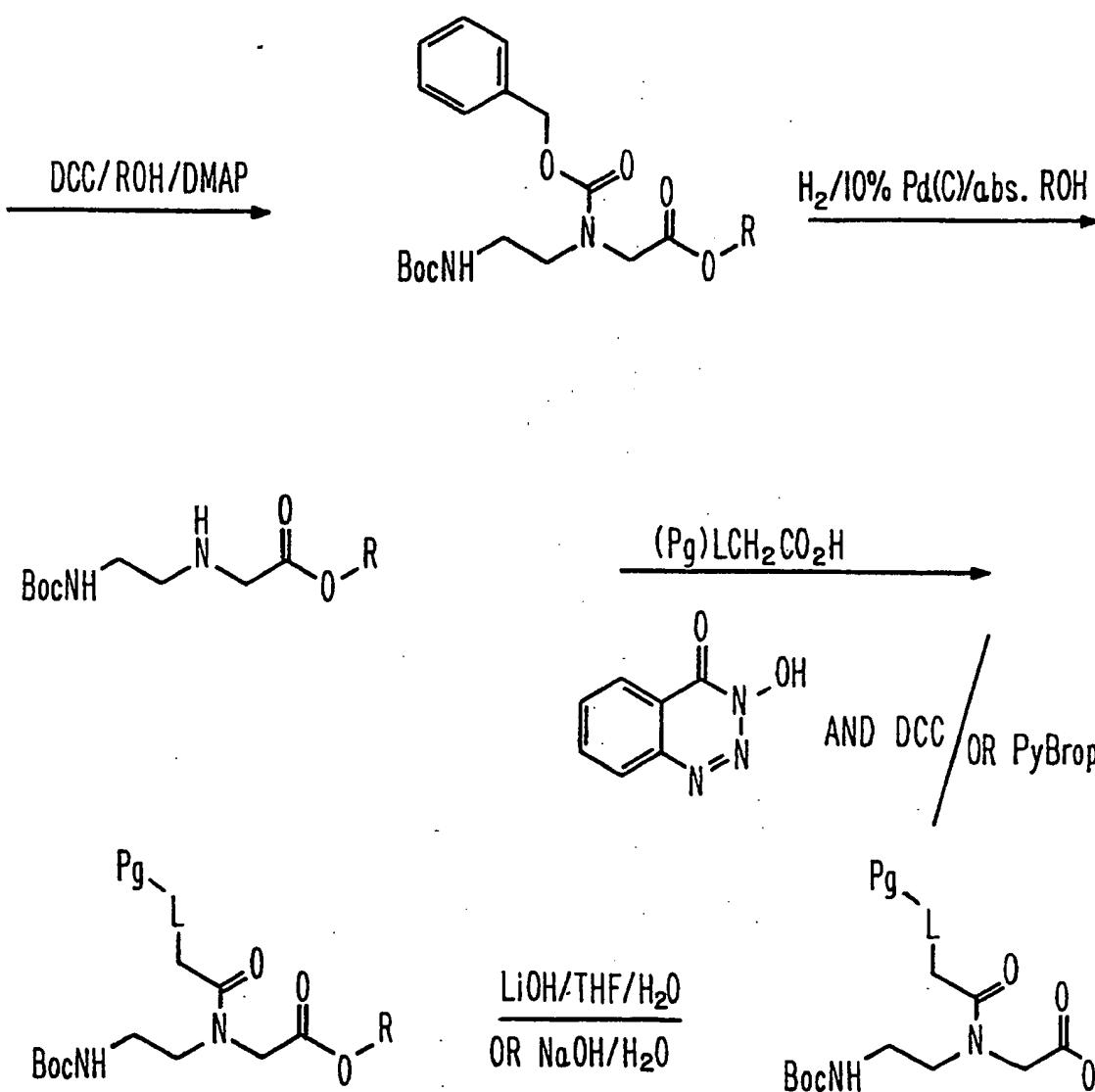
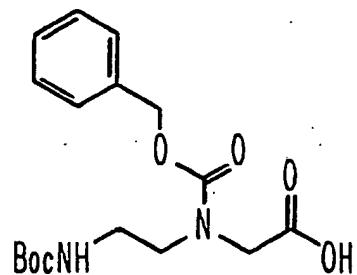
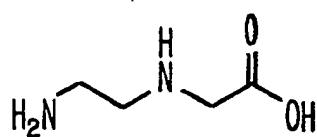
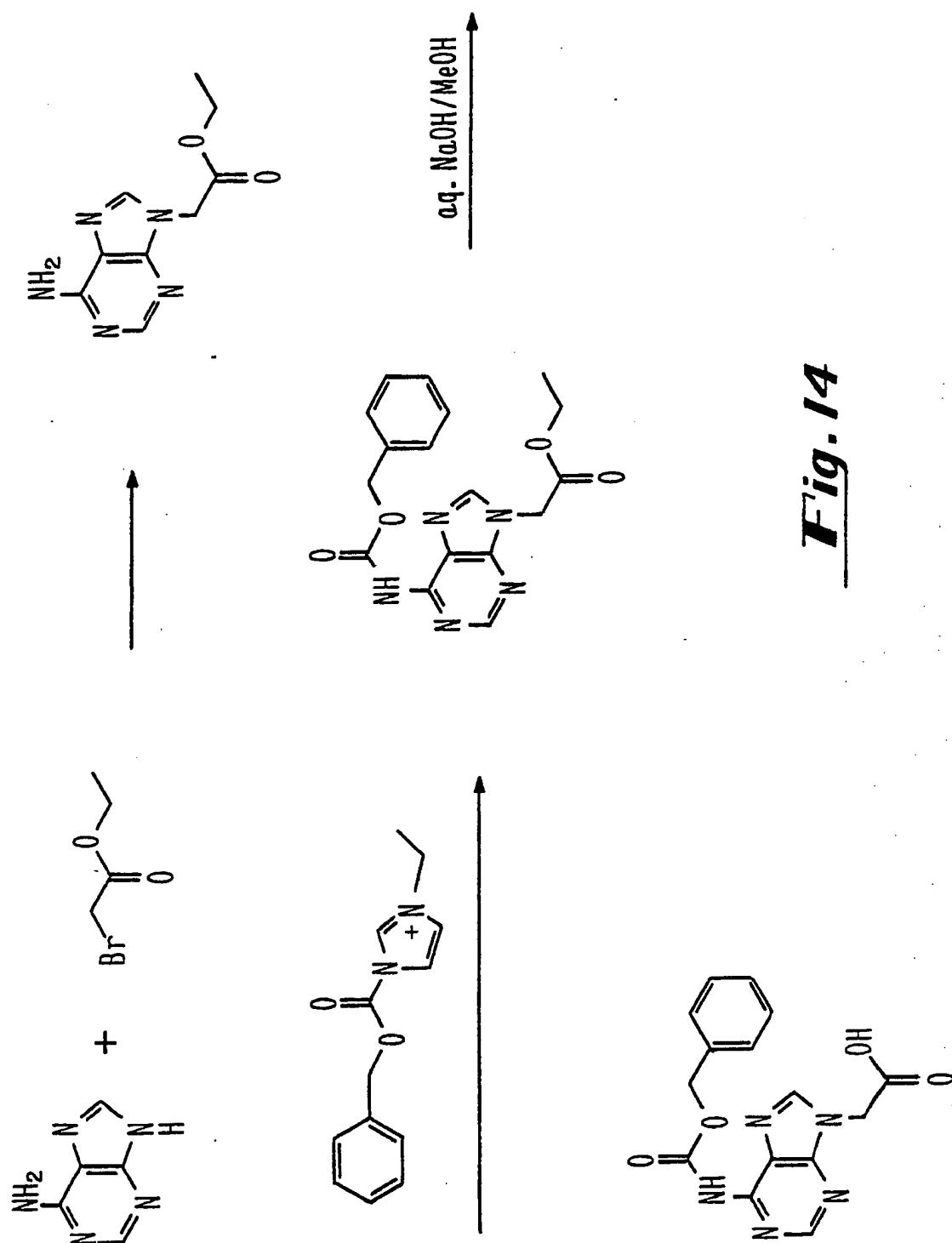


FIG. 12C

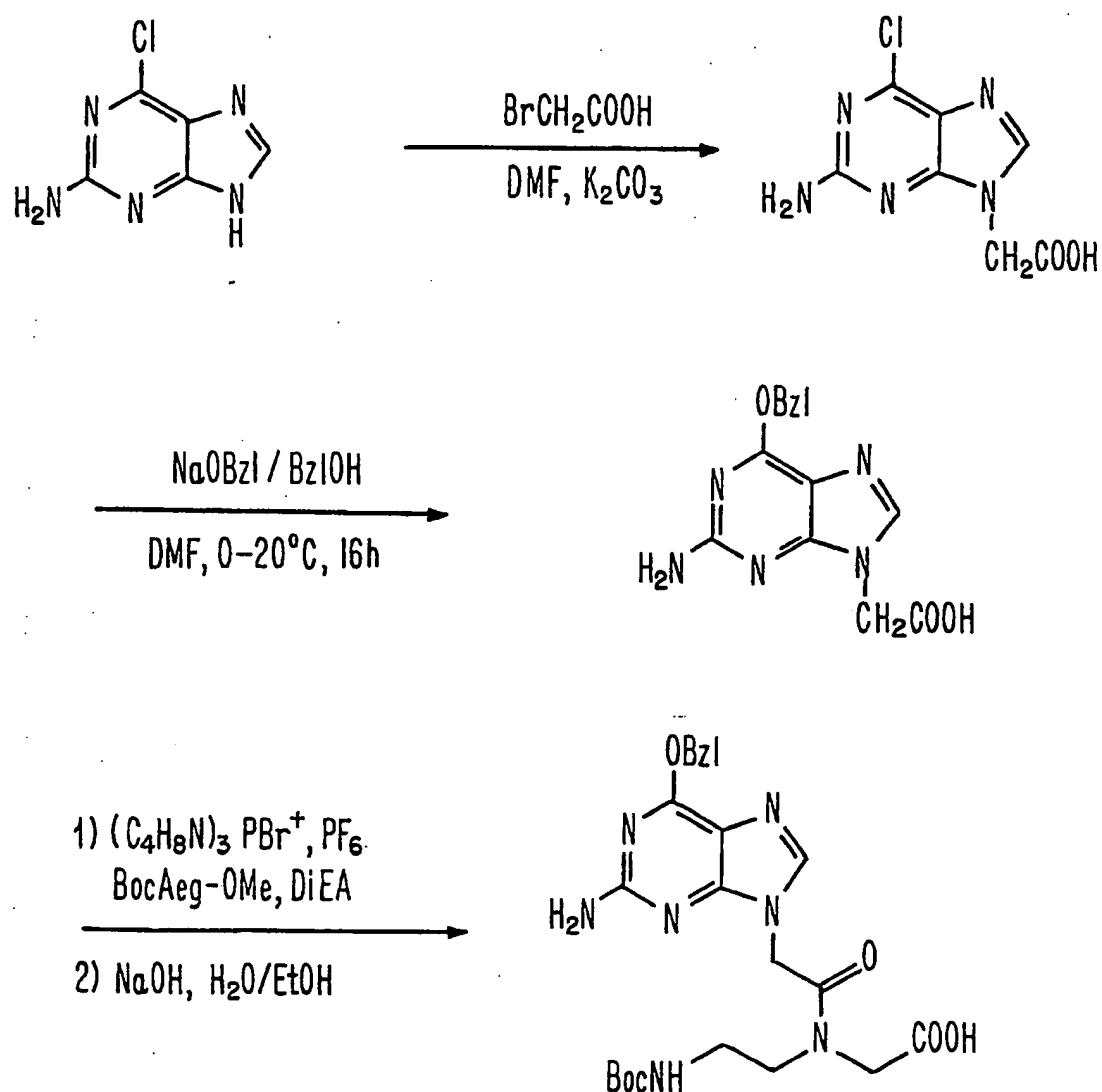
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***Fig. 13***

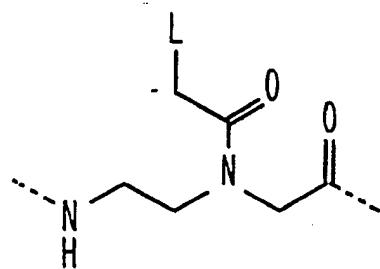
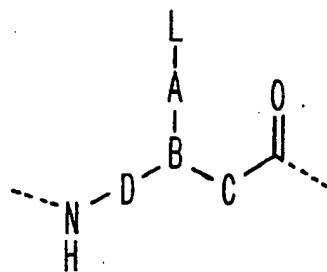
19 / 31



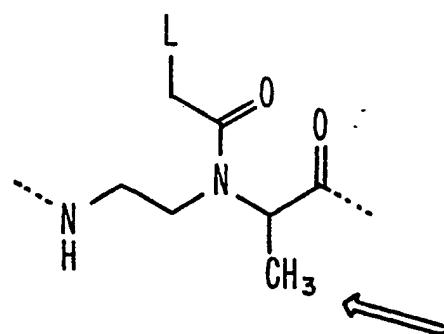
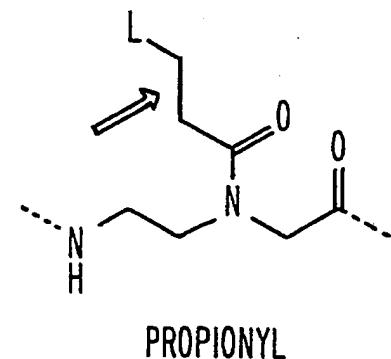
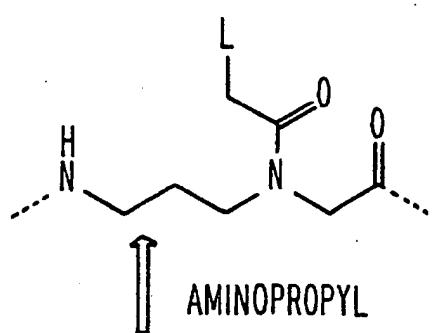
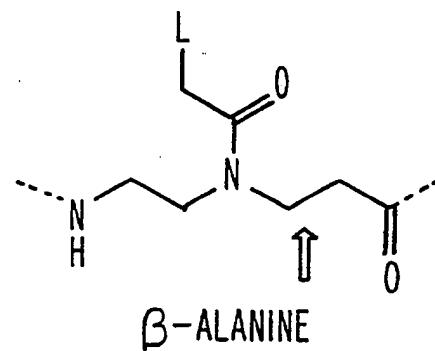
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Fig. 15

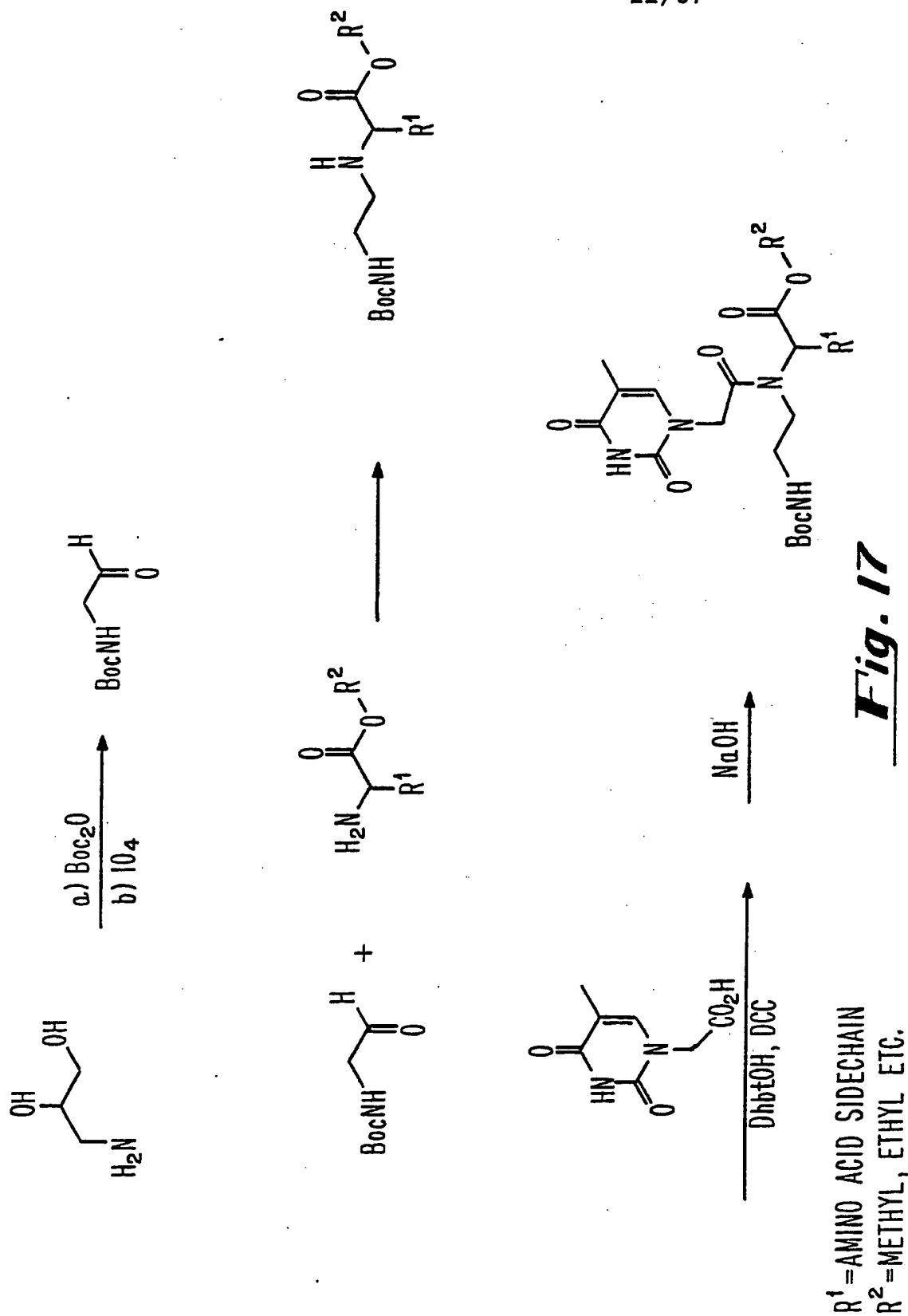
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AMINOETHYL GLYCINE

Fig. 16

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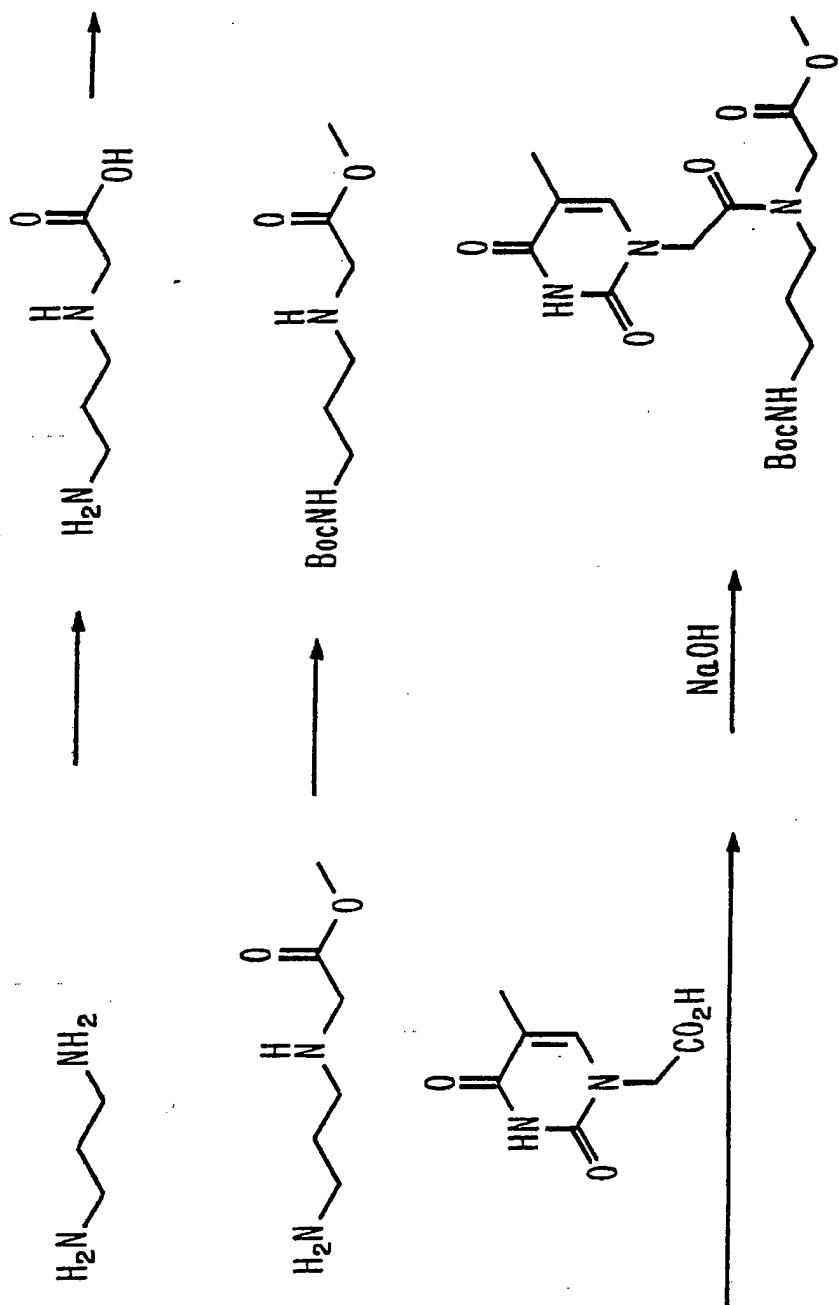
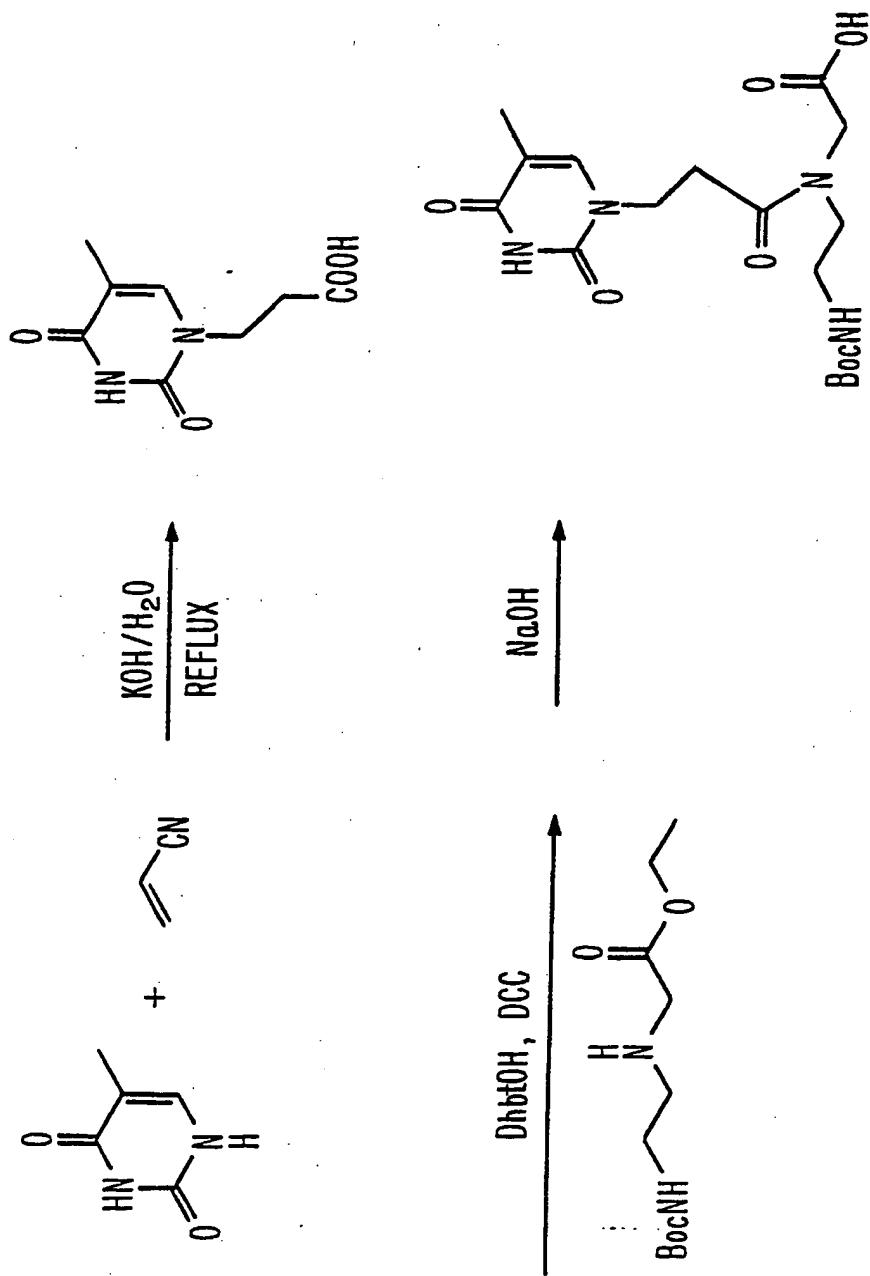
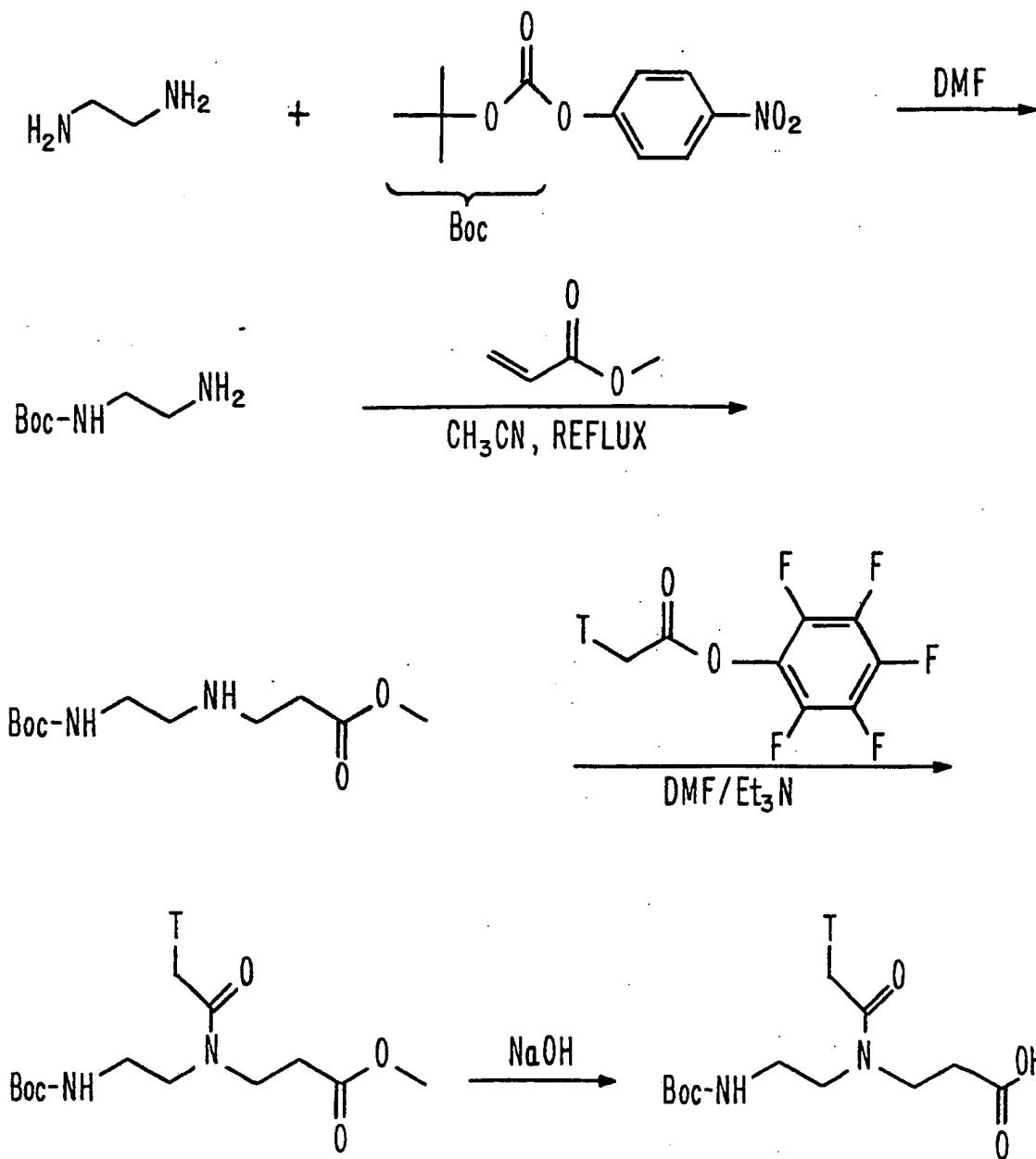


Fig. 18(a)

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**Fig. 19**

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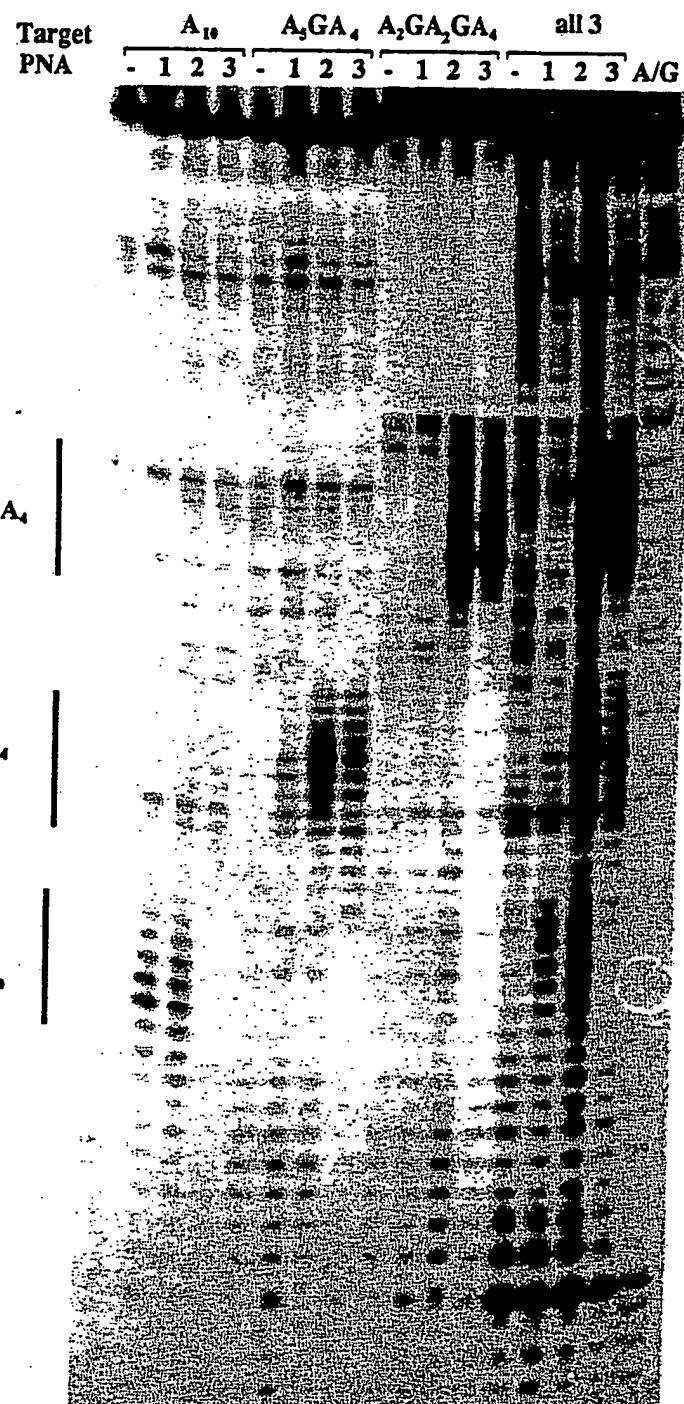
PNA 1: T_{10} PNA 2: T_5CT_4 PNA 3: $T_2CT_2CT_4$

FIG. 20

SUBSTITUTE SHEET

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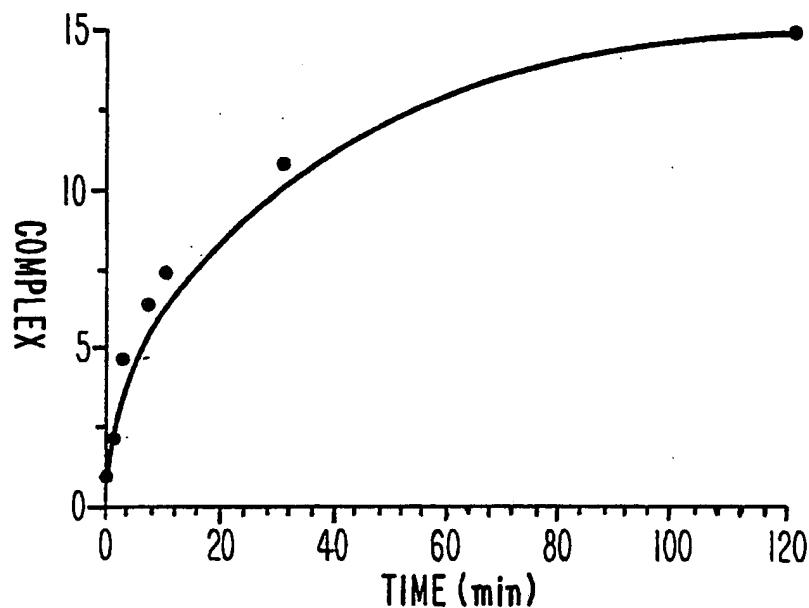


Fig. 21

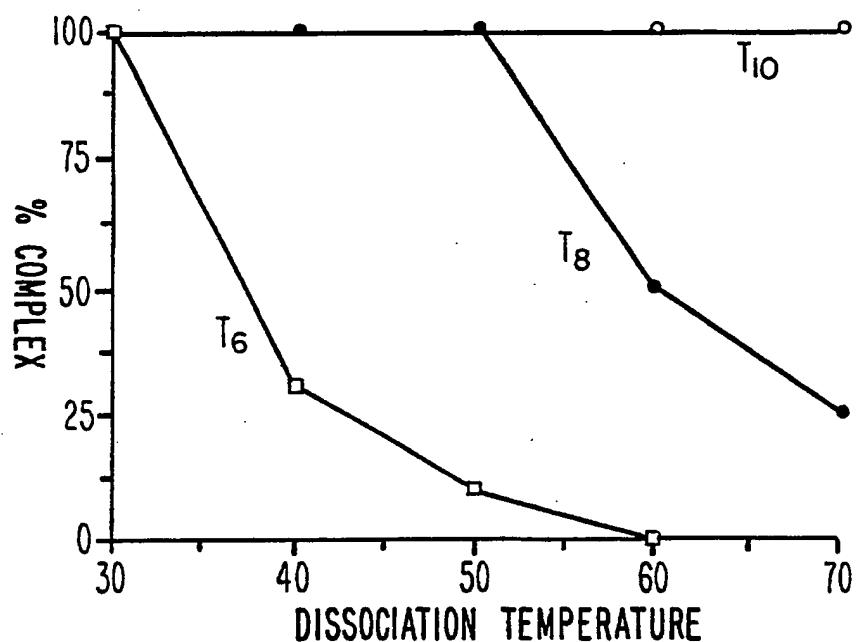
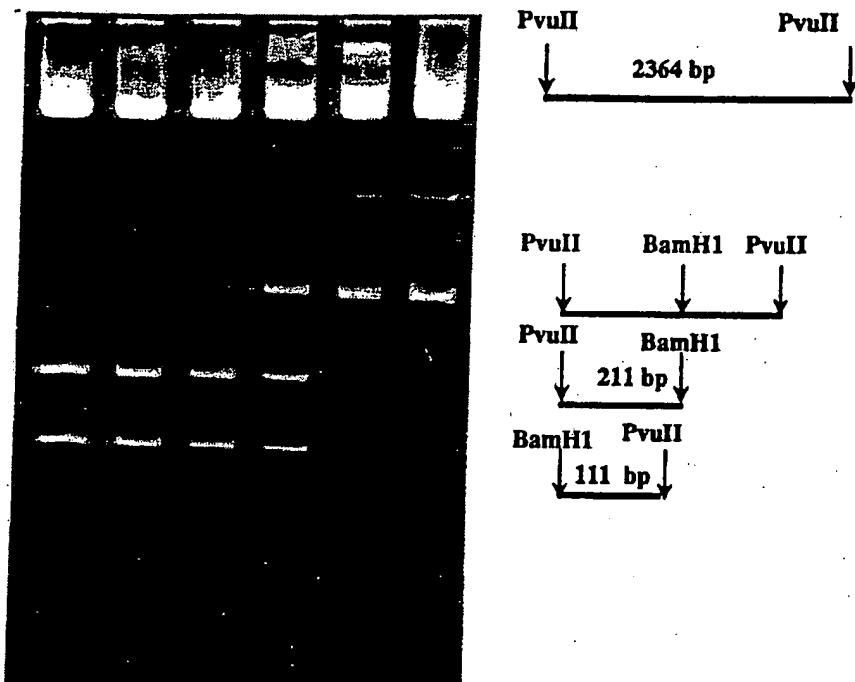


Fig. 22

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PNA/DNA 0 0.006 0.02 0.06 0.2 0.6



PNA Target

5' -----GGATCCAAAAAAAAGGATCC-----
3' -----CCTAGGTTTTTTTCCTAGG-----

BamH1

BamH1

FIG. 23

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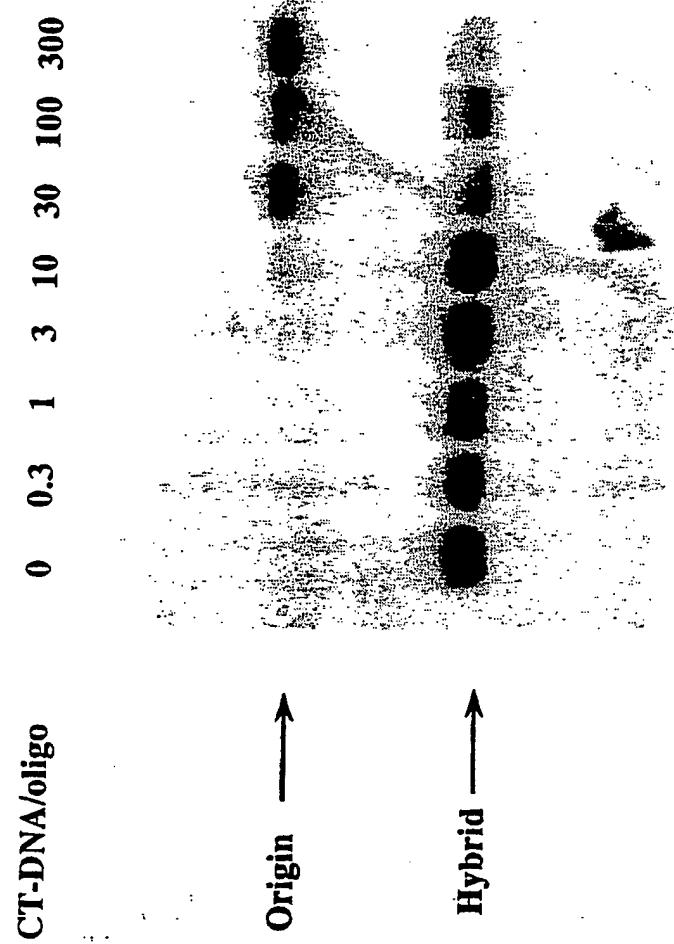
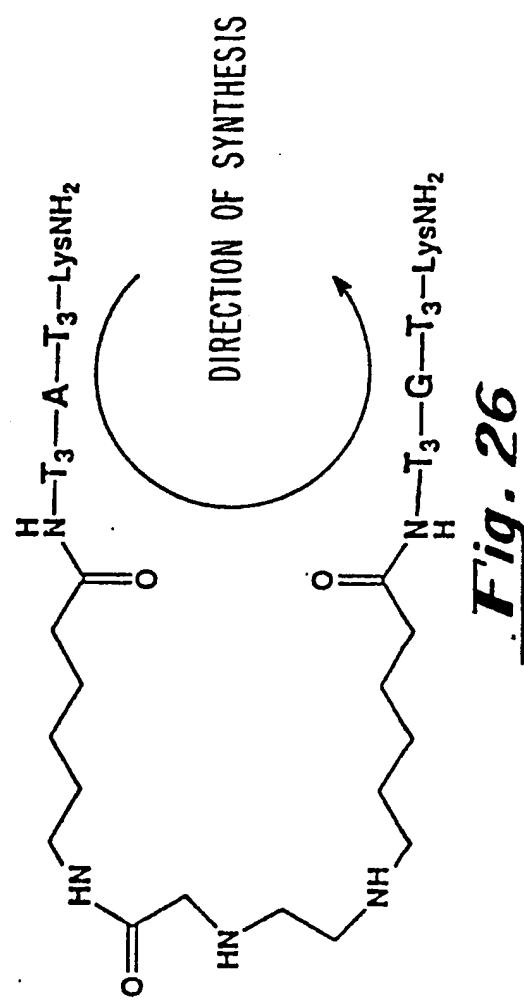
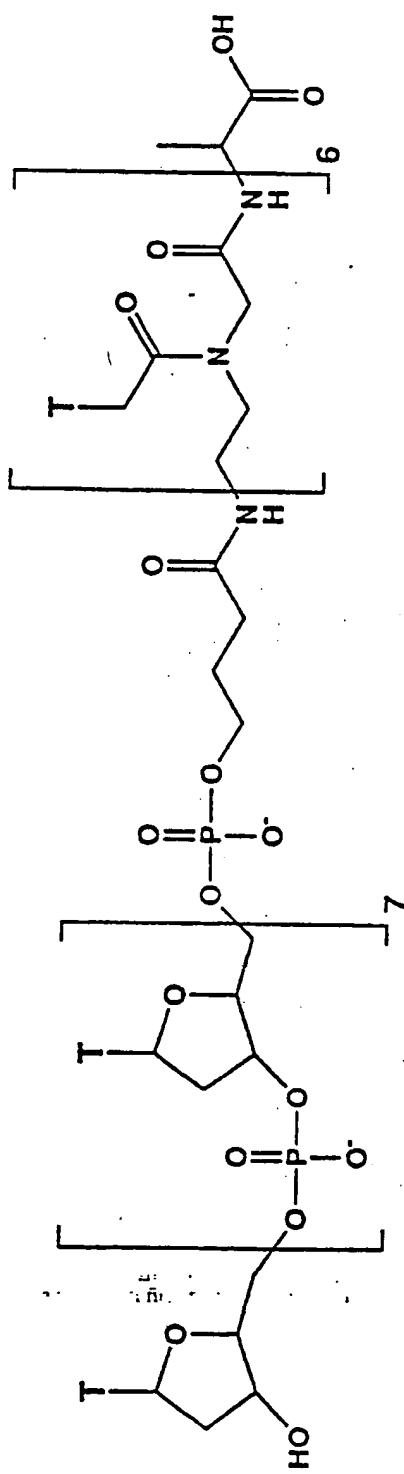


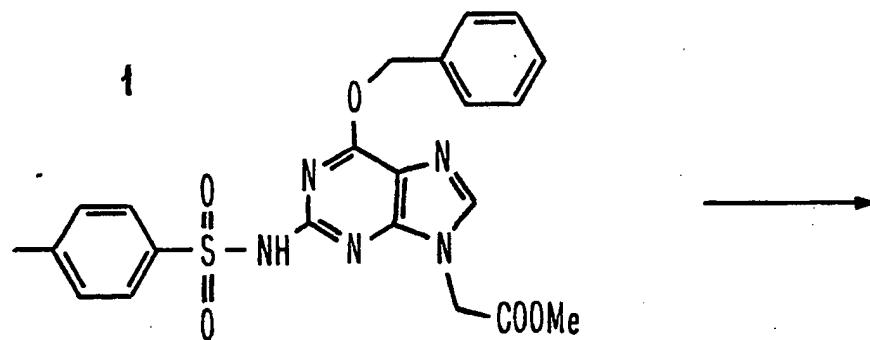
FIG. 24

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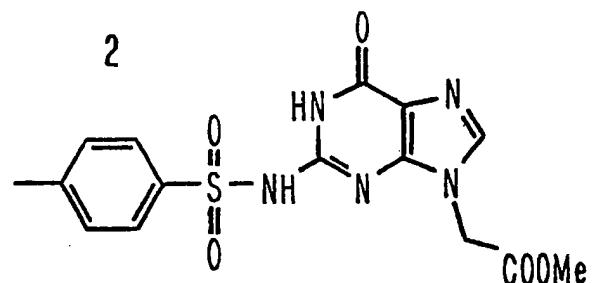


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COMPOUND

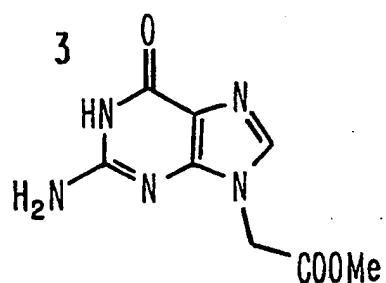


COMPOUND 1 IN
50% TFA: 50% METHYLENE CHLORIDE, 5 h, rt.



QUANTITATIVE DE-BENZYLATION

COMPOUND 1 IN
100% HF, 0°C, 1 h

QUANTITATIVE DE-BENZYLATION
AND DE-SULFONYLATION**Fig. 27**

INTERNATIONAL SEARCH REPORT

International Application No. PCT/EP 92/01219

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ⁶							
According to International Patent Classification (IPC) or to both National Classification and IPC							
Int.C1.5		C 07 K 5/00	C 07 K 7/00	C 12 Q 1/68			
C 08 L 77/00							
II. FIELDS SEARCHED							
Minimum Documentation Searched ⁷							
Classification System		Classification Symbols					
Int.C1.5		C 07 K 5/00	C 07 K 7/00	C 12 Q 1/00			
		C 08 L 77/00					
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁸							
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹							
Category ¹⁰		Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²				Relevant to Claim No. ¹³	
1	X	WO,A,8605518 (J. SUMMERTON) 25 September 1986 (cited in the application) ---					
1	Y	WO,A,8605519 (J. SUMMERTON) 25 September 1986 ---					
5	P,X	Journal of the American Chemical Society, vol. 114, no. 5, 26 February 1992, American Chemical Society, M. EGHOLM et al.: "Peptide nucleic acids (PNA). Oligonucleotide analogues with an achiral peptide backbone", pages 1895-1897 ---					
5	P,X	Science, vol. 254, 6 December 1991, P.E. NIELSEN et al.: "Sequence-selective recognition of DNA by strand displacement with a thymine-substituted polyamide", pages 1497-1500 ---				-/-	
<p>¹⁰ Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed</p> <p>¹¹ T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family</p>							
IV. CERTIFICATION							
Date of the Actual Completion of the International Search				Date of Mailing of this International Search Report			
10-09-1992				07.10.92			
International Searching Authority EUROPEAN PATENT OFFICE				Signature of Authorized Officer <i>[Handwritten Signature]</i>			

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
A	Journal of The American Chemical Society, vol. 110, no. 22, 26 October 1988, American Chemical Society, D.P. MACK et al.: "Design and chemical synthesis of a sequence-specific DNA-cleaving protein", pages 7572-7574 ---	
A	Journal of Medicinal Chemistry, vol. 33, no. 7, 1990, American Chemical Society, L.P.G. WAKELIN et al.: "Kinetic and equilibrium binding studies of amsacrine-4-carboxamides: a class of asymmetrical DNA-intercalating agents which bind by threading through the DNA helix", pages 2039-2044 ---	
A	BIOCHEMISTRY, vol. 27, no. 17, 23 August 1988, American Chemical Society, P.E. NIELSEN et al.: "Photochemical cleavage of DNA by nitrobenzamides linked to 9-aminoacridine", pages 6338-6343 -----	

INTERNATIONAL SEARCH REPORT

International application No.

PCT/EP 92/01219

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 23-31 are directed to a method of treatment of the human/animal body the search has been carried out and based on the alleged effects of the compounds.
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest.
- No protest accompanied the payment of additional search fees.

ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO.

EP 9201219

SA 60822

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report.
The members are as contained in the European Patent Office EDP file on 24/09/92.
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Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A- 8605518	25-09-86	AU-A-	5661386	13-10-86
		AU-A-	5698186	13-10-86
		CA-A-	1268404	01-05-90
		EP-A-	0216860	08-04-87
		EP-A-	0215942	01-04-87
		JP-T-	62502338	10-09-87
		JP-T-	62502357	10-09-87
		WO-A-	8605519	25-09-86
		US-A-	5034506	23-07-91
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WO-A- 8605519	25-09-86	AU-A-	5661386	13-10-86
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		CA-A-	1268404	01-05-90
		EP-A-	0216860	08-04-87
		EP-A-	0215942	01-04-87
		JP-T-	62502338	10-09-87
		JP-T-	62502357	10-09-87
		WO-A-	8605518	25-09-86
		US-A-	5034506	23-07-91
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PCT

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5 : C07K 5/00, 7/00, C12Q 1/68 C08L 77/00		A1	(11) International Publication Number: WO 92/20702 (43) International Publication Date: 26 November 1992 (26.11.92)									
(21) International Application Number: PCT/EP92/01219			(74) Agent: HALLYBONE, Huw, George; Carpmaels & Ransford, 43 Bloomsbury Square, London WC1A 2RA (GB).									
(22) International Filing Date: 22 May 1992 (22.05.92)												
(30) Priority data: <table><tr><td>0986/91</td><td>24 May 1991 (24.05.91)</td><td>DK</td></tr><tr><td>0987/91</td><td>24 May 1991 (24.05.91)</td><td>DK</td></tr><tr><td>0510/92</td><td>15 April 1992 (15.04.92)</td><td>DK</td></tr></table>		0986/91	24 May 1991 (24.05.91)	DK	0987/91	24 May 1991 (24.05.91)	DK	0510/92	15 April 1992 (15.04.92)	DK		(81) Designated States: AT, AT (European patent), AU, BB, BE (European patent), BF (OAPI patent), BG, BJ (OAPI patent), BR, CA, CF (OAPI patent), CG (OAPI patent), CH, CH (European patent), CI (OAPI patent), CM (OAPI patent), CS, DE, DE (European patent), DK, DK (European patent), ES, ES (European patent), FI, FR (European patent), GA (OAPI patent), GB, GB (European patent), GN (OAPI patent), GR (European patent), HU, IT (European patent), JP, KP, KR, LK, LU, LU (European patent), MC (European patent), MG, ML (OAPI patent), MN, MR (OAPI patent), MW, NL, NL (European patent), NO, PL, RO, RU, SD, SE, SE (European patent), SN (OAPI patent), TD (OAPI patent), TG (OAPI patent), US.
0986/91	24 May 1991 (24.05.91)	DK										
0987/91	24 May 1991 (24.05.91)	DK										
0510/92	15 April 1992 (15.04.92)	DK										
(71)(72) Applicants and Inventors: BUCHARDT, Ole [DK/DK]; Søndergårdsvæj 73, DK-3500 Værløse (DK). EGHOLM, Michael [DK/DK]; Sindshvilevej 5, 3. tv., DK-2000 Frederiksberg (DK). NIELSEN, Peter, Eigil [DK/DK]; Hjortevænget 509, DK-2980 Kokkedal (DK). BERG, Rolf, Henrik [DK/DK]; Langelandsvej 20 B, 3. tv, DK-2000 Frederiksberg (DK).			Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>									

(54) Title: PEPTIDE NUCLEIC ACIDS

(57) Abstract

A novel class of compounds, known as peptide nucleic acids, bind complementary ssDNA and RNA strands more strongly than a corresponding DNA. The peptide nucleic acids generally comprise ligands such as naturally occurring DNA bases attached to a peptide backbone through a suitable linker.

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CA	Canada	IT	Italy	RU	Russian Federation
CF	Central African Republic	JP	Japan	SD	Sudan
CG	Congo	KP	Democratic People's Republic of Korea	SE	Sweden
CH	Switzerland	KR	Republic of Korea	SN	Senegal
CI	Côte d'Ivoire	LI	Liechtenstein	SU	Soviet Union
CM	Cameroon	LK	Sri Lanka	TD	Chad
CS	Czechoslovakia	LU	Luxembourg	TG	Togo
DE	Germany	MC	Monaco	US	United States of America
DK	Denmark	MG	Madagascar		
ES	Spain				

PEPTIDE NUCLEIC ACIDS

RELATED APPLICATIONS

This application is a continuation-in-part of the
5 following Danish Patent Applications: No. 986/91, filed May
24, 1991, No. 987/91, filed May 24, 1991, and No. 510/92,
filed April 15, 1992. The entire disclosure of each
application is incorporated herein by reference.

10 FIELD OF THE INVENTION

This invention is directed to compounds that are not
polynucleotides yet which bind to complementary DNA and RNA
strands more strongly than the corresponding DNA. In particular,
the invention concerns compounds wherein naturally-occurring
15 nucleobases or other nucleobase-binding moieties are
covalently bound to a polyamide backbone.

BACKGROUND OF THE INVENTION

Oligodeoxyribonucleotides as long as 100 base pairs
20 (bp) are routinely synthesized by solid phase methods using
commercially available, fully automatic synthesis machines.
The chemical synthesis of oligoribonucleotides, however, is
far less routine. Oligoribonucleotides also are much less
stable than oligodeoxyribonucleotides, a fact which has
25 contributed to the more prevalent use of oligodeoxyribonucleo-
tides in medical and biological research directed to, for
example, gene therapy or the regulation of transcription or
translation.

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The function of a gene starts by transcription of its information to a messenger RNA (mRNA) which, by interaction with the ribosomal complex, directs the synthesis of a protein coded for by its sequence. The synthetic process is known as 5 translation. Translation requires the presence of various co-factors and building blocks, the amino acids, and their transfer RNAs (tRNA), all of which are present in normal cells.

Transcription initiation requires specific recognition 10 of a promoter DNA sequence by the RNA-synthesizing enzyme, RNA polymerase. In many cases in prokaryotic cells, and probably in all cases in eukaryotic cells, this recognition is preceded by sequence-specific binding of a protein transcription factor to the promoter. Other proteins which bind to the promoter, 15 but whose binding prohibits action of RNA polymerase, are known as repressors. Thus, gene activation typically is regulated positively by transcription factors and negatively by repressors.

Most conventional drugs function by interaction with 20 and modulation of one or more targeted endogenous proteins, e.g., enzymes. Such drugs, however, typically are not specific for targeted proteins but interact with other proteins as well. Thus, a relatively large dose of drug must be used to effectively modulate a targeted protein. Typical 25 daily doses of drugs are from 10^{-5} - 10^{-1} millimoles per kilogram of body weight or 10^{-3} - 10 millimoles for a 100 kilogram person. If this modulation instead could be effected by interaction with and inactivation of mRNA, a dramatic reduction in the necessary amount of drug necessary could 30 likely be achieved, along with a corresponding reduction in side effects. Further reductions could be effected if such interaction could be rendered site-specific. Given that a functioning gene continually produces mRNA, it would thus be even more advantageous if gene transcription could be arrested 35 in its entirety.

Oligodeoxynucleotides offer such opportunities. For example, synthetic oligodeoxynucleotides could be used as

-3-

antisense probes to block and eventually lead to the breakdown of mRNA. Thus, synthetic DNA could suppress translation *in vivo*. It also may be possible to modulate the genome of an animal by, for example, triple helix formation using 5 oligonucleotides or other DNA recognizing agents. However, there are a number of drawbacks associated with triple helix formation. For example, it can only be used for homopurine sequences and it requires unphysiologically high ionic strength and low pH.

10 Furthermore, unmodified oligonucleotides are unpractical both in the antisense approach and in the triple helix approach because they have short *in vivo* half-lives, they are difficult to prepare in more than milligram quantities and, thus, are prohibitively costly, and they are 15 poor cell membrane penetrators.

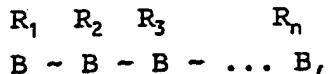
These problems have resulted in an extensive search for improvements and alternatives. For example, the problems arising in connection with double-stranded DNA (dsDNA) recognition through triple helix formation have been 20 diminished by a clever "switch back" chemical linking whereby a sequence of polypurine on one strand is recognized, and by "switching back", a homopurine sequence on the other strand can be recognized. See, e.g., McCurdy, Moulds, and Froehler, *Nucleosides*, in press. Also, good helix formation has been 25 obtained by using artificial bases, thereby improving binding conditions with regard to ionic strength and pH.

In order to improve half life as well as membrane penetration, a large number of variations in polynucleotide backbones has been undertaken, although so far not with 30 desired results. These variations include the use of methylphosphonates, monothiophosphates, dithiophosphates, phosphoramidates, phosphate esters, bridged phosphoroamidates, bridged phosphorothioates, bridged methylenephosphonates, dephospho internucleotide analogs with siloxane bridges, 35 carbonate bridges, carboxymethyl ester bridges, acetamide bridges, carbamate bridges, thio ther, sulfoxy, sulfono

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bridges, various "plastic" DNAs, α -anomeric bridges, and borane derivatives.

International patent application WO 86/05518 broadly claims a polymeric composition effective to bind to a single-stranded polynucleotide containing a target sequence of bases. The composition is said to comprise non-homopolymeric, substantially stereoregular polymer molecules of the form:

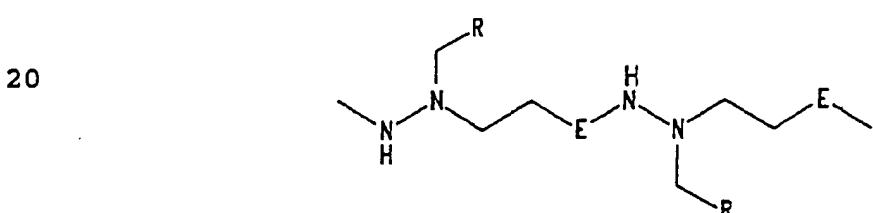
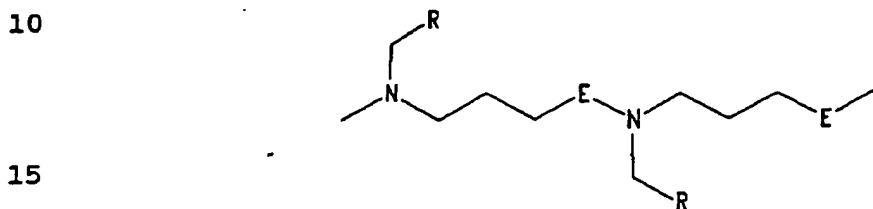
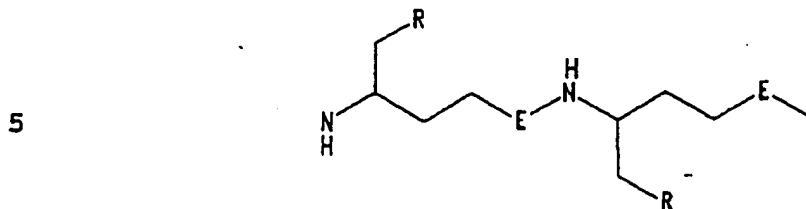


10 where:

- (a) R_1-R_n are recognition moieties selected from purine, purine-like, pyrimidine, and pyrimidine like heterocycles effective to bind by Watson/Crick pairing to corresponding, in-sequence bases in the target sequence;
- (b) n is such that the total number of Watson/Crick hydrogen bonds formed between a polymer molecule and target sequence is at least about 15;
- (c) $B - B$ are backbone moieties joined predominantly by chemically stable, substantially uncharged, predominantly achiral linkages;
- (d) the backbone moiety length ranges from 5 to 7 atoms if the backbone moieties have a cyclic structure, and ranges from 4 to 6 atoms if the backbone moieties have an acyclic structure; and
- (e) the backbone moieties support the recognition moieties at position which allow Watson/Crick base pairing between the recognition moieties and the corresponding, in-sequence bases of the target sequence.

According to WO 86/05518, the recognition moieties are various natural nucleobases and nucleobase-analogs and the backbone moieties are either cyclic backbone moieties comprising furan or morpholine rings or acyclic backbone moieties of the following forms:

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where E is -CO- or -SO₂-.

The specification of the application provides general descriptions for the synthesis of subunits, for backbone coupling reactions, and for polymer assembly strategies. However, the specification provides no example wherein a claimed compound or structure is actually prepared. Although WO 86/05518 indicates that the claimed polymer compositions can bind target sequences and, as a result, have possible diagnostic and therapeutic applications, the application contains no data relating to the binding affinity of a claimed polymer.

International patent application WO 86/05519 claims diagnostic reagents and systems that comprise polymers described in WO 86/05518, but attached to a solid support. WO 86/05519 also provides no examples concerning actually preparation of a claimed diagnostic reagent, much less data showing the diagnostic efficiency of such a reagent.

International patent application WO 89/12060 claims various building blocks for synthesizing oligonucleotide analogs, as well as oligonucleotide analogs formed by joining

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such building blocks in a defined sequence. The building blocks may be either "rigid" (containing a ring) or "flexible" (lacking a ring). In both cases the building blocks contain a hydroxy group and a mercapto group, through which the 5 building blocks are said to join to form oligonucleotide analogs. The linking moiety in the oligonucleotide analogs is selected from the group consisting of sulfide (-S-), sulfoxide (-SO-), and sulfone (-SO₂-). WO 89/12060 provides a general description concerning synthesis of the building 10 blocks and coupling reactions for the synthesis of oligonucleotide analogs, along with experimental examples describing the preparation of building blocks. However, the application provides no examples directed to the preparation of a claimed oligonucleotide analog and no data confirming the 15 specific binding of an oligonucleotide analog to a target oligonucleotide.

Furthermore, oligonucleotides or their derivatives have been linked to intercalators in order to improve binding, to polylysine or other basic groups in order to improve binding 20 both to double-stranded and single-stranded DNA, and to peptides in order to improve membrane penetration. However, such linking has not resulted in satisfactory binding for either double-stranded or single-stranded DNA. Other problems which resulted from, for example, methylphosphonates and 25 monothiophosphates were the occurrence of chirality, insufficient synthetic yield or difficulties in performing solid phase assisted syntheses.

In most cases only a few of these modifications could be used. Even then, only short sequences -- often only dimers 30 -- or monomers could be generated. Furthermore, the oligomers actually produced have rarely been shown to bind to DNA or RNA or have not been examined biologically.

The great majority of these backbone modifications led to decreased stability for hybrids formed between the modified 35 oligonucleotide and its complementary native oligonucleotide, as assayed by measuring T_m values. Consequently, it is generally understood in the art that backbone modifications

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d stabilize such hybrids, i.e., result in lower T_m values, and should be kept to a minimum.

OBJECTS OF THE INVENTION

5 It is one object of the present invention to provide compounds that bind ssDNA and RNA strands to form stable hybrids therewith.

It is a further object of the invention to provide compounds that bind ssDNA and RNA strands more strongly than the corresponding DNA.

It is another object to provide compounds wherein naturally-occurring nucleobases or other nucleobase-binding moieties are covalently bound to a peptide backbone.

It is yet another object to provide compounds other than RNA that can bind one strand of a double-stranded polynucleotide, thereby displacing the other strand.

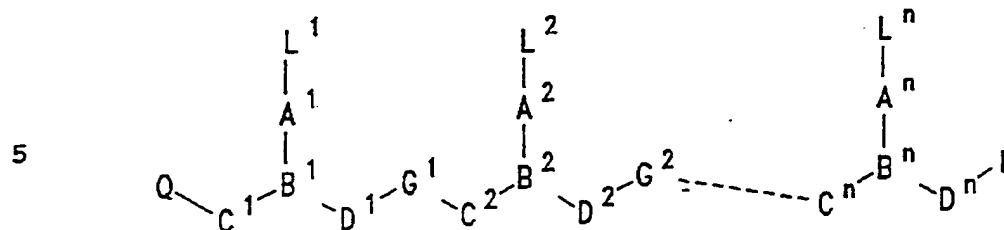
It is still another object to provide therapeutic and prophylactic methods that employ such compounds.

20 SUMMARY OF THE INVENTION

The present invention provides a novel class of compounds, known as peptide nucleic acids (PNAs), that bind complementary ssDNA and RNA strands more strongly than a corresponding DNA. The compounds of the invention generally comprise ligands linked to a peptide backbone via an aza nitrogen. Representative ligands include either the four main naturally occurring DNA bases (i.e., thymine, cytosine, adenine or guanine) or other naturally occurring nucleobases (e.g., inosine, uracil, 5-methylcytosine or thiouracil) or artificial bases (e.g., bromothymine, azaadenines or azaguuanines, etc.) attached to a peptide backbone through a suitable linker.

In certain preferred embodiments, the peptide nucleic acids of the invention have the general formula (I):

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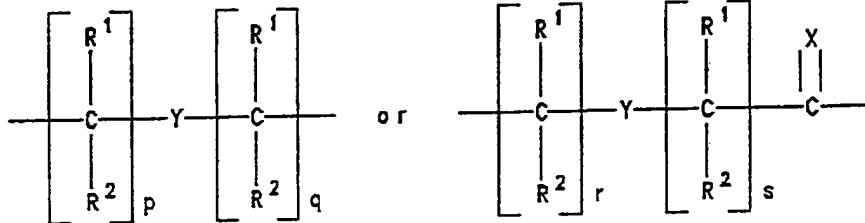
wherein:

n is at least 2,

each of L^1-L^n is independently selected from the group consisting of hydrogen, hydroxy, (C_1-C_4) alkanoyl, naturally occurring nucleobases, non-naturally occurring nucleobases, aromatic moieties, DNA intercalators, nucleobase-binding groups, heterocyclic moieties, and reporter ligands, at least one of L^1-L^n being a naturally occurring nucleobase, a non-naturally occurring nucleobase, a DNA intercalator, or a nucleobase-binding group;

each of A^1-A^n is a single bond, a methylene group or a group of formula (IIa) or (IIb):

25



30

(IIa)

(IIb)

where:

35

x is O, S, Se, NR^3 , CH_2 or $C(CH_3)_2$;y is a single bond, O, S or NR^4 ;

each of p and q is zero or an integer from 1 to 5, the sum p+q being not more than 10;

40

each of r and s is zero or an integer from 1 to 5, the sum r+s being not more than 10;

each R^1 and R^2 is independently selected from the group consisting of hydrogen, (C_1-C_4) alkyl which may be hydroxy- or alkoxy- or alkylthio-substituted, hydroxy, alkoxy, alkylthio, amino and halogen; and

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each R³ and R⁴ is independently selected from the group consisting of hydrogen, (C₁-C₄)alkyl, hydroxy- or alkoxy- or alkylthio-substituted (C₁-C₄)alkyl, hydroxy, alkoxy, alkylthio and amino;

5 each of B¹-Bⁿ is N or R³N⁺, where R³ is as defined above; each of C¹-Cⁿ is CR⁶R⁷, CHR⁶CHR⁷ or CR⁶R⁷CH₂, where R⁶ is hydrogen and R⁷ is selected from the group consisting of the side chains of naturally occurring alpha amino acids, or R⁶ and R⁷ are independently selected from the group consisting of

10 hydrogen, (C₂-C₆)alkyl, aryl, aralkyl, heteroaryl, hydroxy, (C₁-C₆)alkoxy, (C₁-C₆)alkylthio, NR³R⁴ and SR⁵, where R³ and R⁴ are as defined above, and R⁵ is hydrogen, (C₁-C₆)alkyl, hydroxy-, alkoxy-, or alkylthio- substituted (C₁-C₆)alkyl, or R⁶ and R⁷ taken together complete an alicyclic or heterocyclic

15 system;

each of D¹-Dⁿ is CR⁶R⁷, CH₂CR⁶R⁷ or CHR⁶CHR⁷, where R⁶ and R⁷ are as defined above;

each of G¹-Gⁿ⁻¹ is -NR³CO-, -NR³CS-, -NR³SO- or -NR³SO₂-, Y in either orientation, where R³ is as defined above;

20 Q is -CO₂H, -CONR'R'', -SO₃H or -SO₂NR'R'' or an activated derivative of -CO₂H or -SO₃H; and

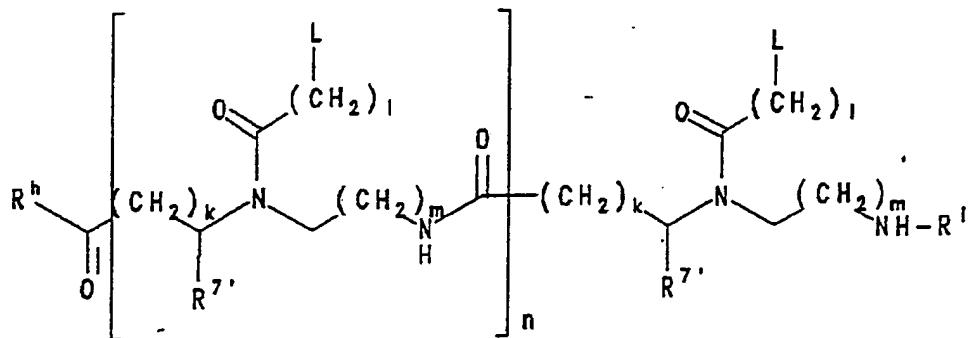
I is -NHR'''R'''' or -NR'''C(O)R''''', where R', R'', R''' and R'''' are independently selected from the group consisting of hydrogen, alkyl, amino protecting groups,

25 reporter ligands, intercalators, chelators, peptides, proteins, carbohydrates, lipids, steroids, oligonucleotides and soluble and non-soluble polymers.

The peptide nucleic acids of the invention differ from those disclosed in WO 86/05518 in that their recognition moieties are attached to an aza nitrogen atom in the backbone, rather than to an amide nitrogen atom, a hydrazine moiety or a carbon atom in the backbone.

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Preferred peptide nucleic acids have general formula (III):



(III)

wherein:

5 each L is independently selected from the group consisting of hydrogen, phenyl, heterocyclic moieties, naturally occurring nucleobases, and non-naturally occurring nucleobases;

10 each R' is independently selected from the group consisting of hydrogen and the side chains of naturally occurring alpha amino acids;

n is an integer from 1 to 60;

each of k, l and m is independently zero or an integer from 1 to 5;

15 R^h is OH, NH₂ or -NHLysNH₂; and

Rⁱ is H or COCH₃.

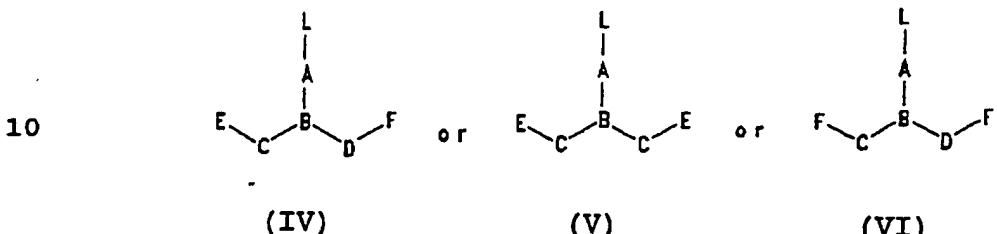
Particularly preferred are compounds having formula (III) wherein each L is independently selected from the group consisting of the nucleobases thymine (T), adenine (A), 20 cytosine (C), guanine (G) and uracil (U), k and m are zero or 1, and n is an integer from 1 to 30, in particular from 4 to 20. An example of such a compound is provided in Figure 1, which shows the structural similarity between such compounds and single-stranded DNA.

25 The peptide nucleic acids of the invention are synthesized by adaptation of standard peptide synthesis procedures, either in solution or on a solid phase. The synthons used are specially designed monomer amino acids or their activated derivatives, protected by standard protecting

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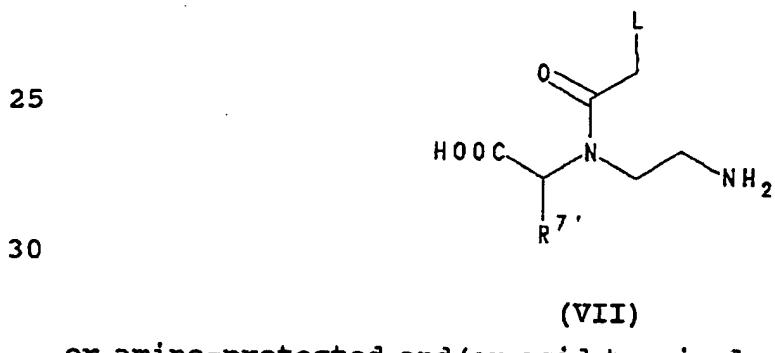
groups. The oligonucleotide analogs also can be synthesized by using the corresponding diacids and diamines.

Thus, the novel monomer synthons according to the invention are selected from the group consisting of amino acids, diacids and diamines having general formulae:



15 wherein L, A, B, C and D are as defined above, except that any amino groups therein may be protected by amino protecting groups; E is COOH, CSOH, SOOH, SO₂OH or an activated derivative thereof; and F is NHR³ or NPgR³, where R³ is as defined above and Pg is an amino protecting group.

20 Preferred monomer synthons according to the invention
are amino acids having formula (VII):



or amino-protected and/or acid terminal activated derivatives thereof, wherein L is selected from the group consisting of
35 hydrogen, phenyl, heterocyclic moieties, naturally occurring nucleobases, non-naturally occurring nucleobases, and protected derivatives thereof; and R⁷ is independently selected from the group consisting of hydrogen and the side chains of naturally occurring alpha amino acids. Especially
40 preferred are such synthons having formula (VII) wherein R⁷ is hydrogen and L is selected from the group consisting of the nucleobases thymine (T), adenine (A), cytosine (C), guanine (G) and uracil (U) and protected derivatives thereof.

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Unexpectedly, these compounds also are able to recognize duplex DNA by displacing one strand, thereby presumably generating a double helix with the other one. Such recognition can take place to dsDNA sequences 5-60 base pairs long. Sequences between 10 and 20 bases are of interest since this is the range within which unique DNA sequences of prokaryotes and eukaryotes are found. Reagents which recognize 17-18 bases are of particular interest since this is the length of unique sequences in the human genome. The compounds of the invention also should be able to form triple helices with dsDNA.

Whereas the improved binding of the compounds of the invention should render them efficient as antisense agents, it is expected that an extended range of related reagents may cause strand displacement, now that this surprising and unexpected new behavior of dsDNA has been discovered.

Thus, in one aspect, the present invention provides methods for inhibiting the expression of particular genes in the cells of an organism, comprising administering to said organism a reagent as defined above which binds specifically to sequences of said genes.

Further, the invention provides methods for inhibiting transcription and/or replication of particular genes or for inducing degradation of particular regions of double stranded DNA in cells of an organism by administering to said organism a reagent as defined above.

Still further, the invention provides methods for killing cells or virus by contacting said cells or virus with a reagent as defined above which binds specifically to sequences of the genome of said cells or virus.

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BRIEF DESCRIPTION OF THE DRAWINGS

The numerous objects and advantages of the present invention may be better understood by those skilled in the art by reference to the accompanying figures, in which:

- 5 Figure 1 shows a naturally occurring deoxyribooligonucleotide (A) and a peptide nucleic acid (PNA) of the invention (B).

Figure 2 provides examples of naturally occurring and non-naturally occurring nucleobases for DNA recognition and
10 reporter groups.

Figure 3 provides a schematic illustration of (a) photocleavage by Acr¹-(Taeg)₁₀-Lys-NH₂ (Acr-T10-LysNH₂); (b) photofootprint by the diazo-linked acridine of Acr¹-(Taeg)₁₀-Lys-NH₂ and preferred KMnO₄-cleavage; and (c) S₁-nuclease
15 enhanced cleavage and (d) micrococcus nuclease cleavage of Acr¹-(Taeg)₁₀-Lys-NH₂ binding site.

Figure 4 provides examples of PNA monomer synthons of the invention.

Figure 5 shows the Acr¹ ligand and a PNA, Acr¹-(Taeg)₁₀-
20 Lys-NH₂.

Figure 6 provides a general scheme for the preparation of monomer synthons.

Figure 7 provides a general scheme for the preparation of the Acr¹ ligand.

25 Figure 8 provides a general scheme for solid-phase PNA synthesis illustrating the preparation of linear unprotected PNA amides.

Figure 9 shows analytical HPLC chromatograms of: (A) crude H-[Taeg]₁₅-NH₂ after HF-cleavage (before lyophilization);
30 (B) crude Acr¹-[Taeg]₁₅-NH₂ after HF-cleavage (before lyophilization); and (C) purified Acr¹-[Taeg]₁₅-NH₂. Buffer A, 5% CH₃CN/95% H₂O/0.0445% TFA; buffer B, 60% CH₃CN/40% H₂O/0.0390% TFA; linear gradient, 0-100% of B in 30 min; flow rate, 1.2 ml/min; column, Vydac C₁₈ (5 μm, 0.46 x 25 cm).

35 Figure 10 shows analytical HPLC chromatograms of: (A) purified H-[Taeg]₁₀-Lys-NH₂ and (B) purified H-[Taeg]₅-Caeg-[Taeg]₄-Lys-NH₂ employing the same conditions as in Figure 9.

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Figures 11a and 11b show binding of AcrT10-Lys to dA₁₀. 5'-³²P-labeled oligonucleotide (1) (5'-GATCCA₁₀G) was incubated in the absence or presence of Acr-T10-LysNH₂ and in the absence or presence of oligonucleotide (2) (5'-GATCCT₁₀G) and 5 the samples were analyzed by polyacrylamide gel electrophoresis (PAGE) and autoradiography under "native conditions" (Figure 11a) or under "denaturing conditions" (Figure 11b).

Figures 12a-c show chemical, photochemical and 10 enzymatic probing of dsDNA-Acr-T10-LysNH₂ complex. Complexes between Acr-T10-LysNH₂ and a ³²P-endlabeled DNA fragment containing a dA₁₀/dT₁₀ target sequence were probed by affinity photocleavage (Figure 12a, lanes 1-3; Figure 12b, lanes 1-3), 15 photofootprinting (Figure 12a, lanes 5-6), potassium permanganate probing (Figure 12b, lanes 4-6) or probing by staphylococcus nuclease (Figure 12b, lanes 8-10) or by nuclease S₁ (Figure 12c). Either the A-strand (Figure 12a) or 20 the T-strand (Figures 12b,c) was probed.

Figure 13 provides a procedure for the synthesis of 20 protected PNA synthons.

Figure 14 provides a procedure for the synthesis of a protected adenine monomer synthon.

Figure 15 provides a procedure for the synthesis of a protected guanine monomer synthon.

25 Figure 16 provides examples of PNA backbone alterations.

Figure 17 provides a procedure for synthesis of thymine monomer synthons with side chains corresponding to the normal amino acids.

30 Figures 18a and 18b provide procedures for synthesis of an aminopropyl analogue and a propionyl analogue, respectively, of a thymine monomer synthon.

Figure 19 provides a procedure for synthesis of an aminoethyl-β-alanine analogue of thymine monomer synthon.

35 Figure 20 shows a PAGE autoradiograph demonstrating that PNAS-T₁₀, -T₉C and -T₈C₂ bind to double stranded DNA with high sequence specificity.

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Figure 21 shows a graph based on densitometric scanning of PAGE autoradiographs demonstrating the kinetics of the binding of PNA-T₁₀ to a double stranded target.

Figure 22 shows a graph based on densitometric scanning 5 of PAGE autoradiographs demonstrating the thermal stabilities of PNAs of varying lengths bound to an A₁₀/T₁₀ double stranded DNA target.

Figure 23 shows an electrophoretic gel staining demonstrating that restriction enzyme activity towards DNA is 10 inhibited when PNA is bound proximal to the restriction enzyme recognition site.

Figure 24 shows a PAGE autoradiograph demonstrating that ¹²⁵I-labeled PNA-T₁₀ binds to a complementary dA₁₀ oligonucleotide.

15 Figure 25 shows a peptide nucleic acid according to the invention.

Figure 26 shows the direction of synthesis for a peptide nucleic acid according to the invention.

Figure 27 provides a test for the tosyl group as a 20 nitrogen protecting group in the synthesis of peptide nucleic acids.

DETAILED DESCRIPTION OF THE INVENTION

In the oligonucleotide analogs and monomer synthons 25 according to the invention, ligand L is primarily a naturally occurring nucleobase attached at the position found in nature, i.e., position 9 for adenine or guanine, and position 1 for thymine or cytosine. Alternatively, L may be a non-naturally occurring nucleobase (nucleobase analog), another base-binding 30 moiety, an aromatic moiety, (C₁-C₄)alkanoyl, hydroxy or even hydrogen. Some typical nucleobase ligands and illustrative synthetic ligands are shown in Figure 2. Furthermore, L can be a DNA intercalator, a reporter ligand such as, for example, a fluorophor, radio label, spin label, hapten, or a protein-35 recognizing ligand such as biotin.

In monomer synthons, L may be blocked with protecting groups. This is illustrated in Figure 4, where Pg¹ is an

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acid, a base or a hydrogenolytically or photochemically cleavable protecting group such as, for example, t-butoxycarbonyl (Boc), fluorenylmethyloxycarbonyl (Fmoc) or 2-nitrobenzyl (2Nb).

Linker A can be a wide variety of groups such as -CR¹R²CO-, -CR¹R²CS-, -CR¹R²CSe-, -CR¹R²CNHR³-, -CR¹R²C=CH₂- and -CR¹R²C=C(CH₃)₂-, where R¹, R² and R³ are as defined above. Preferably, A is methylenecarbonyl (-CH₂CO-). Also, A can be a longer chain moiety such as propanoyl, butanoyl or pentanoyl, or corresponding derivative, wherein O is replaced by another value of X or the chain is substituted with R¹R² or is heterogenous, containing Y. Further, A can be a (C₂-C₆)alkylene chain, a (C₂-C₆)alkylene chain substituted with R¹R² or can be heterogenous, containing Y. In certain cases, A can just be a single bond.

In the preferred form of the invention, B is a nitrogen atom, thereby presenting the possibility of an achiral backbone. B can also be R³N⁺, where R³ is as defined above.

In the preferred form of the invention, C is -CR⁶R⁷-, but can also be a two carbon unit, i.e. -CHR⁶CHR⁷- or -CR⁶R⁷CH₂-, where R⁶ and R⁷ are as defined above. R⁶ and R⁷ also can be a heteroaryl group such as, for example, pyrrolyl, furyl, thieryl, imidazolyl, pyridyl, pyrimidinyl, indolyl, or can be taken together to complete an alicyclic system such as, for example, 1,2-cyclobutenediyl, 1,2-cyclopentanediyI or 1,2-cyclohexanediyI.

In the preferred form of the invention, E in the monomer synthon is COOH or an activated derivative thereof, and G in the oligomer is -CONR³- . As defined above, E may also be CSOH, SOOH, SO₂OH or an activated derivative thereof, whereby G in the oligomer becomes -CSNR³-, -SONR³-and -SO₂NR³-, respectively. The activation may, for example, be achieved using an acid anhydride or an active ester derivative, wherein hydrogen in the groups represented by E is replaced by a leaving group suited for generating the growing backbone.

The amino acids which form the backbone may be identical or different. We have found that those based on 2-

-17-

aminoethylglycine are especially well suited to the purpose of the invention.

In some cases it may be of interest to attach ligands at either terminus (Q, I) to modulate the binding characteristics of the PNAs. Representative ligands include DNA intercalators which will improve dsDNA binding or, basic groups, such as lysine or polylysine, which will strengthen the binding of PNA due to electrostatic interaction. To decrease negatively charged groups such as carboxy and sulfo groups could be used. The design of the synthons further allows such other moieties to be located on non-terminal positions.

In a further aspect of the invention, the PNA oligomers are conjugated to low molecular effector ligands such as ligands having nuclease activity or alkylating activity or reporter ligands (fluorescent, spin labels, radioactive, protein recognition ligands, for example, biotin or haptens). In a further aspect of the invention, the PNAs are conjugated to peptides or proteins, where the peptides have signaling activity and the proteins are, for example, enzymes, transcription factors or antibodies. Also, the PNAs can be attached to water-soluble or water-insoluble polymers. In another aspect of the invention, the PNAs are conjugated to oligonucleotides or carbohydrates. When warranted, a PNA oligomer can be synthesized onto some moiety (e.g., a peptide chain, reporter, intercalator or other type of ligand-containing group) attached to a solid support.

Such conjugates can be used for gene modulation (e.g., gene targeted drugs), for diagnostics, for biotechnology, and for scientific purposes.

As a further aspect of the invention, PNAs can be used to target RNA and ssDNA to produce both antisense-type gene regulating moieties and hybridization probes for the identification and purification of nucleic acids. Furthermore, the PNAs can be modified in such a way that they can form triple helices with dsDNA. Reagents that bind sequence-specifically to dsDNA have applications as gene

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targeted drugs. These are foreseen as extremely useful drugs for treating diseases like cancer, AIDS and other virus infections, and may also prove effective for treatment of some genetic diseases. Furthermore, these reagents may be used for 5 research and in diagnostics for detection and isolation of specific nucleic acids.

The triple helix principle is believed to be the only known principle in the art for sequence-specific recognition of dsDNA. However, triple helix formation is largely limited 10 to recognition of homopurine-homopyrimidine sequences. Strand displacement is superior to triple helix recognition in that it allows for recognition of any sequence by use of the four natural bases. Also, in strand displacement recognition readily occurs at physiological conditions, that is, neutral 15 pH, ambient (20-40 C) temperature and medium (100-150 mM) ionic strength.

Gene targeted drugs are designed with a nucleobase sequence (containing 10-20 units) complementary to the regulatory region (the promoter) of the target gene. 20 Therefore, upon administration of the drug, it binds to the promoter and block access thereto by RNA polymerase. Consequently, no mRNA, and thus no gene product (protein), is produced. If the target is within a vital gene for a virus, no viable virus particles will be produced. Alternatively, 25 the target could be downstream from the promoter, causing the RNA polymerase to terminate at this position, thus forming a truncated mRNA/protein which is nonfunctional.

Sequence-specific recognition of ssDNA by base complementary hybridization can likewise be exploited to target 30 specific genes and viruses. In this case, the target sequence is contained in the mRNA such that binding of the drug to the target hinders the action of ribosomes and, consequently, translation of the mRNA into protein. The peptide nucleic acids of the invention are superior to prior reagents in that 35 they have significantly higher affinity for complementary ssDNA. Also, they possess no charge and water soluble, which should facilitate cellular uptake, and they contain amides of

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non-biological amino acids, which should make them biostable and resistant to enzymatic degradation by, for example, proteases.

Certain biochemical/biological properties of PNA 5 oligomers are illustrated by the following experiments.

1. Sequence discrimination at the dsDNA level (Example 63, Figure 20).

Using the S₁-nuclease probing technique, the discrimination of binding of the T₁₀, T₅CT₄(T₉C) & T₂CT₂CT₄(T₈C₂) PNA 10 to the recognition sequences A₁₀, A₅GA₄ (A₉G) & A₂GA₂GA₄ (A₈G₂) cloned into the BamHI, SalI or PstI site of the plasmid pUC19 was analyzed. The results (Figure 20) show that the three PNAs bind to their respective recognition sequences with the following relative efficiencies: PNA - T₁₀: A₁₀ > A₉G >> A₈G₂, 15 PNA - T₉C: A₉G > A₁₀ ~ A₈G₂, PNA - T₈C₂: A₈G₂ ≥ A₉G >> A₁₀. Thus at 37°C one mismatch out of ten gives reduced efficiency (5-10 times estimated) whereas two mismatches are not accepted.

2. Kinetics of PNA-T₁₀ - dsDNA strand displacement complex formation (Example 66, Figure 21).

20 Complex formation was probed by S₁-nuclease at various times following mixing of PNA and ³²P-endlabeled dsDNA fragment (Figure 21).

3. Stability of PNA-dsDNA complex (Example 67, Figure 22)

Complexes between PNA-T_n and ³²P-dsDNA (A₁₀/T₁₀) target 25 were formed (60 min, 37°C). The complexes were then incubated at the desired temperature in the presence of excess oligo-dA₁₀ for 10 min, cooled to RT and probed with KMnO₄. The results (Figure 22) show that the thermal stability of the PNA-dsDNA complexes mirror that of the PNA oligonucleotide 30 complexes in terms of "T_m".

4. Inhibition of restriction enzyme cleavage by PNA (Example 65, Figure 23)

The plasmid construct, pT10, contains a dA₁₀/dT₁₀ tract cloned into the BamHI site in pUC19. Thus, cleavage of pT10 35 with BamHI and PvuII results in two small DNA fragments of 211 and 111 bp, respectively. In the presence of PNA-T₁₀, a 336 bp fragment is obtained corresponding to cleavage only by

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PvuII (Figure 23). Thus cleavage by BamHI is inhibited by PNA bound proximal to the restriction enzyme site. The results also show that the PNA-dsDNA complex can be formed in 100% yield. Similar results were obtained using the pT8C2 plasmid 5 and PNA-T8C2.

5. Binding of 125 I-labeled PNA to oligonucleotides (Example 63, Figure 24)

A Tyr-PNA-T₁₀-Lys-NH₂ was labeled with 125 I using Na 125 I and chloramine-T and purified by HPLC. The 125 I-PNA-T₁₀ was 10 shown to bind to oligo-dA₁₀ by PAGE and autoradiography (Figure 24). The binding could be competed by excess denatured calf thymus DNA.

The sequence-specific recognition of dsDNA is illustrated by the binding of a PNA, consisting of 10 thymine substituted 2-aminoethylglycyl units, which C-terminates in a lysine amide and N-terminates in a complex 9-aminoacridine ligand (9-Acr¹-(Taeg)₁₀-Lys-NH₂, Figure 11a, 11b) to a dA₁₀/dT₁₀ target sequence. The target is contained in a 248 bp 32 P-end-labelled DNA-fragment.

20 Strand displacement was ascertained by the following type of experiments:

1) The 9-Acr¹ ligand (Figure 5), which is equipped with a 4-nitrobenzamido group to ensures cleavage of DNA upon irradiation, is expected only to cleave DNA in close proximity 25 to its binding site. Upon irradiation of the PNA with the above 248 bp DNA fragment, selective cleavage at the dA₁₀/dT₁₀ sequence is observed (Figure 3a).

2) In a so-called photofootprinting assay, where a synthetic diazo-linked acridine under irradiation cleaves DNA 30 (except where the DNA is protected by said binding substance) upon interaction with DNA in the presence of a DNA-binding substance.

Such an experiment was performed with the above 248 bp dsDNA fragment, which showed clear protection against photocleavage of the PNA binding site (Figure 3b).

3) In a similar type of experiment, the DNA-cleaving enzyme micrococcus nuclease, which is also hindered in its

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action by most DNA-binding reagents, showed increased cleavage at the T₁₀-target (Figure 3c).

4) In yet another type of experiment, the well-known high susceptibility of single strand thymine ligands (as opposed to double strand thymine ligands) towards potassium permanganate oxidation was employed. Oxidation of the 248 bp in the presence of the reagent showed only oxidation of the T₁₀-strand of the target (Figure 3b).

5) In a similar type of demonstration, the single strand specificity of S₁ nuclease clearly showed that only the T₁₀-strand of the target was attacked (Figure 3d).

The very efficient binding of (Taeg)₁₀, (Taeg)₁₀-Lys-NH₂ and Acr¹-(Taeg)₁₀-Lys-NH₂ (Figures 11a. 11b) to the corresponding dA₁₀ was furthermore illustrated in two ways:

15 1. Ligand-oligonucleotide complexes will migrate slower than the naked oligonucleotide upon electrophoresis in polyacrylamide gels. Consequently, such experiments were performed with Acr¹-(Taeg)₁₀-Lys-NH₂ and ³²P-end-labelled dA₁₀. This showed retarded migration under conditions where a normal 20 dA₁₀/dT₁₀ duplex is stable, as well as under conditions where such a duplex is unstable (denaturing gel). A control experiment was performed with a mixture of Acr¹-(Taeg)₁₀-Lys-NH₂ and ³²P-end-labelled dT₁₀ which showed no retardation under the above conditions.

25 2. Upon formation of DNA duplexes (dsDNA) from single strand DNA, the extinction coefficient decreases (hypochromicity). Thus, the denaturing of DNA can be followed by measuring changes in the absorbance, for example, as a function of T_m, the temperature where 50% of a duplex has 30 disappeared to give single strands.

Duplexes were formed from the single-stranded oligodeoxyribonucleotides and the PNAs listed below. Typically 0.3 OD₂₆₀ of the T-rich strand was hybridized with 1 equivalent of the other strand by heating to 90 C for 5 min, 35 cooling to room temperature and kept for 30 min and finally stored in a refrigerator at 5 C for at least 30 min. The buffers used were all 10 mM in phosphate and 1 mM in EDTA.

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The low salt buffer contained no sodium chloride, whereas the medium salt buffer contained 140 mM NaCl and the high salt buffer 500 mM NaCl. The pH of all the buffers was 7.2. The melting temperature of the hybrids were determined on a 5 Gilford Response apparatus. The following extinction coefficients were used A: 15.4 ml/ μ mol·cm; T: 8.8; G: 11.7 and C: 7.3 for both normal oligonucleotides and PNA. The melting curves were recorded in steps of 0.5 °C/min. The T_m were determined from the maximum of the 1st derivative of the plot 10 of A_{260} vs temperature.

List of oligodeoxyribonucleotides:

1. 5'-AAA-AAA-AA
2. 5'-AAA-AAA-AAA-A
- 15 3. 5'-TTT-TTT-TTT-T
4. 5'-AAA-AAG-AAA-A
5. 5'-AAG-AAG-AAA-A
6. 5'-AAA-AGA-AAA-A
7. 5'-AAA-AGA-AGA-A
- 20 8. 5'-TTT-TCT-TTT-T
9. 5'-TTT-TCT-TCT-T
10. 5'-TTT-TTC-TTT-T
11. 5'-TTT-TTC-TTC-T
12. 5'-TTC-TTC-TTT-T
- 25 13. 5'-TTT-TTT-TTT-TTT-TTT
14. 5'-AAA-AAA-AAA-AAA-AAA

List of PNAs

- a. TTT-TTT-TTT-T-Lys-NH₂
- 30 b. TTT-TTT-TT-Lys-NH₂
- c. TTT-TTC-TTT-T-Lys-NH₂
- d. TTC-TTC-TTT-T-Lys-NH₂
- e. Acr-TTT-TTT-TTT-T-Lys-NH₂
- f. Ac-TTT-TTT-TTT-T-Lys-NH₂

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Oligo/PNA	Low Salt	Medium Salt	High Salt
1+b	56.0	51.5	50.0
2+a	73.0	72.5	73.0
2+c		41.5 and 52.0*	
5	2+e	84.5	~90
	2+f		74
	4+a	60.0	61.5
	4+c	74.5	72.5
	4+f		62.0
10	5+a		47.0
	5+c		57.5
	5+f		46.5
	7+a		46.0
	7+c		58.0
15	7+f		43.5
	7+12		23.0
	13+14		39.0

* = Two distinct melting temperatures are seen,
indicating local melting before complete
denaturation.

The hybrid formed between RNA-A (poly rA) and PNA-T₁₀-Lys-NH₂ melts at such high temperature that it cannot be measured (>90 C). But specific hybridization is demonstrated
25 by the large drop in A₂₆₀ by mixing with RNA-A but not G,C and U. The experiment is done by mixing 1 ml of a solution of the PNA and 1 ml of a solution the RNA, each with A₂₆₀ = 0.6, and then measure the absorbance at 260 nm. Thereafter the sample is heated to 90 C for 5 min, cooled to room temperature and
30 kept at this temperature for 30 minutes and finally stored at 5C for 30 min.

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RNA	PNA	A ₂₆₀ Before Mixing	A ₂₆₀ After Mixing	A ₂₆₀ After Mixing and Heating
RNA-A	PNA-T ₁₀ -lys-NH ₂	0.600	0.389	0.360
RNA-U	PNA-T ₁₀ -lys-NH ₂	0.600	0.538	0.528
RNA-G	PNA-T ₁₀ -lys-NH ₂	0.600	0.514	0.517
5 RNA-C	PNA-T ₁₀ -lys-NH ₂	0.600	0.540	0.532

- From the above measurements the following conclusions can be made. There is base stacking, since a melting curve is observed. The PNA-DNA hybrid is more stable than a normal DNA-DNA hybrid, and the PNA-RNA is even more stable. Mismatches cause significant drops in the T_m-value, whether the mispaired base is in the DNA or in the PNA-strand. The T_m-value is only slightly dependent on ionic strength, as opposed to normal oligonucleotides.
- 15 The synthesis of the PNAs according to the invention is discussed in detail in the following, where Figure 1 illustrates one of the preferred PNA examples and compares its structure to that of a complementary DNA.

20 Synthesis of PNA Oligomers and Polymers

The principle of anchoring molecules onto a solid matrix, which helps in accounting for intermediate products during chemical transformations, is known as Solid-Phase Synthesis or Merrifield Synthesis (see, e.g., Merrifield, J. Am. Chem. Soc., 1963, 85, 2149 and Science, 1986, 232, 341). Established methods for the stepwise or fragmentwise solid-phase assembly of amino acids into peptides normally employ a beaded matrix of slightly cross-linked styrene-divinylbenzene copolymer, the cross-linked copolymer having been formed by the pearl polymerization of styrene monomer to which has been added a mixture of divinylbenzenes. A level of 1-2% cross-linking is usually employed. Such a matrix also can be used in solid-phase PNA synthesis in accordance with the present invention (Figure 8).

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Concerning the initial functionalization of the solid phase, more than fifty methods have been described in connection with traditional solid-phase peptide synthesis (see, e.g., Barany and Merrifield in "The Peptides" Vol. 2, 5 Academic Press, New York, 1979, pp. 1-284, and Stewart and Young, "Solid Phase Peptide Synthesis", 2nd Ed., Pierce Chemical Company, Illinois, 1984). Reactions for the introduction of chloromethyl functionality (Merrifield resin; via a chloromethyl methyl ether/SnCl₄ reaction), aminomethyl 10 functionality (via an N-hydroxymethylphthalimide reaction; see, Mitchell, et al., Tetrahedron Lett., 1976, 3795), and benzhydryl amino functionality (Pietta, et al., J. Chem. Soc., 1970, 650) are the most widely applied. Regardless of its nature, the purpose of the functionality is normally to form 15 an anchoring linkage between the copolymer solid support and the C-terminus of the first amino acid to be coupled to the solid support. As will be recognized, anchoring linkages also can be formed between the solid support and the amino acid N-terminus. It is generally convenient to express the 20 "concentration" of a functional group in terms of millimoles per gram (mmol/g). Other reactive functionalities which have been initially introduced include 4-methylbenzhydryl amino and 4-methoxybenzhydryl amino. All of these established methods are in principle useful within the context of the present invention. Preferred methods for PNA synthesis employ aminomethyl as the initial functionality, in that aminomethyl is particularly advantageous with respect to the incorporation 25 of "spacer" or "handle" groups, owing to the reactivity of the amino group of the aminomethyl functionality with respect to the essentially quantitative formation of amide bonds to a carboxylic acid group at one end of the spacer-forming reagent. A vast number of relevant spacer- or handle-forming bifunctional reagents have been described (see, Barany, et al., Int. J. Peptide Protein Res., 1987, 30, 705), especially 30 35 reagents which are reactive towards amino groups such as found in the aminomethyl function. Representative bifunctional reagents include 4-(haloalkyl)aryl-lower alkanoic acids such

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as 4-(bromomethyl)phenylacetic acid, Boc-aminoacyl-4-(oxymethyl)aryl-lower alkanoic acids such as Boc-aminoacyl-4-(oxymethyl)phenylacetic acid, N-Boc-p-acylbenzhydrylamines such as N-Boc-p-glutaroylbenzhydrylamine, N-Boc-4'-lower alkyl-p-acylbenzhydrylamines such as N-Boc-4'-methyl-p-glutaroylbenzhydrylamine, N-Boc-4'-lower alkoxy-p-acylbenzhydrylamines such as N-Boc-4'-methoxy-p-glutaroyl-benzhydrylamine, and 4-hydroxymethylphenoxyacetic acid. One type of spacer group particularly relevant within the context of the present invention is the phenylacetamidomethyl (Pam) handle (Mitchell and Merrifield, *J. Org. Chem.*, 1976, 41, 2015) which, deriving from the electron withdrawing effect of the 4-phenylacetamidomethyl group, is about 100 times more stable than the classical benzyl ester linkage towards the Boc-amino deprotection reagent trifluoroacetic acid (TFA).

Certain functionalities (e.g., benzhydrylamo, 4-methylbenzhydrylamo and 4-methoxybenzhydrylamo) which may be incorporated for the purpose of cleavage of a synthesized PNA chain from the solid support such that the C-terminal of the PNA chain is in amide form, require no introduction of a spacer group. Any such functionality may advantageously be employed in the context of the present invention.

An alternative strategy concerning the introduction of spacer or handle groups is the so-called "preformed handle" strategy (see, Tam, et al., *Synthesis*, 1979, 955-957), which offers complete control over coupling of the first amino acid, and excludes the possibility of complications arising from the presence of undesired functional groups not related to the peptide or PNA synthesis. In this strategy, spacer or handle groups, of the same type as described above, are reacted with the first amino acid desired to be bound to the solid support, the amino acid being N-protected and optionally protected at the other side-chains which are not relevant with respect to the growth of the desired PNA chain. Thus, in those cases in which a spacer or handle group is desirable, the first amino acid to be coupled to the solid support can either be coupled to the free reactive end of a spacer group which has been

